

# Network Inference and Properties

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SciLifeLab

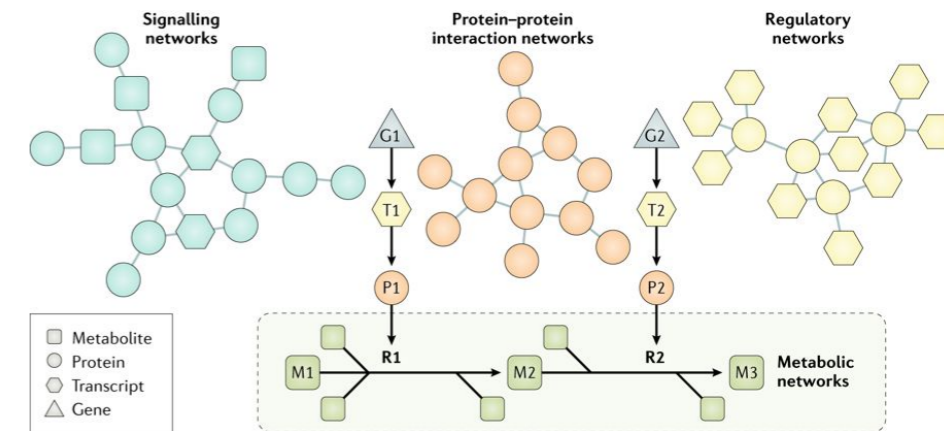
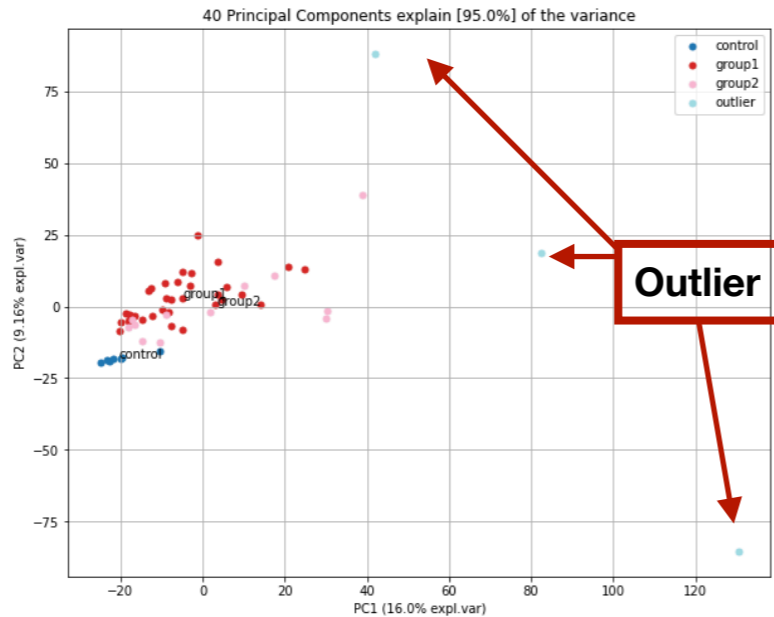


# Overview

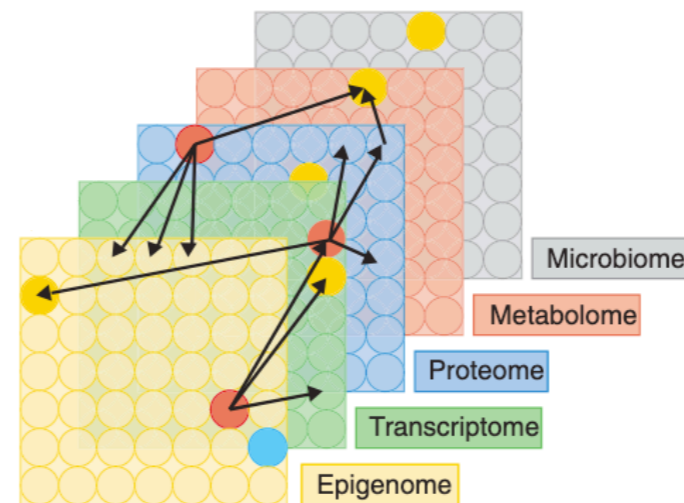
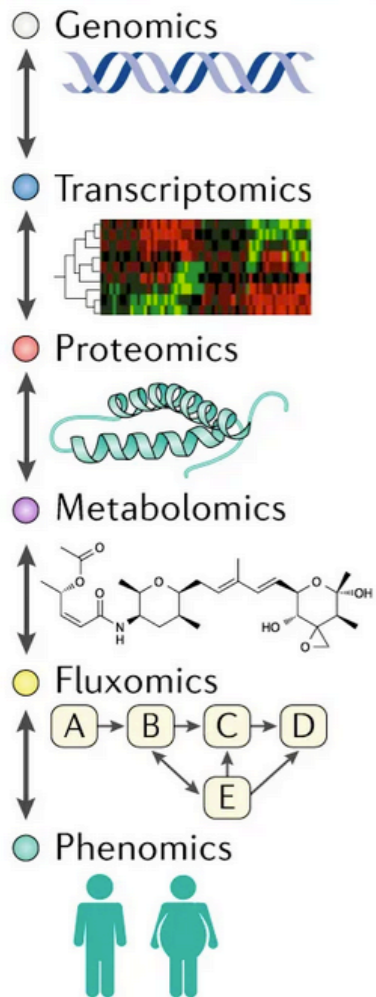
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1. Introduction to network analysis
2. Terminology
- 3. Network inference**
4. Key network properties
5. Community analysis

# Building networks



Raw → Pre-processing → Distance calculation → Graph analysis



Hasin 2017  
 Piening 2018  
 Mardinoglu 2018

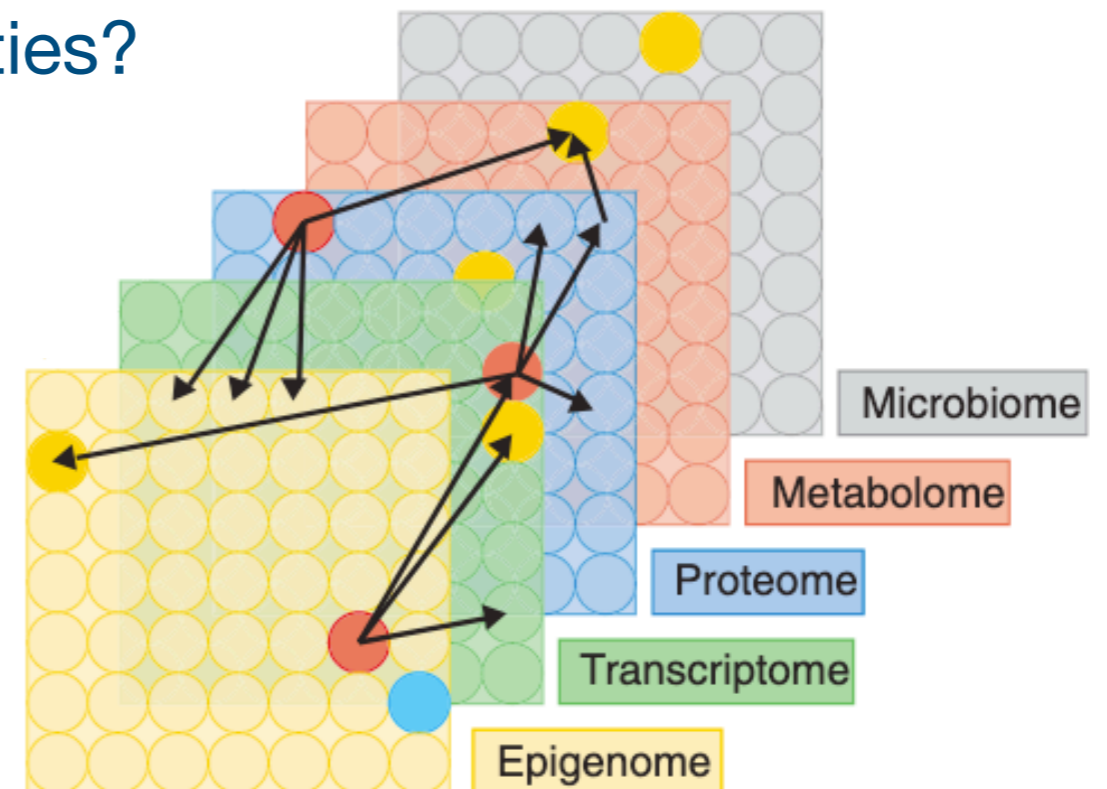
# Interomic vs Intraomic networks

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Networks may be build for individual omics or for their integration

What is my biological question?

- Do I want to analyse vertical relationships between features?
- Biological motivation for integrating omics with different coverage (e.g. transcriptomic and proteomic)
- Do I want to extract functional properties?



# Different approaches for network inference

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- |   |                                   |
|---|-----------------------------------|
| 1. Feature association                            | <b>No prior graph structure</b>   |
| 2. K-nearest neighbour graph (k-NNG) construction |                                   |
| 3. Knowledge-based                                | <b>Based on prior information</b> |
| 4. Genome-scale metabolic models                  |                                   |

# 1. Association analysis

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Balanced dataset for group sizes

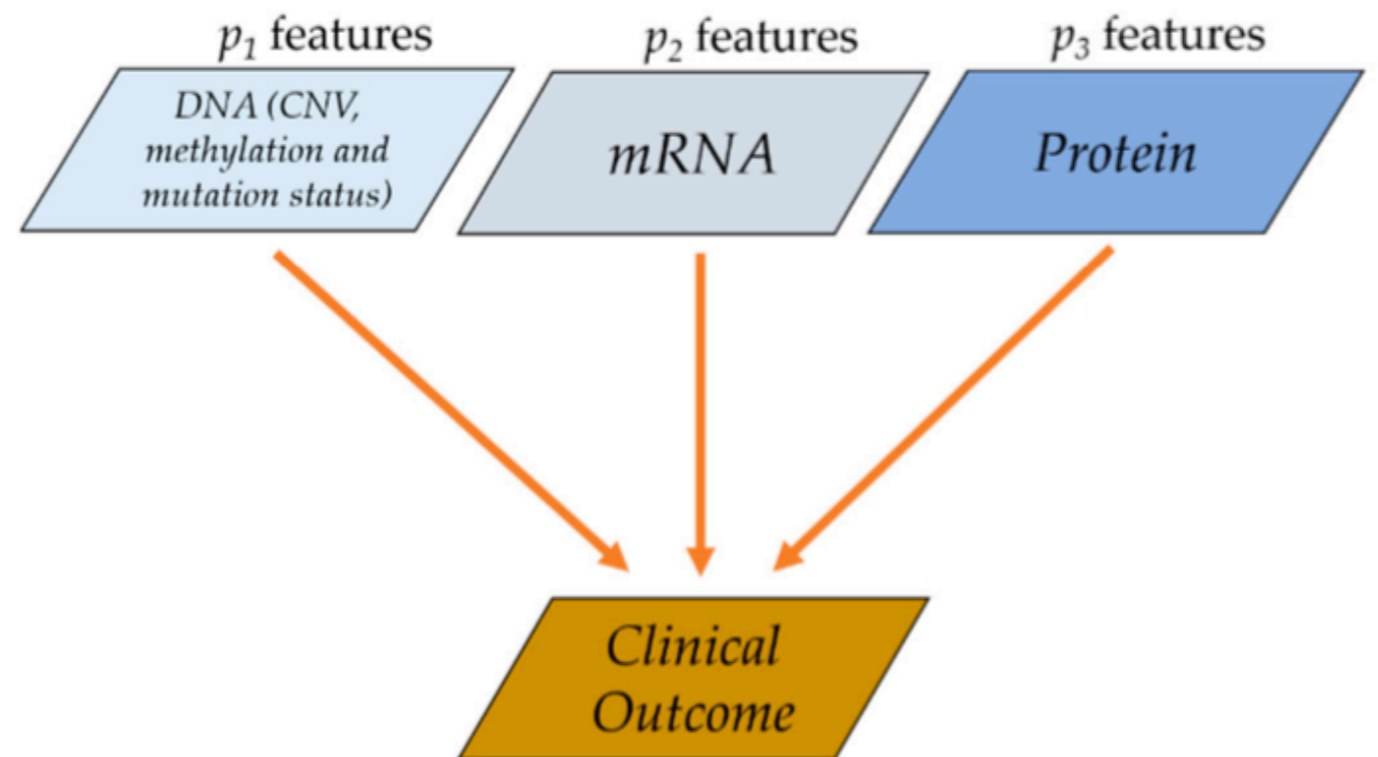
GroupA (80 samples) vs GroupB (20 samples)

GroupA (50 samples) vs GroupB (50 samples)

Common approach: compute correlations between different features

- Spearman
- Pearson

Extend known associations



# 1. Association analysis

Easy to interpret

Unweighted vs weighted ( $-1 \leq \rho \leq 1$ )

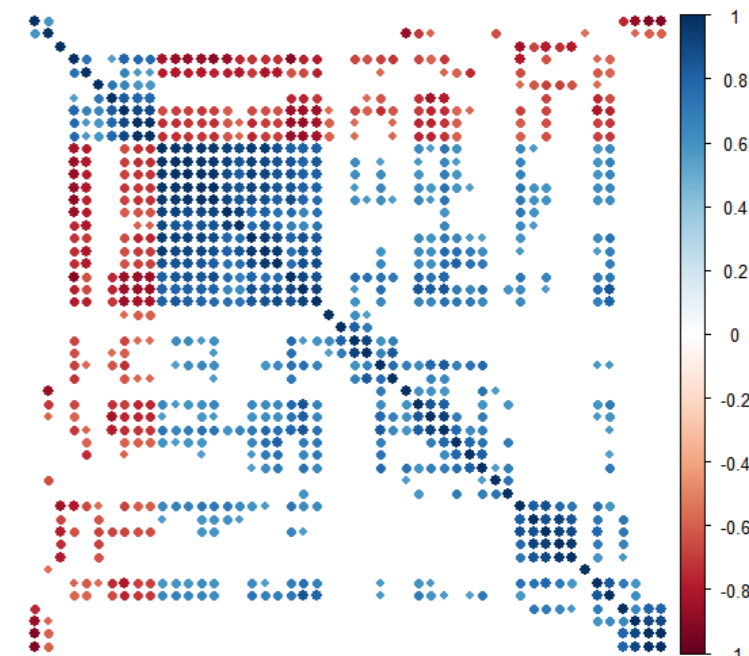
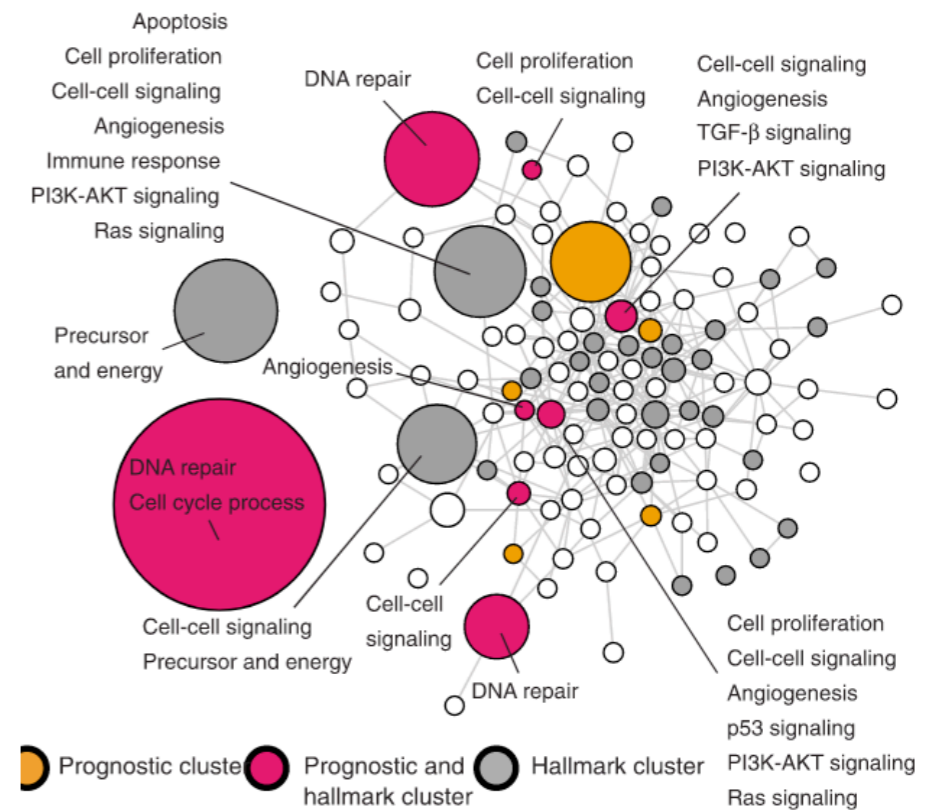
Unbalanced networks

Prone to type I errors

Filtering

- FDR vs Bonferroni
- Correlation coefficient cutoff

Need adjustment to possible confounding factors

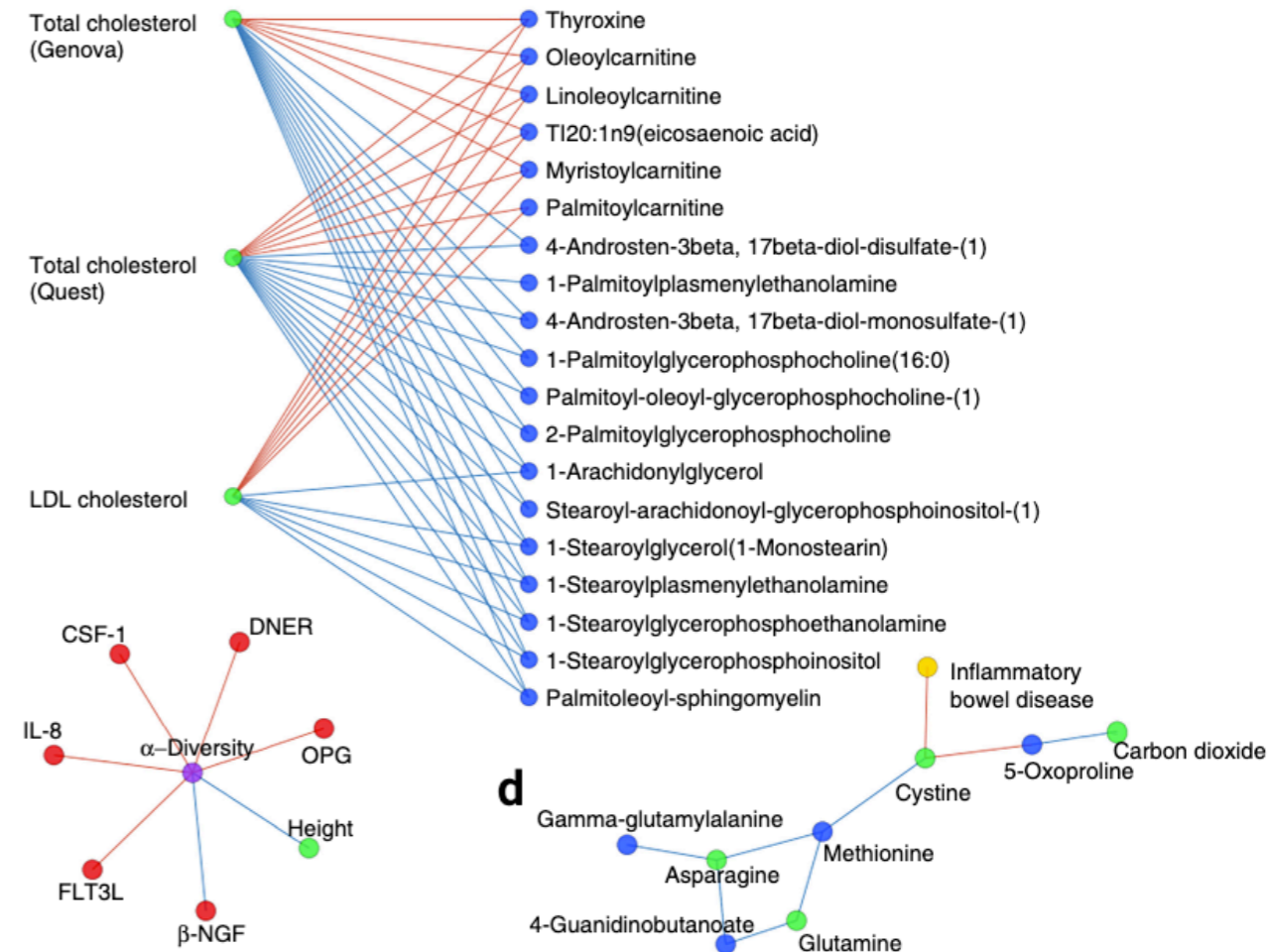
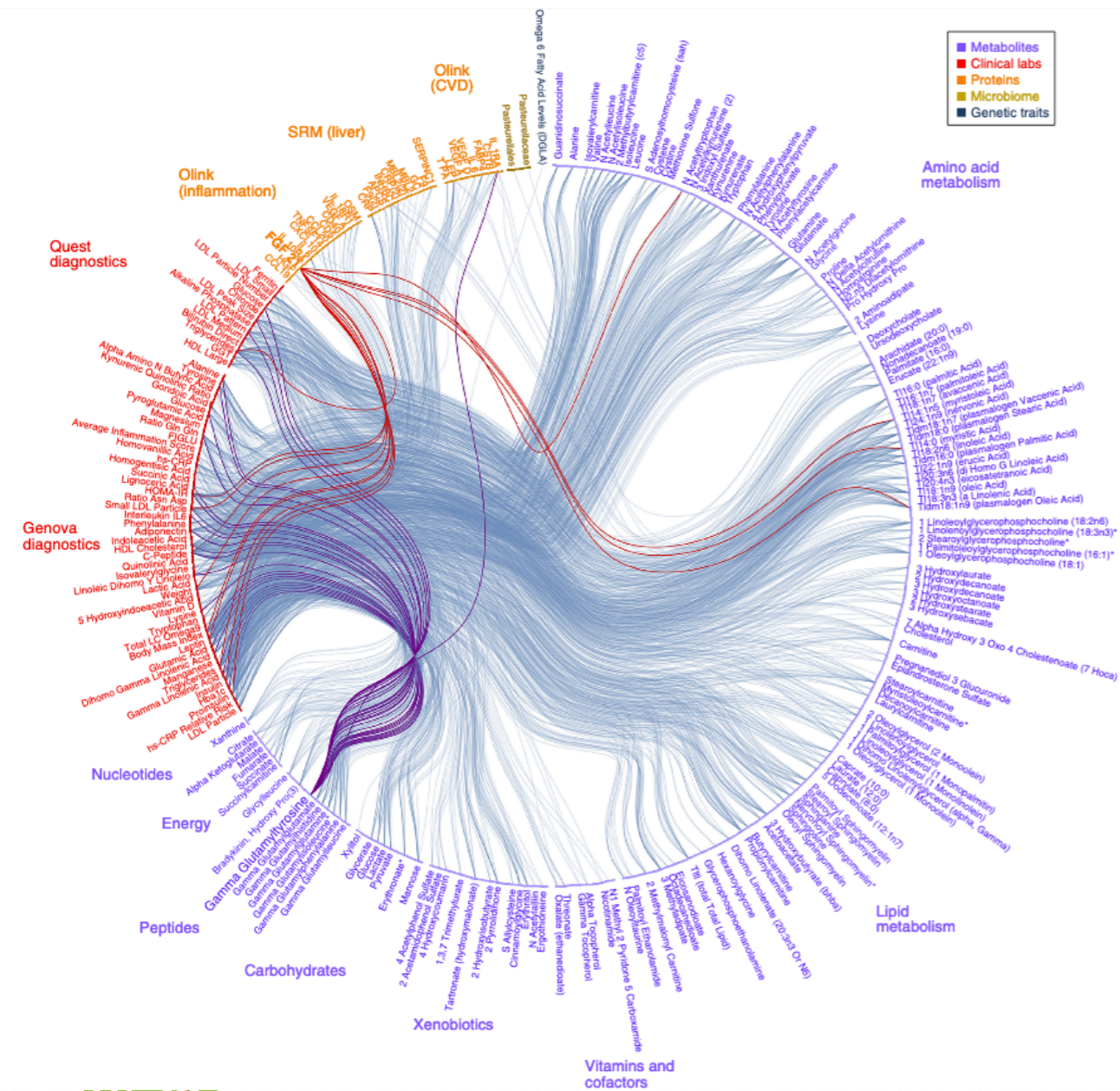


# 1. Association analysis

Adjusting for confounding factors: partial correlation analysis

Below:

- gender and age are known confounding factors
- feature regression on confounding factors, followed by correlation on the residuals of each model



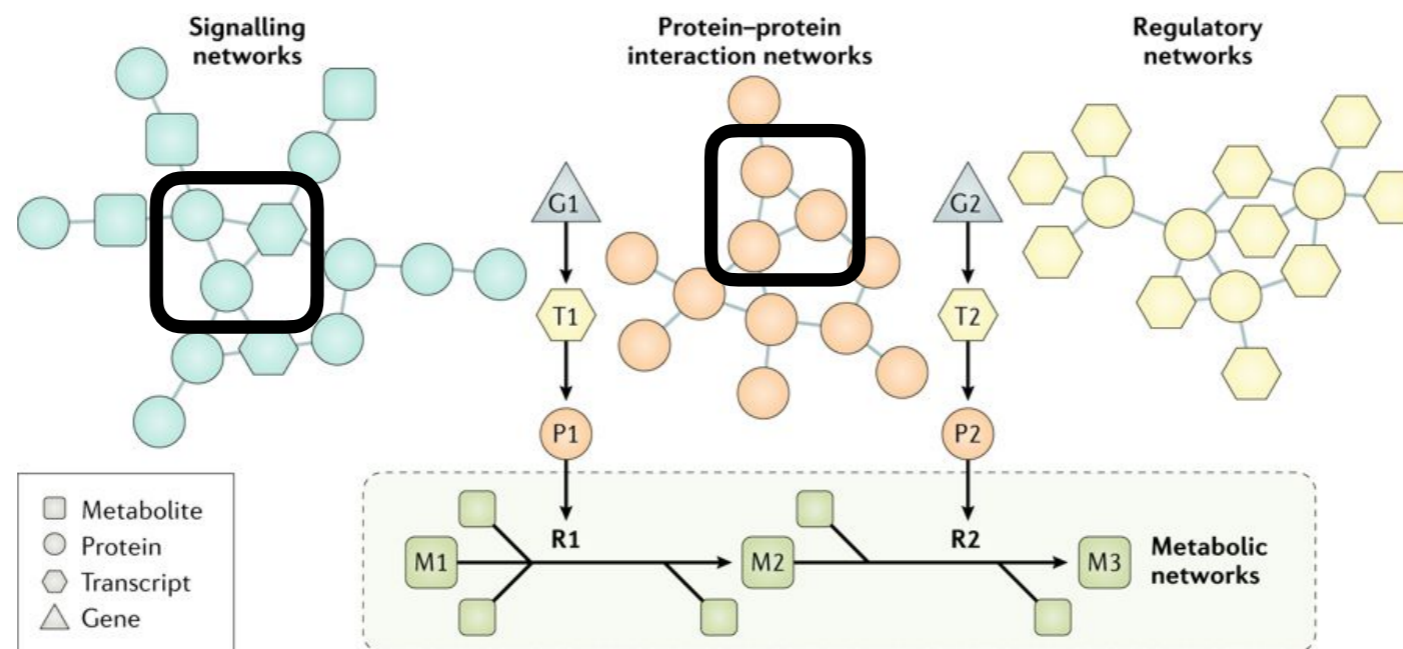


# Computationally expensive representations may be simplified

Does your graph have many cliques? **Possibly noisy**

**Graph contraction** simplifies the graph by successively grouping cliques

**Problem:** reduces information and prevents studying many properties of the graph



Clarke 2011  
Krzywinski 2013  
Sham 2014  
Nygaard 2016  
Piening 2018  
other refs as links

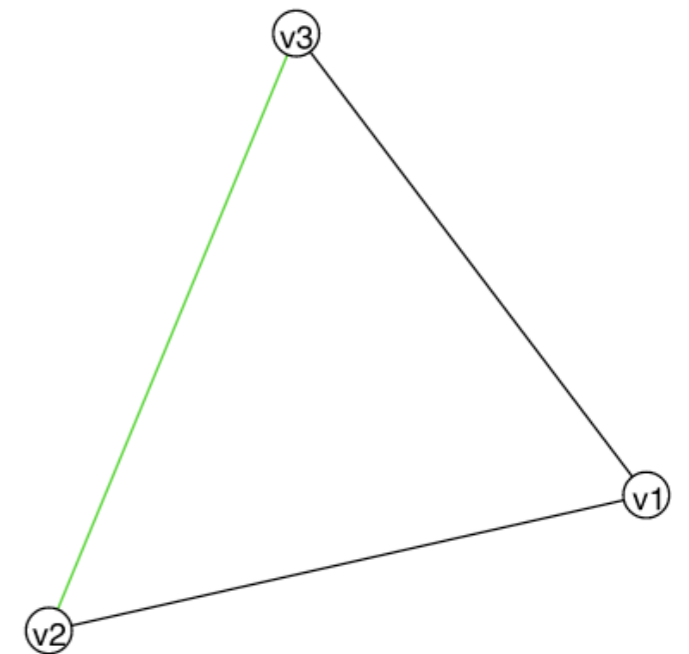
## 2. $k$ -nearest neighbour graph

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1. For each pair of features  $(u, v)$ , compute a distance metric:

- Correlation
- Euclidean
- Jaccard
- ...

2. For each feature, select the *closest*  $k$  neighbours



Efficiency (not scalable, compute all neighbours for every node)

Generates well-structured graph

Simple as it reduces the number of features

Loses potentially important information because  $k$  is fixed

# 2. $k$ -nearest neighbour graph

High  $k$  is smooth, but biased (underfitting)

Low  $k$  is accurate, but noisy (overfitting)

Optimum  $k$  may be identified:

- cross validation
- ad-hoc

**A. query of a gene on each network**

Liver integrated network

\* "target to source" option used

Liver co-expression network

Liver cancer co-expression network

**B. multiple gene search ( $\leq$  three genes)**

\* FASN and G6PD were searched in liver coexpression network

**C. expression of neighbors**

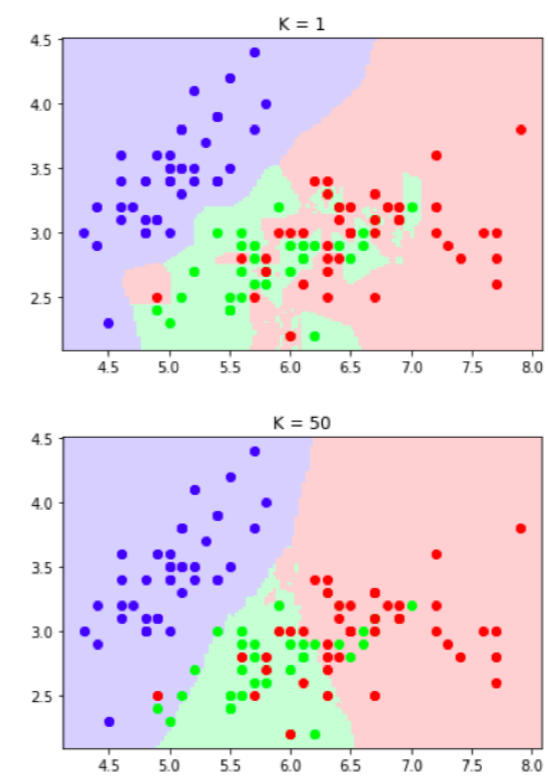
#	Ensembl ID	Symbol	Mean Expr	Min Expr	Max Expr
1	ENSG00000143627	PKLR	56.20 (n= 43.90)	0.00	495.00
2	ENSG00000134824	PDZD2	23.21 (n= 25.90)	0.00	240.00
3	ENSG00000113161	HMGCR	11.93 (n= 8.32)	1.00	52.00
4	ENSG00000160205	LSS	15.84 (n= 11.70)	2.00	91.00
5	ENSG00000131069	ACSS2	19.71 (n= 17.80)	2.00	151.00
6	ENSG00000135100	HNF1A	3.69 (n= 1.05)	0.00	12.00

**D. tissue expressions in HPA**

\* PKLR tissue expressions are shown.

**E. co-expression of edges**

#	M Source Node	M Target Node	Score	PValue
1	FASN	ELNVL6	0.590055	4.930E-06
2	FASN	TMEM135	0.519170	3.939E-06
3	FASN	TH8SP	0.483330	5.760E-03
4	FASN	FNPLA3	0.480324	1.839E-02
5	FASN	SOD	0.477626	3.370E-02



# 3. Knowledge-based graph creation

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## Database-derived

- PPI
- TFRN
- Metabolic Atlas
- ...

## Many reference databases

KEGG

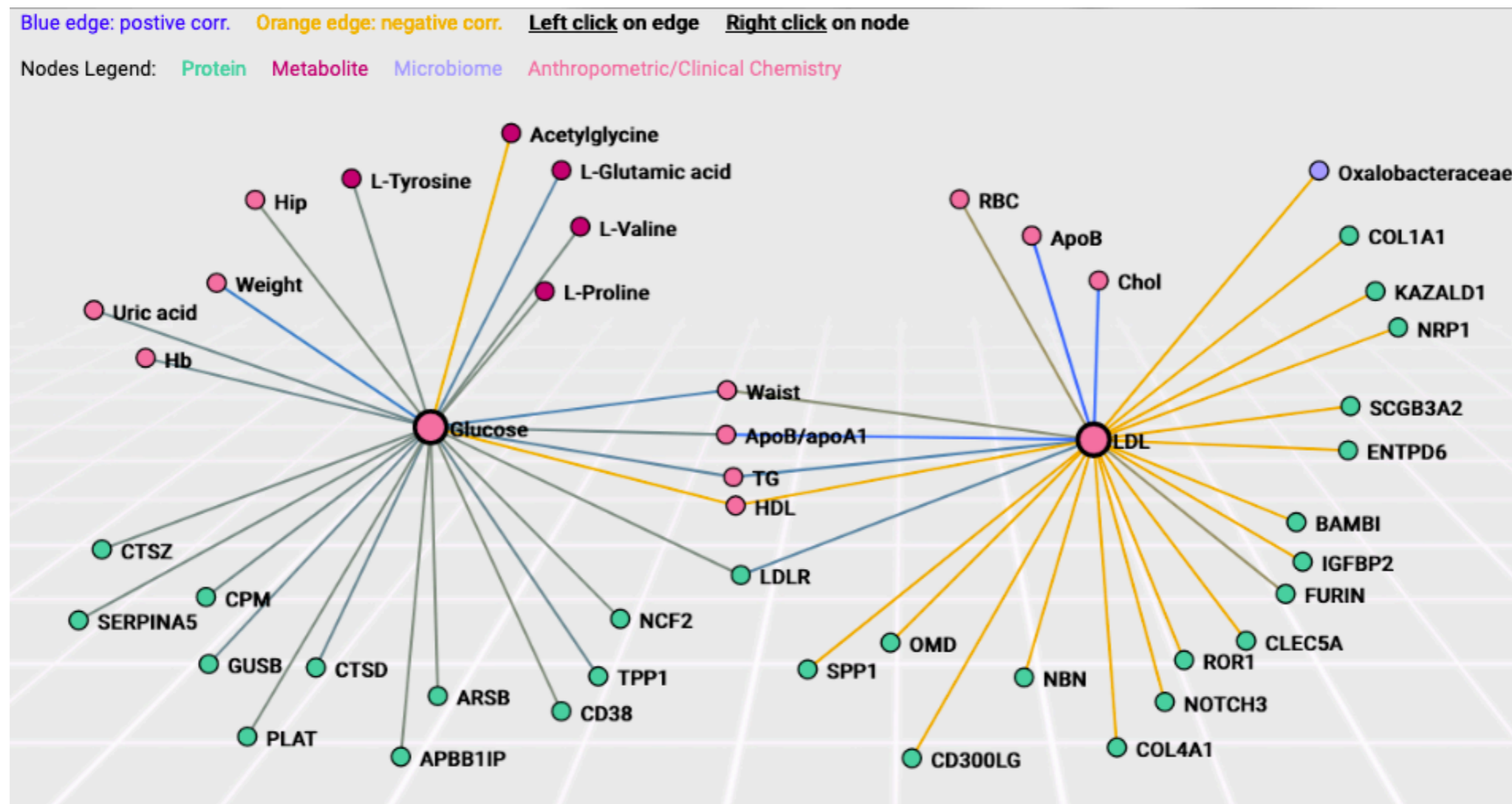
Reactome

WikiPathways

STRING-DB

# 3. Knowledge-based graph creation

## Multi-omic biological networks



# 3. Knowledge-based graph creation

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# 3. Knowledge-based graph creation

Little overlap among reference pathways

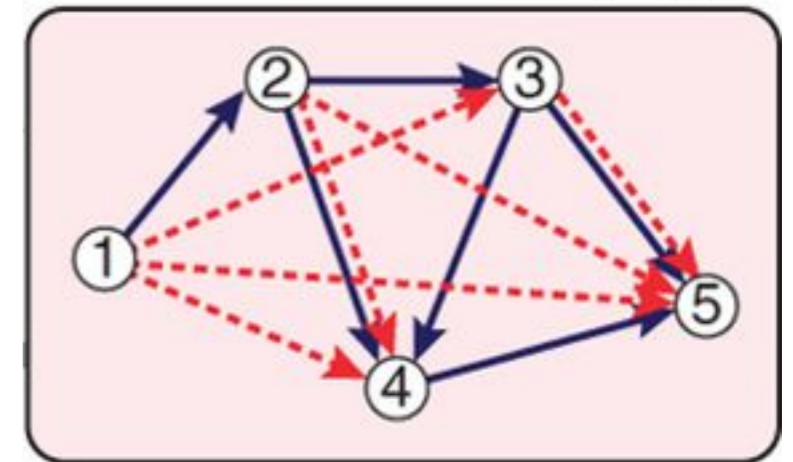
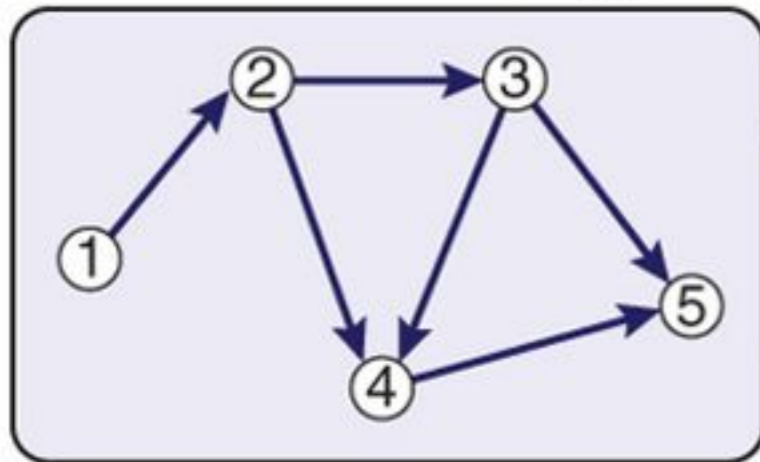


# 3. Knowledge-based graph creation

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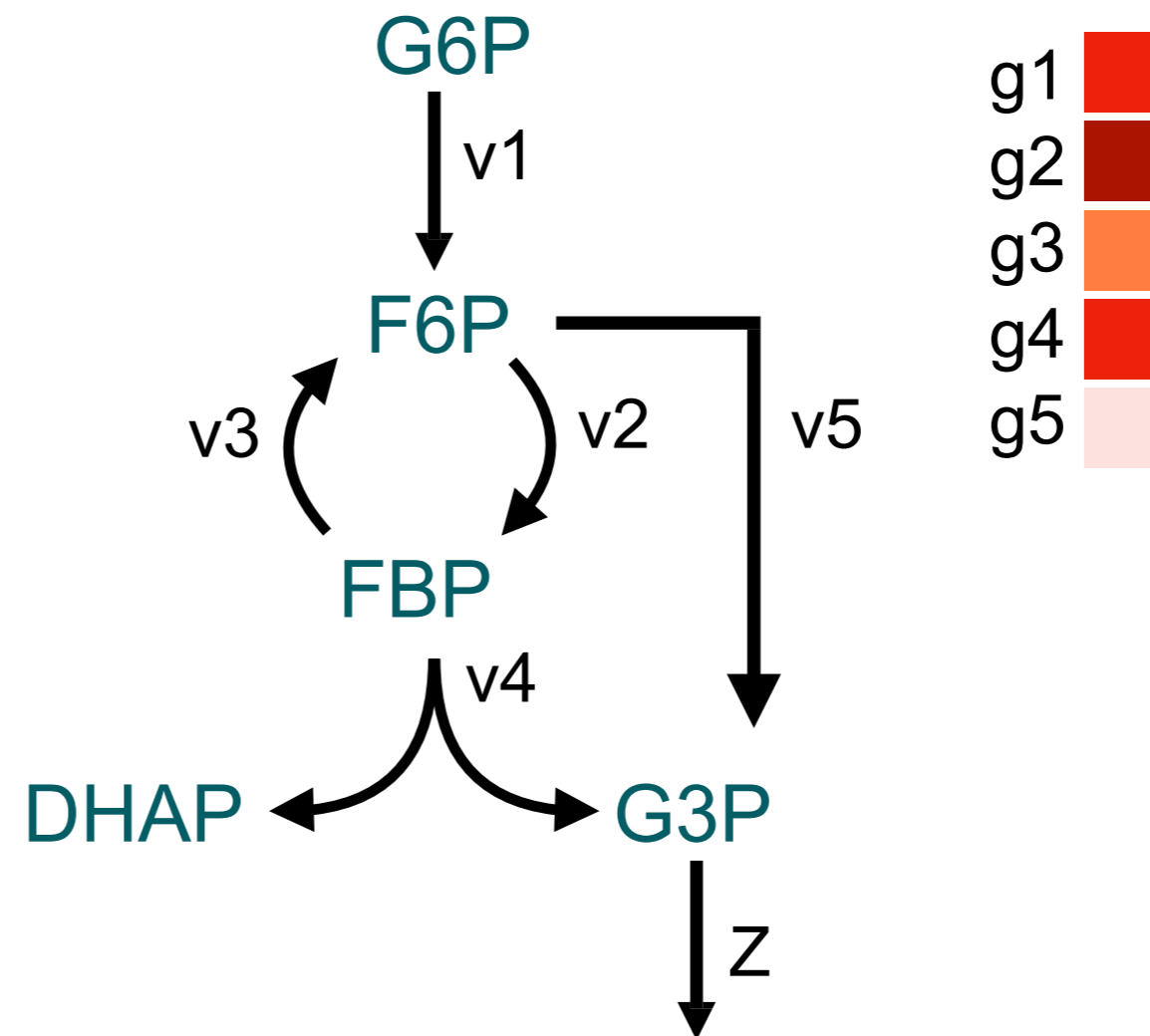
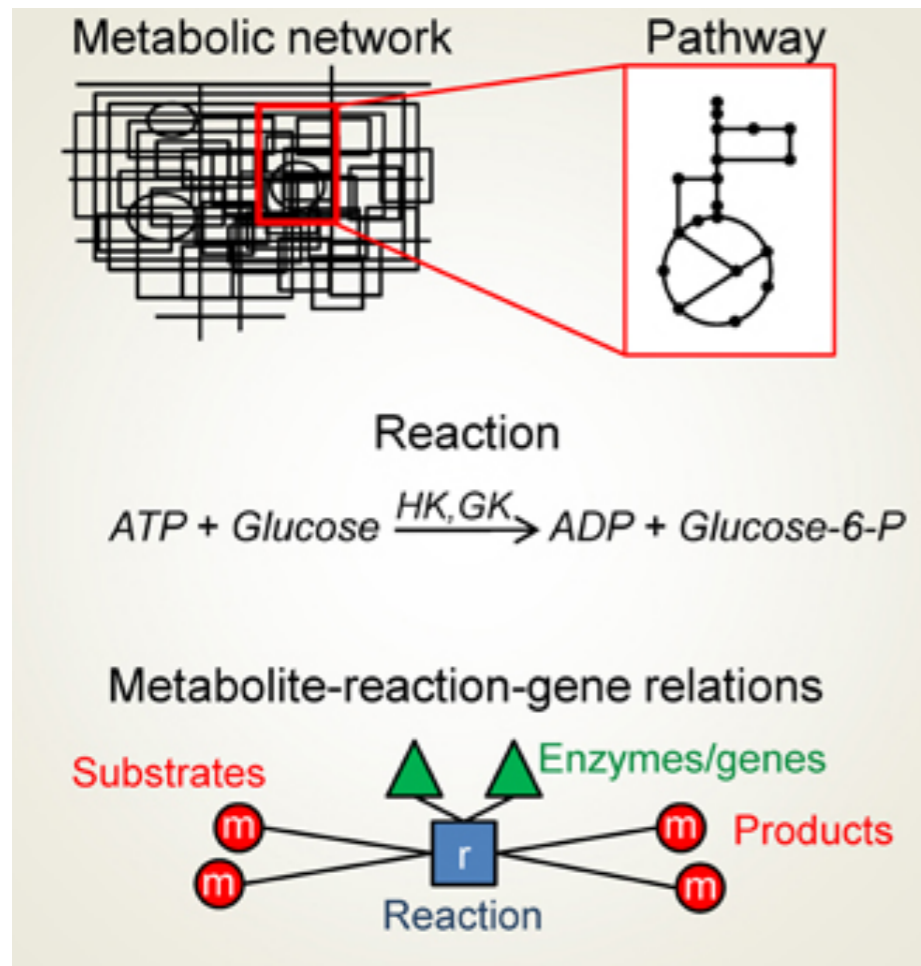
How to overlay your data based on known interactions?

- Filter your predicted interactions based on known information? (intersection)
- Add interactions that are not found in the reference networks?

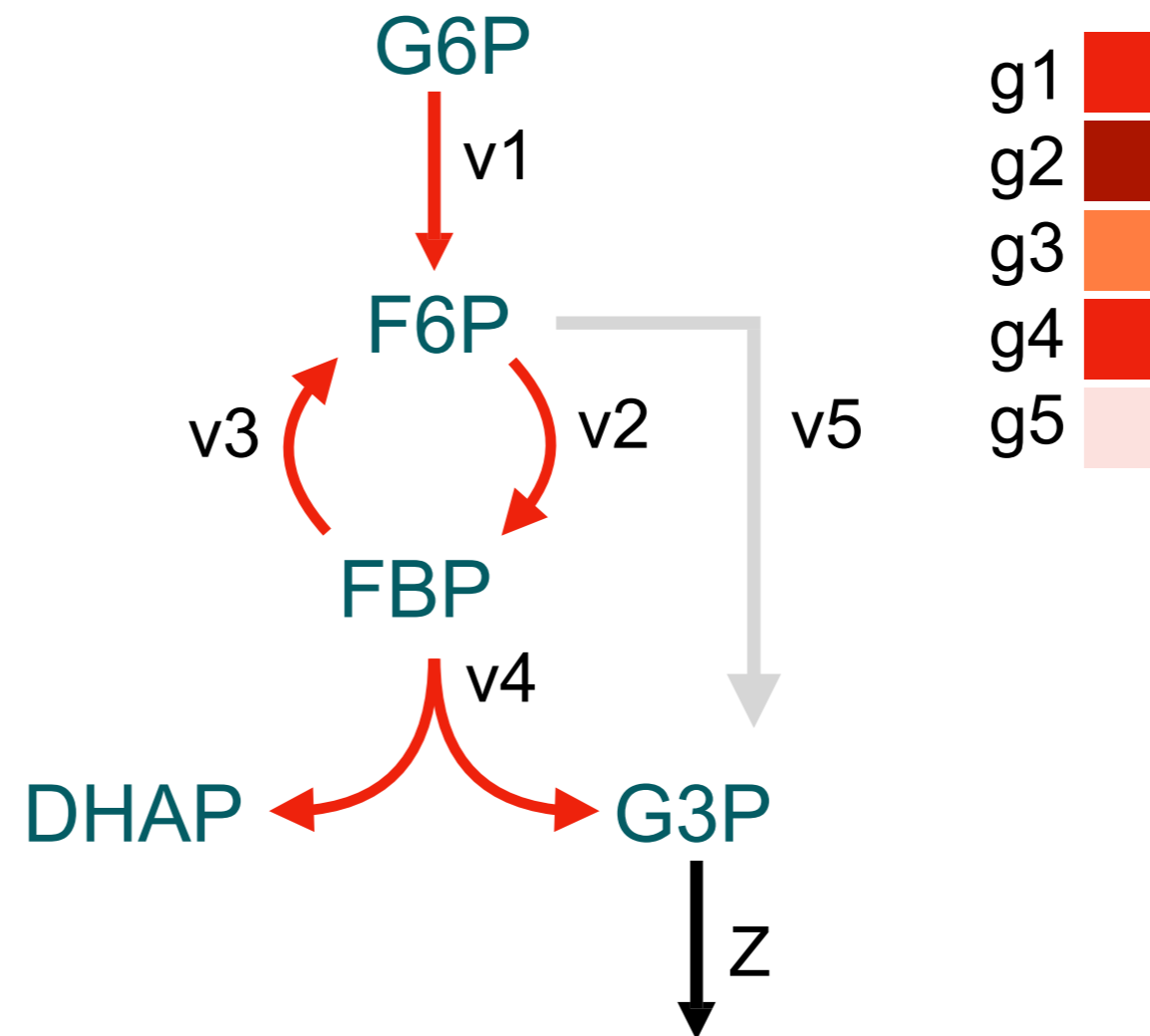
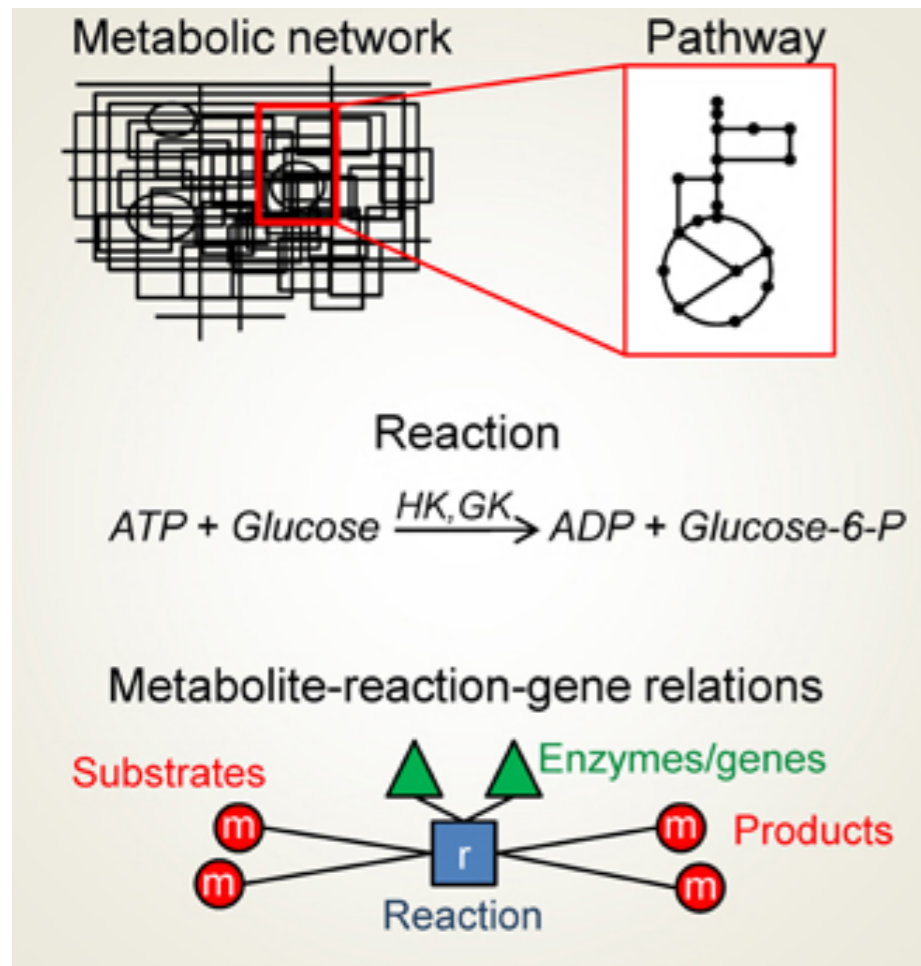




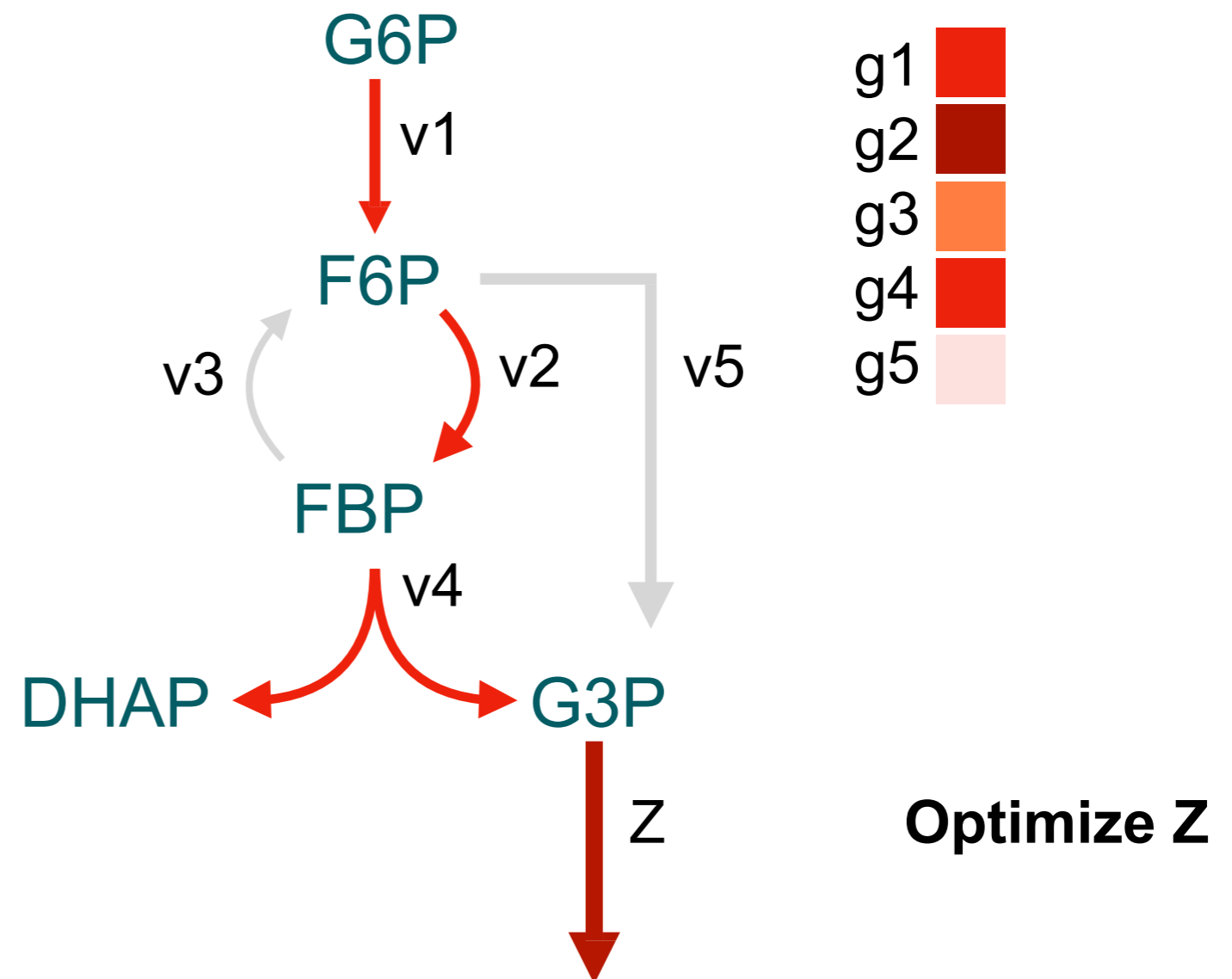
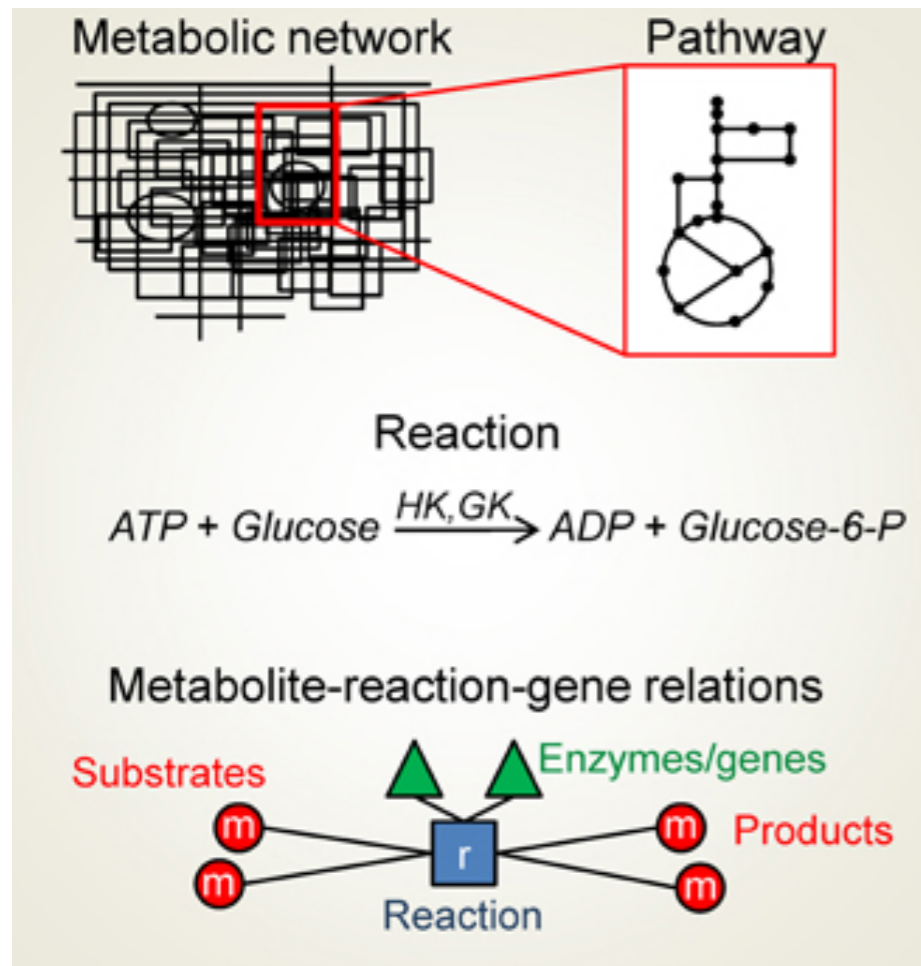
# 4. Genome-scale metabolic models as integrative networks



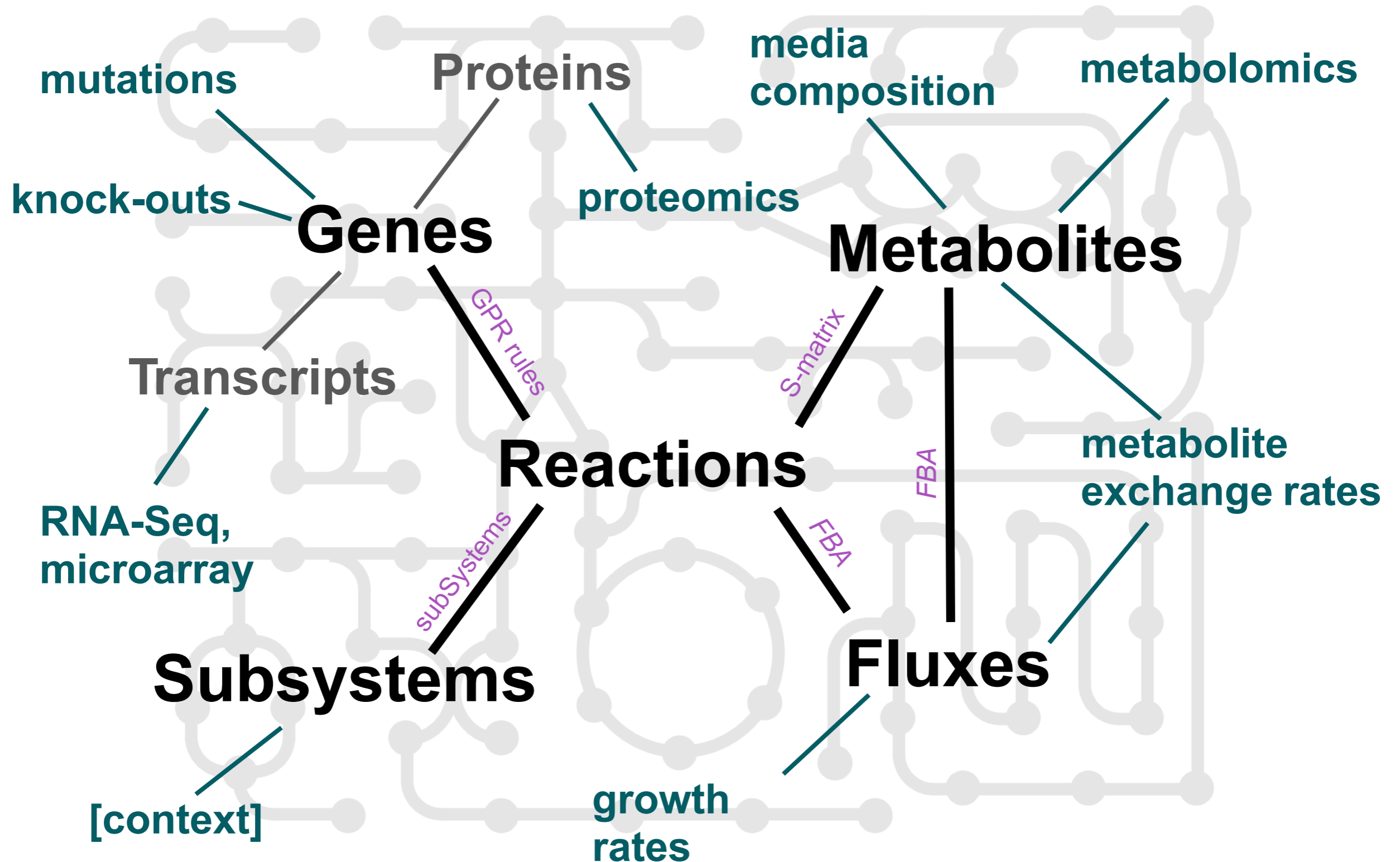
# 4. Genome-scale metabolic models as integrative networks



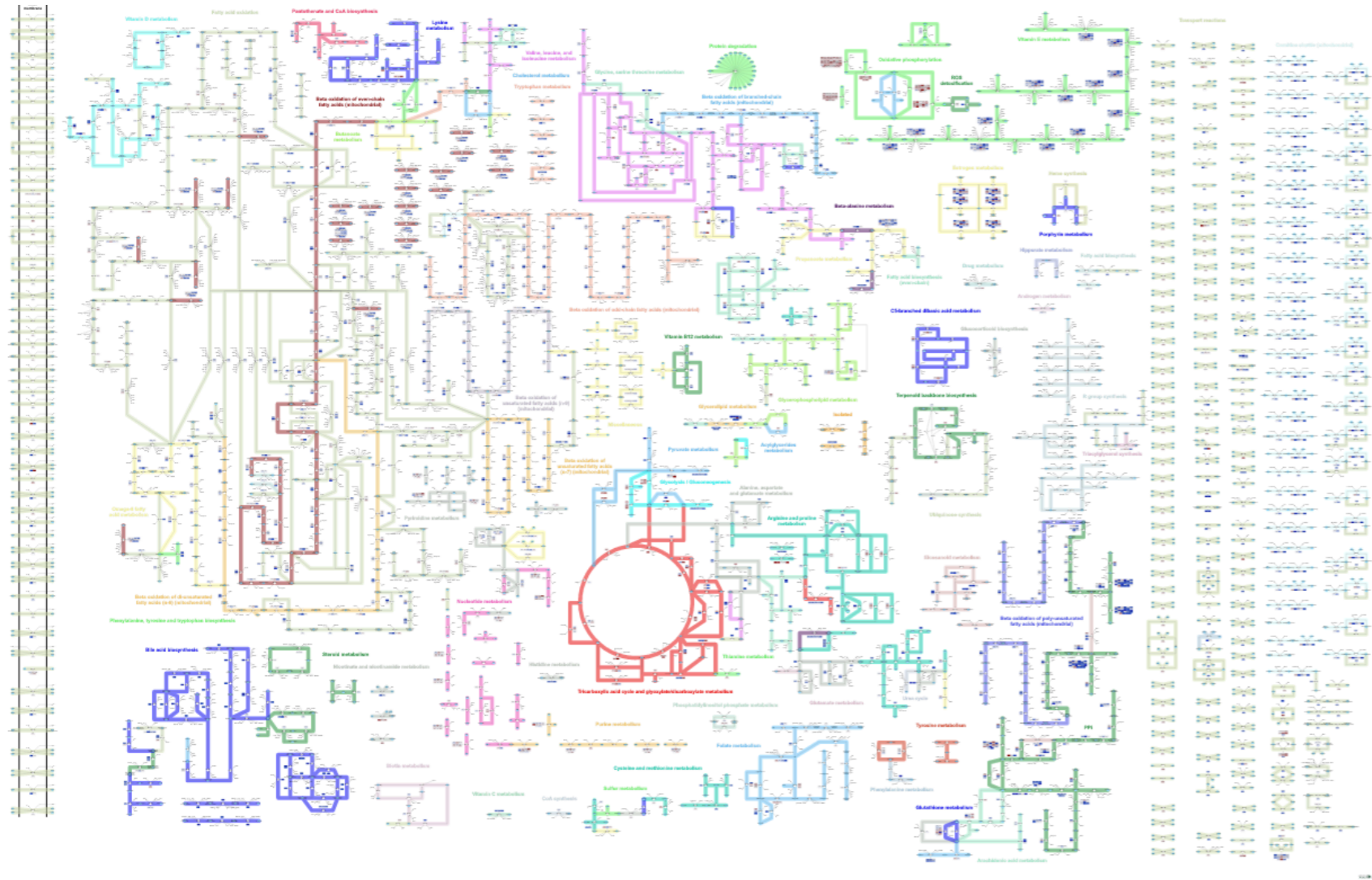
# 4. Genome-scale metabolic models as integrative networks



# 4. Genome-scale metabolic models as integrative networks



# 4. Genome-scale metabolic models as integrative networks



Simulate flux distributions

Dysregulated pathways

Reporter metabolites

Essential genes

Targetable enzymes

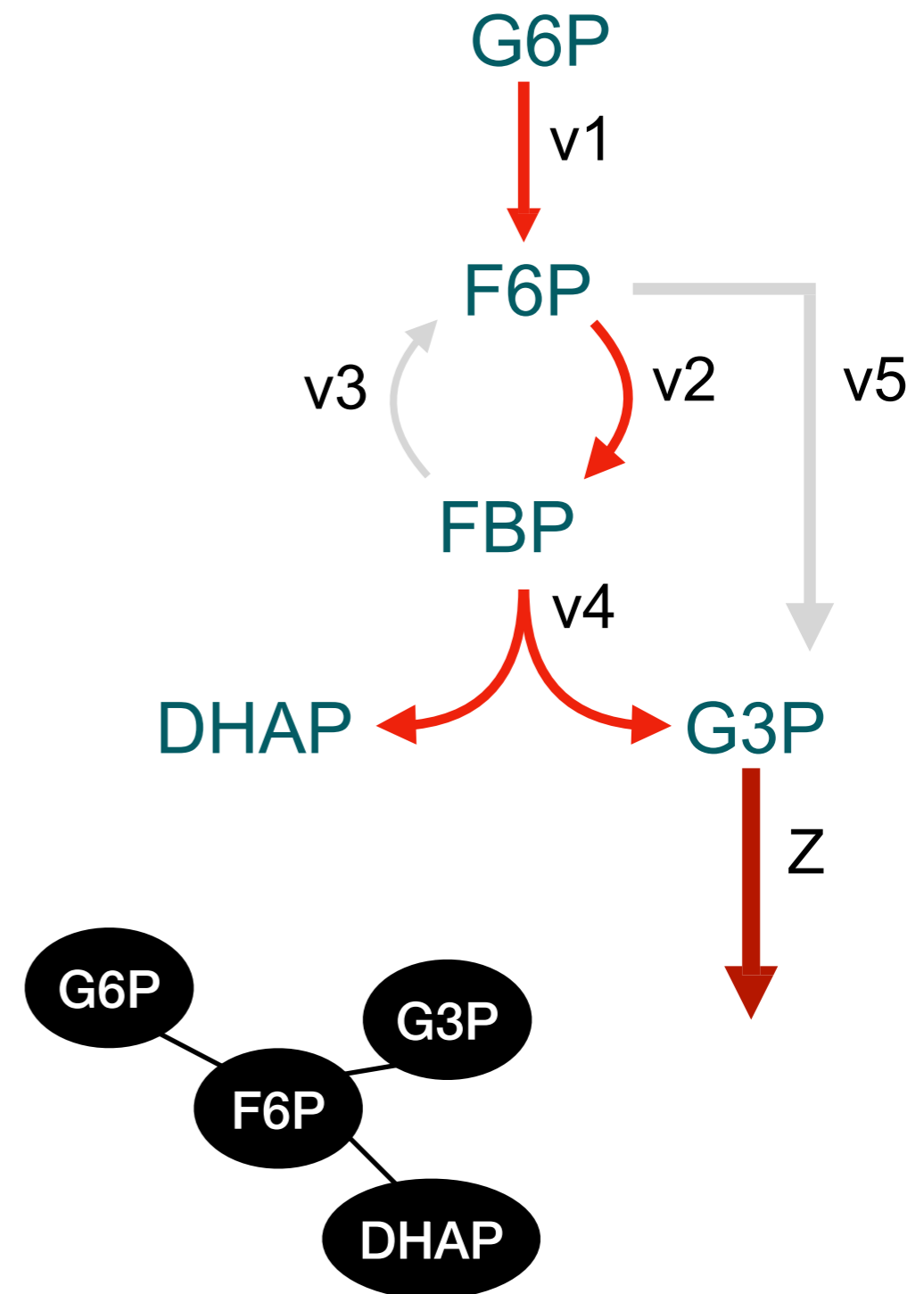
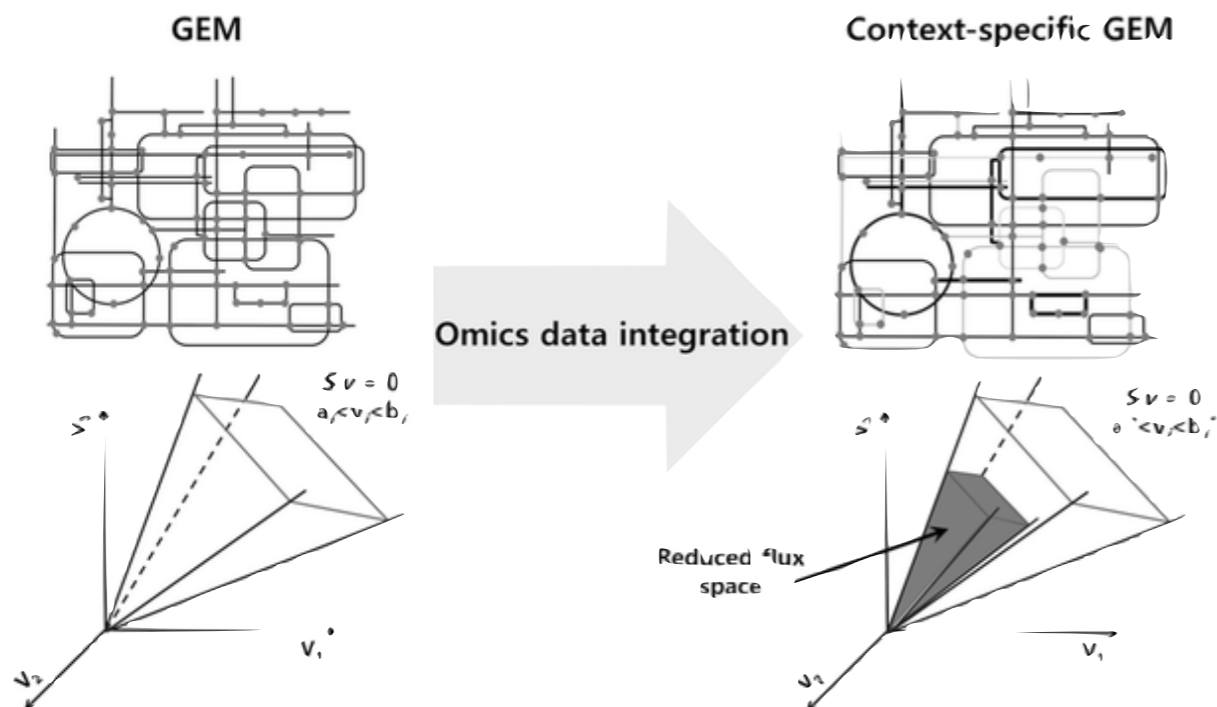
May be combined with  
standard graph analysis

# 4. Genome-scale metabolic models as integrative networks

GEMs may be used to find such missing relationships, but there is a coverage issue

The overall strategy follows

1. Integrate proteomic, transcriptomic, metabolomic, fluxomic
2. Flux distribution
3. Compute metabolite-reaction-gene relationships
4. Extract relevant relationships (met-met, gene-gene)
- 4b. Exclude unnecessary interactions (e.g. cofactors)
5. Downstream analysis (e.g. topology, stratification)



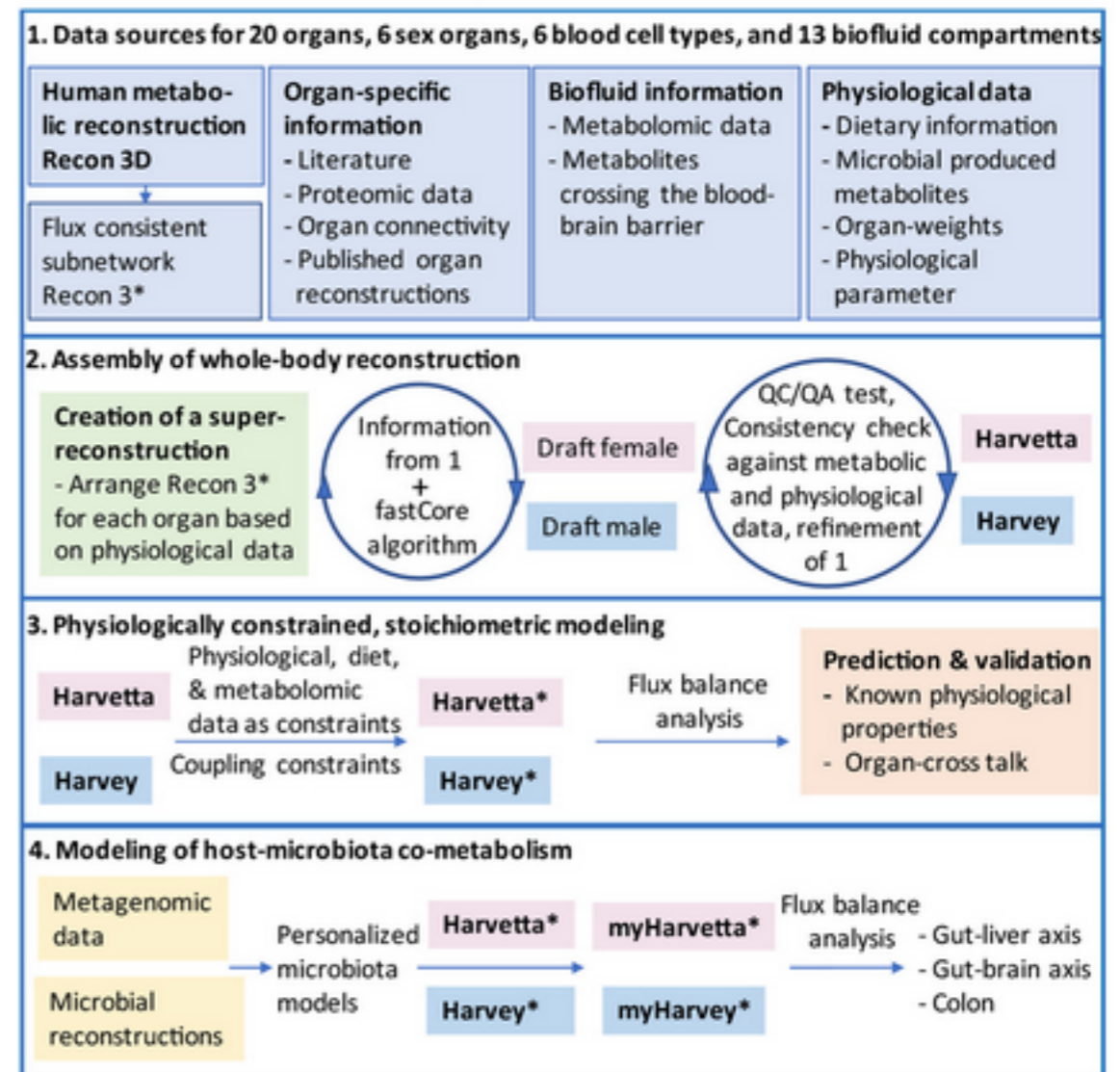
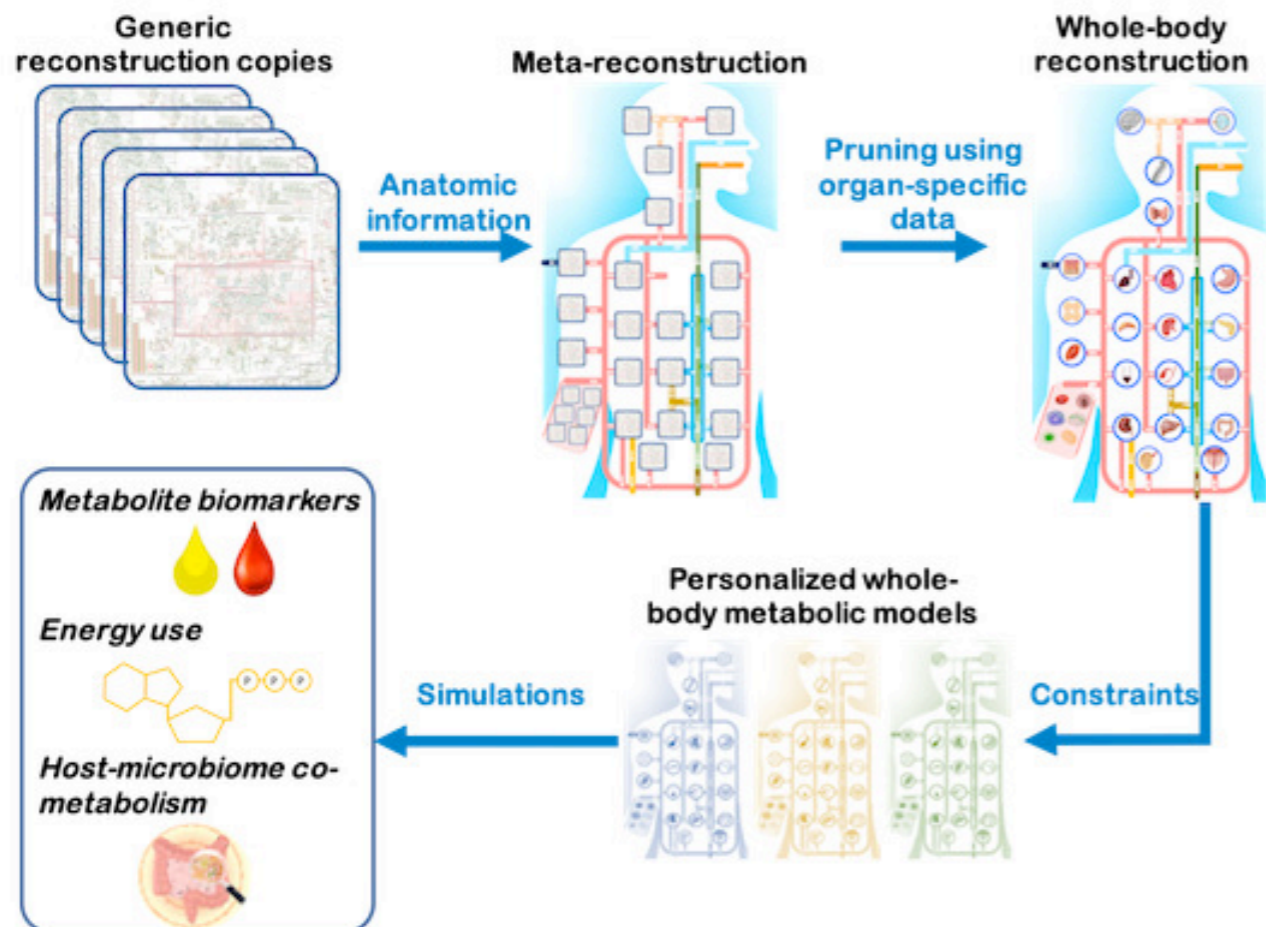
# 4. Genome-scale metabolic models as integrative networks

Personalised whole-body models of host + gut microbiome

26 organs

6 blood cell types

80,000 biochemical reactions

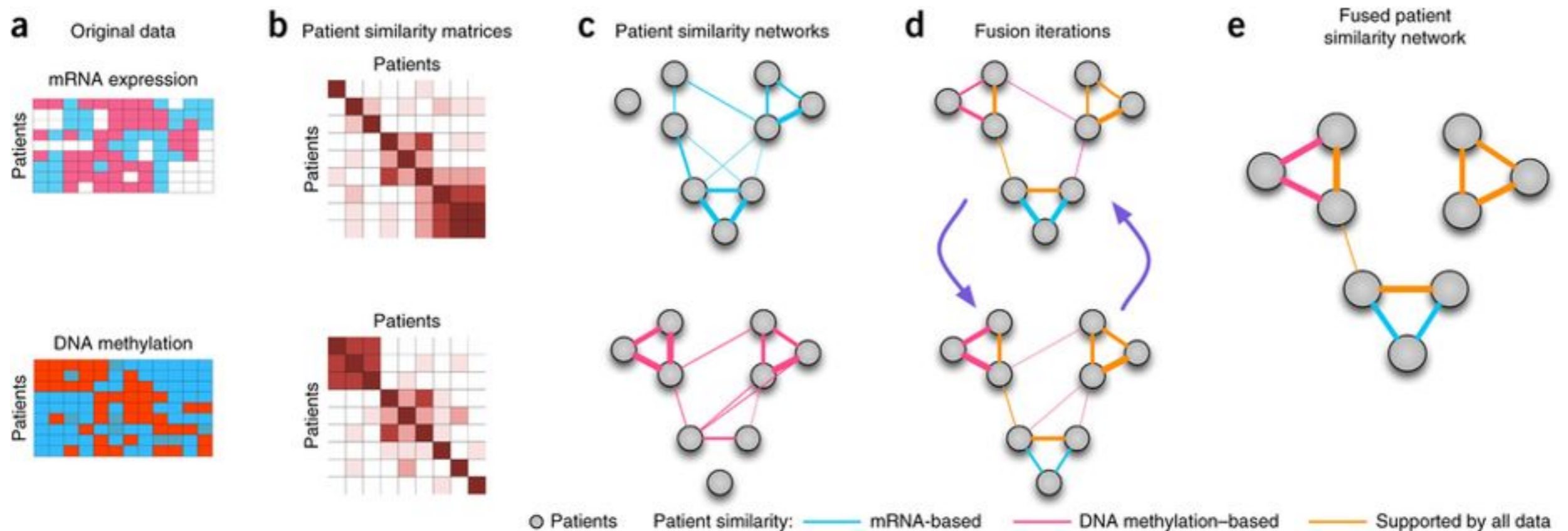


# Similarity network fusion

Sample-sample clustering based on multi-omic data improves clustering

Single-omics present complementary (non-redundant) information

Enables further comparisons between clusters





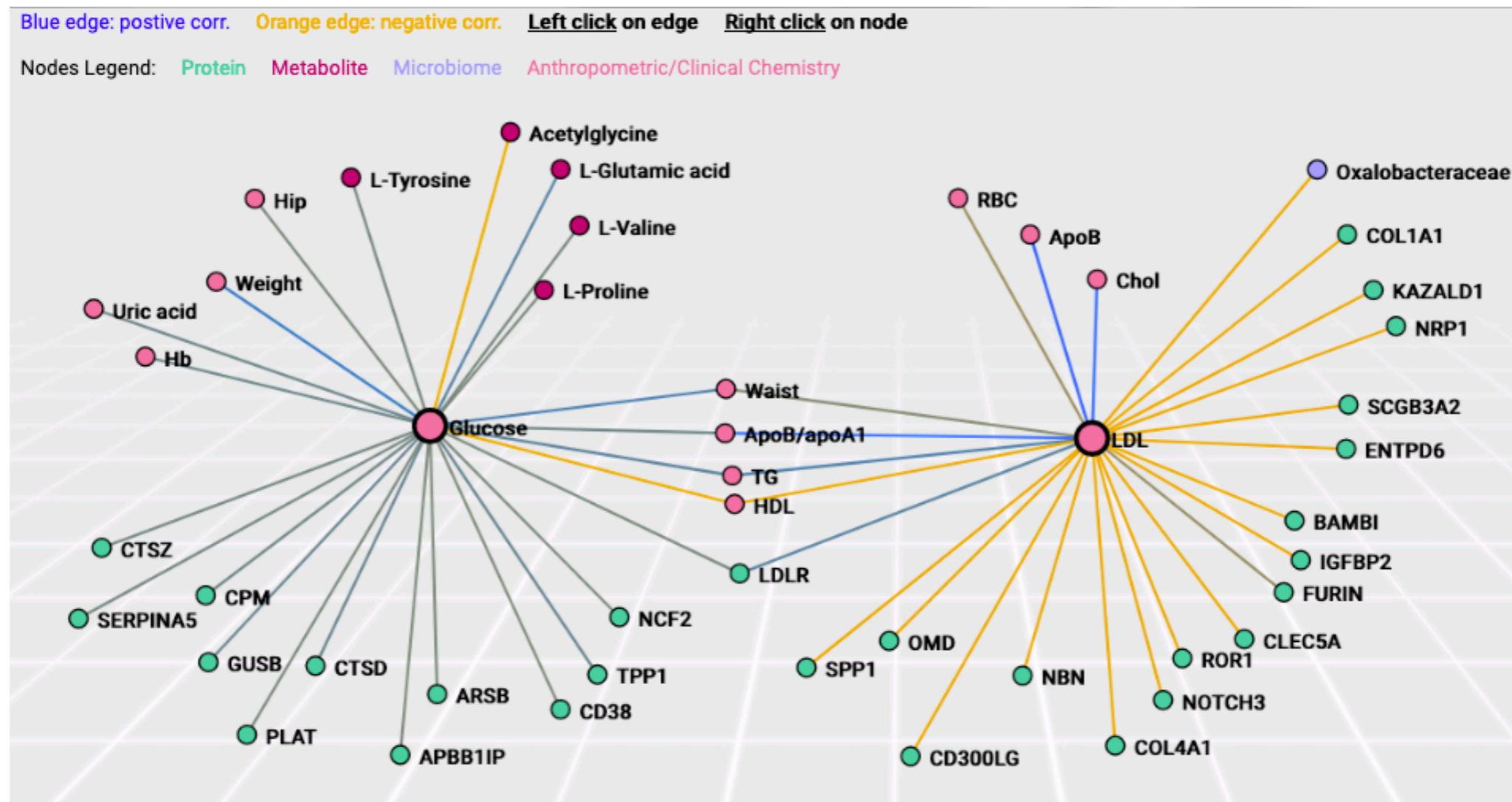
# Overview

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1. Introduction to network analysis
2. Terminology
3. Network inference
- 4. Key network properties**
5. Community analysis

# Motivation

You have built an association network (e.g. PPI, multi-omic). How to identify pivotal features, their organization, and biological characteristics?



# Key network properties to discuss

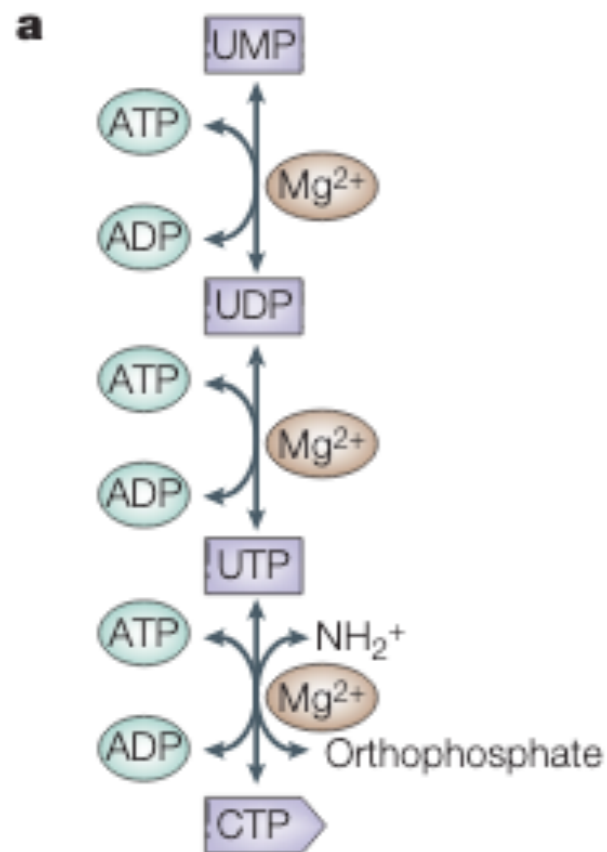
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1. Network representations
2. Network density
3. Paths
4. Centrality
5. Clustering coefficient
6. Degree and connectivity distributions

# 1. Network representations

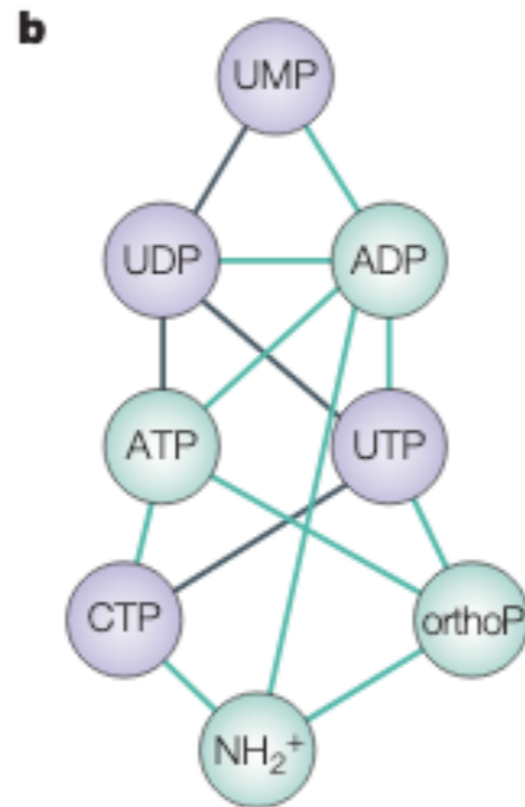
## Representations of a metabolic network: pyrimidine metabolism

Metabolism



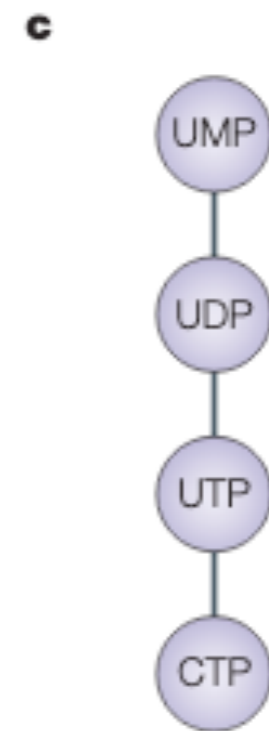
(directed graph)

Graph representation:  
metabolites and co-factors



(undirected graph)

metabolite-metabolite  
association



(undirected graph)

Other representations: Protein-Protein, Protein-Metabolite

## 2. Network density

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A **dense graph** is a graph where the number of edges approximates the maximum possible number of edges for the given node number.

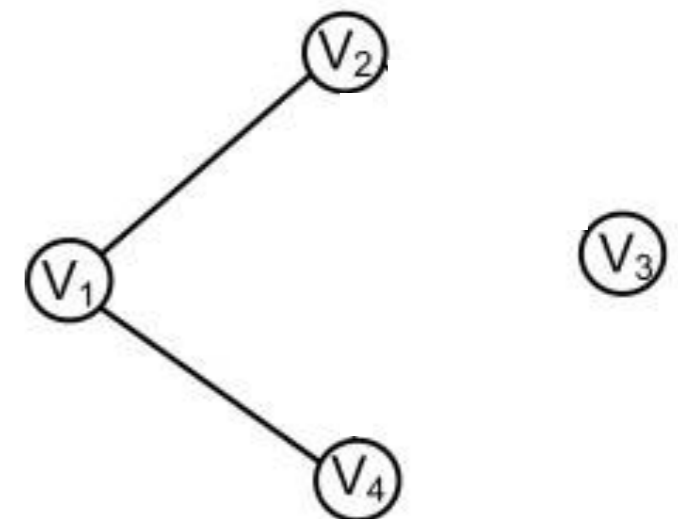
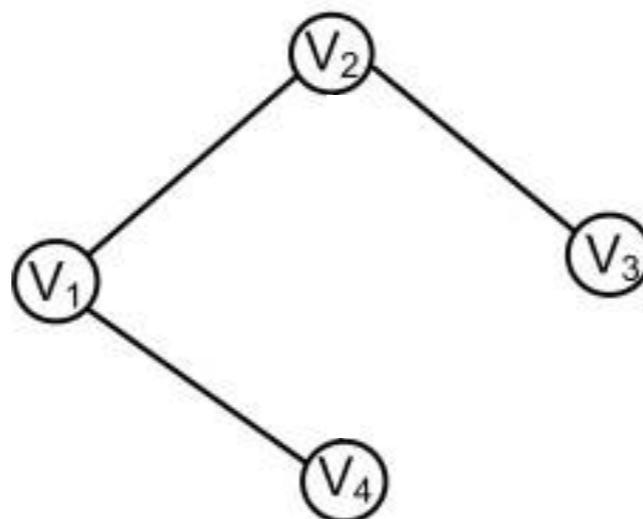
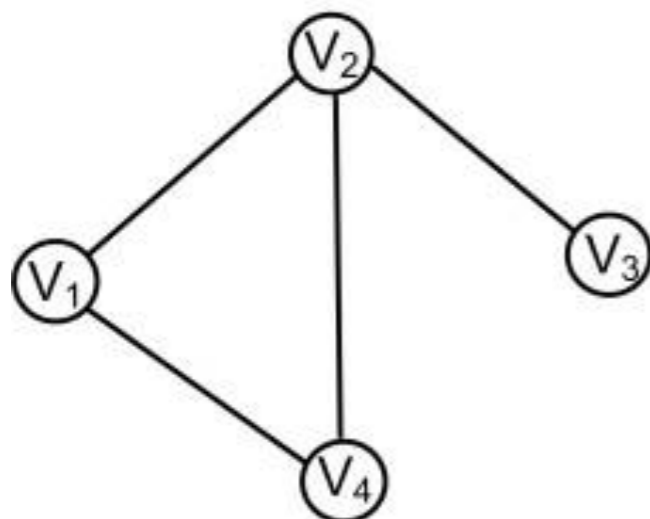
We can thus compute the network **density** (or **global connectivity**) as

$$\text{Undirected graphs: } D = \frac{2 * E}{V \cdot (V - 1)}$$

$E$  : number of edges

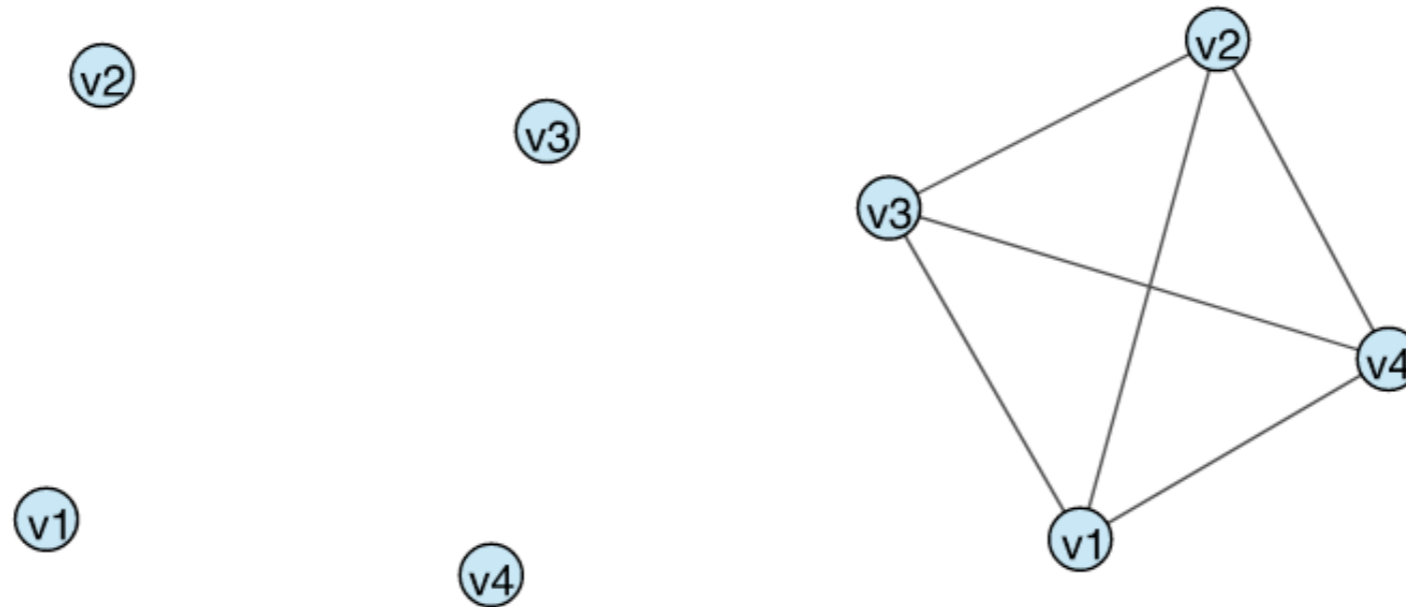
$V$  : number of vertices

$$\text{Possible edges} = \frac{V \cdot (V - 1)}{2}$$



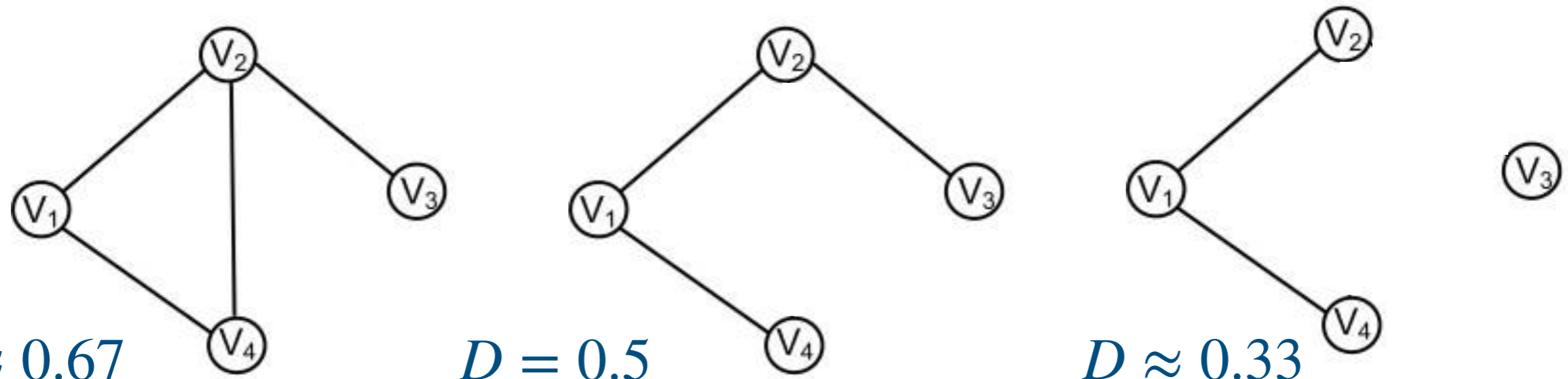
## 2. Network density

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$$0 \leq D \leq 1$$

Higher density indicates higher associations in the network, which implies lower resilience to changes.



# 2. Biological network density

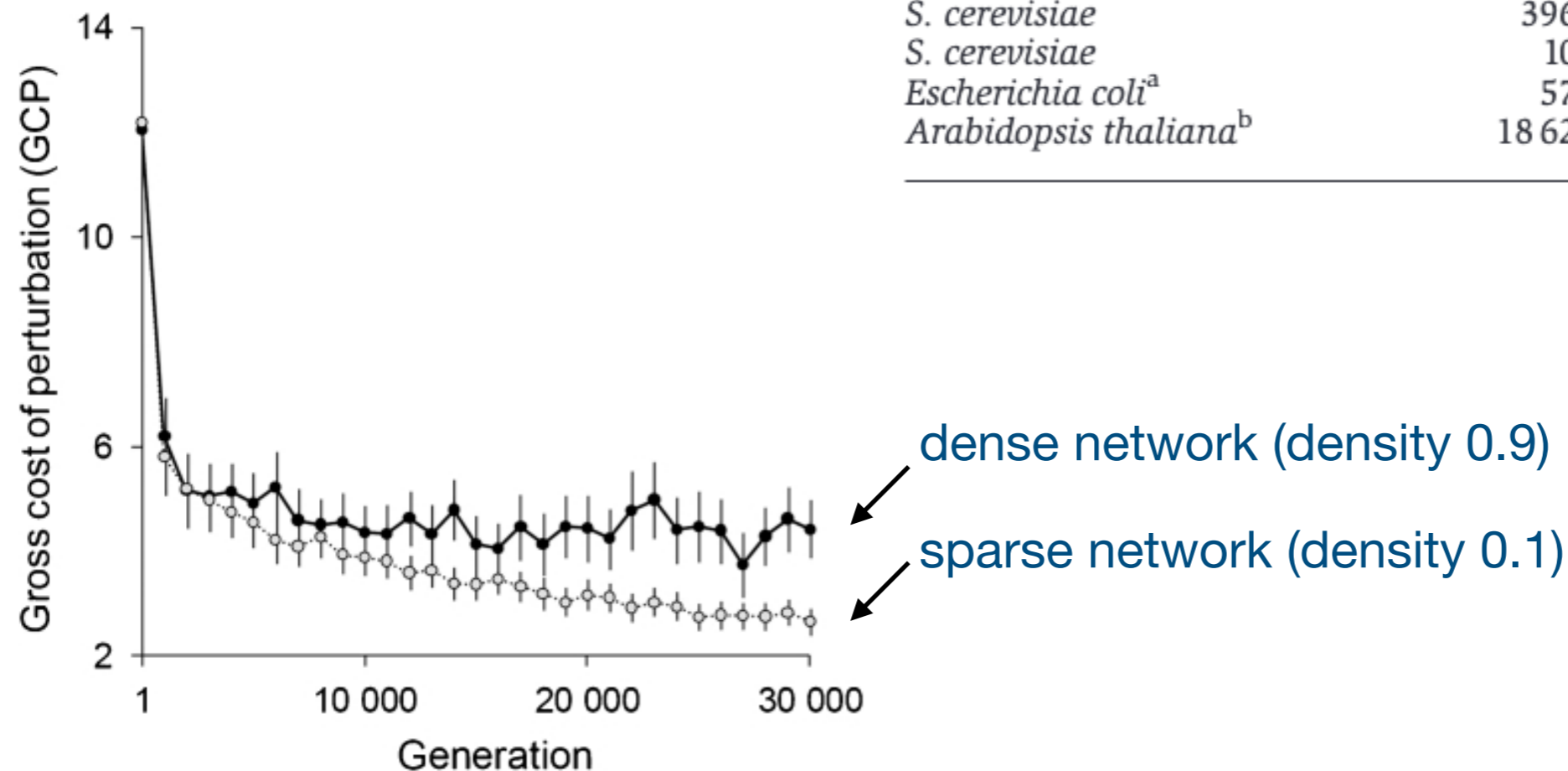
Evolutionary analysis of biological networks indicates general sparsity

Network structure must balance robustness to mutation, stochasticity and environmental queues

Sparse networks show higher robustness when accounting for costs and benefits of complexity

**Table I** Biological networks are sparsely connected

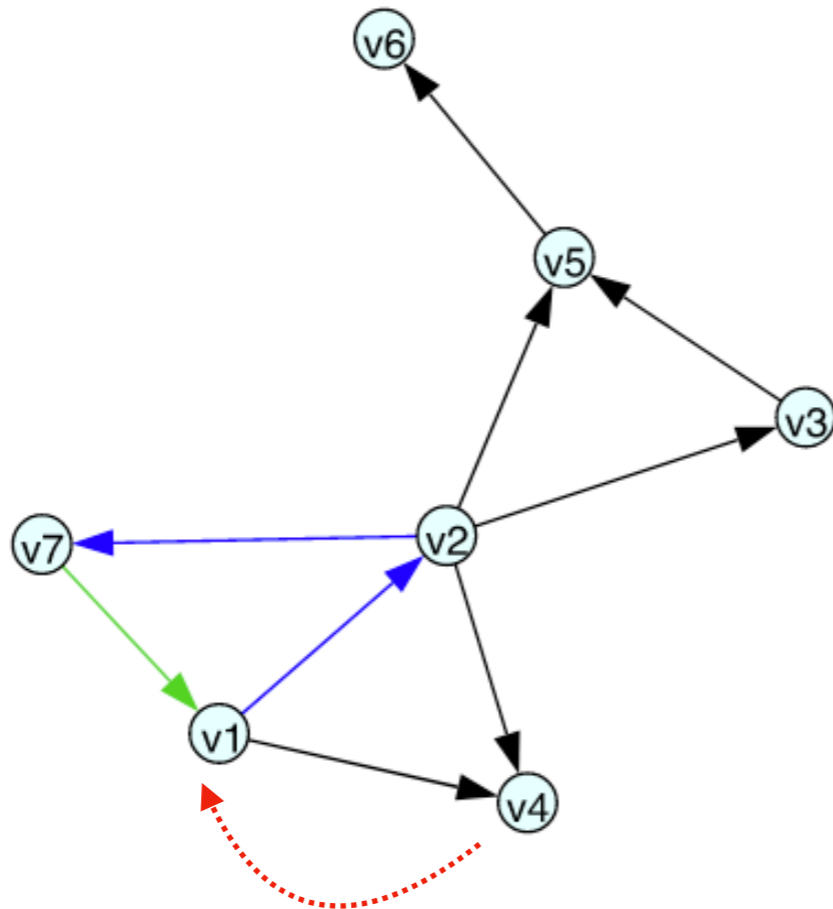
Organism	Interactions	Genes	$D$
<i>Drosophila melanogaster</i>	29	14	0.148
<i>D. melanogaster</i>	45	25	0.072
Sea urchin	82	44	0.0065
<i>Saccharomyces cerevisiae</i>	1052	678	0.0023
<i>S. cerevisiae</i>	3969	2341	0.0007
<i>S. cerevisiae</i>	106	56	0.0338
<i>Escherichia coli</i> <sup>a</sup>	578	423	0.0032
<i>Arabidopsis thaliana</i> <sup>b</sup>	18 625	6760	0.0004



# 3. Paths

Distance between nodes is measured in path length

In directed graphs, the shortest path between  $(a, b) \neq (b, a)$



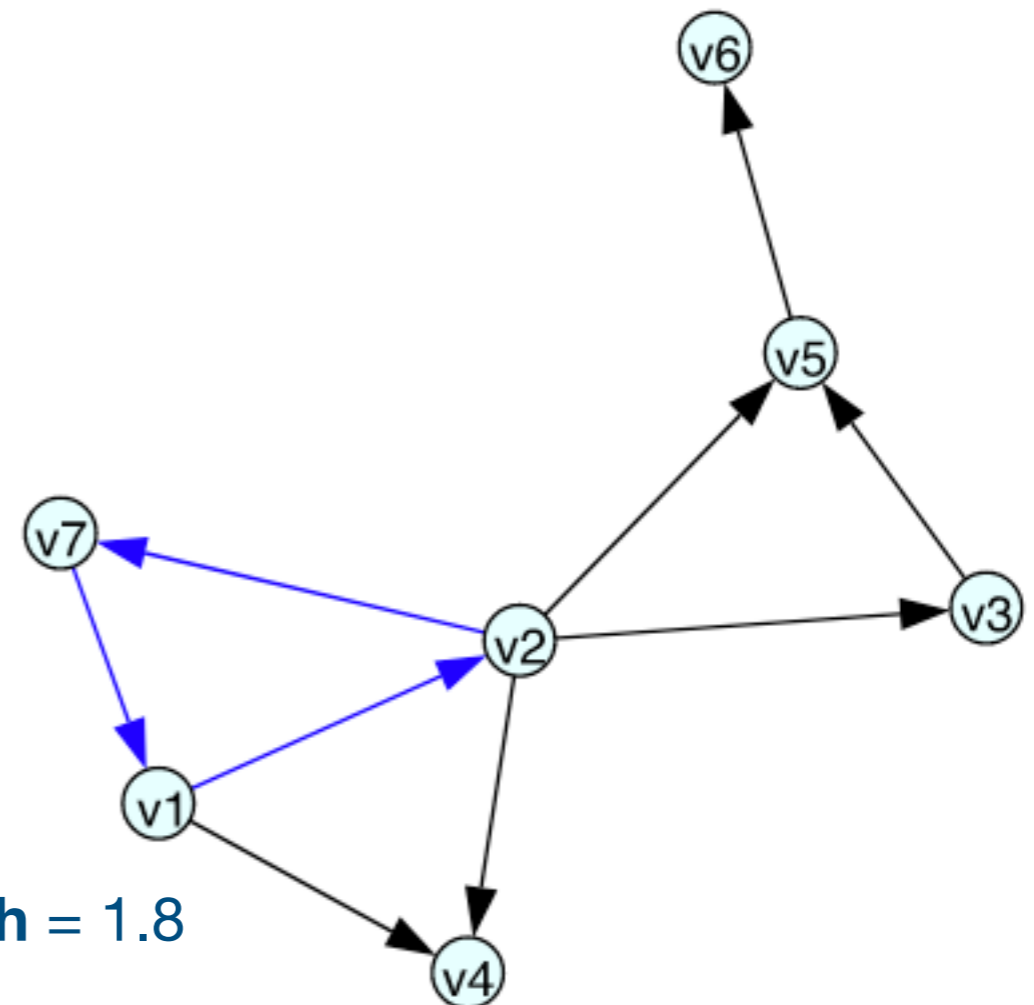
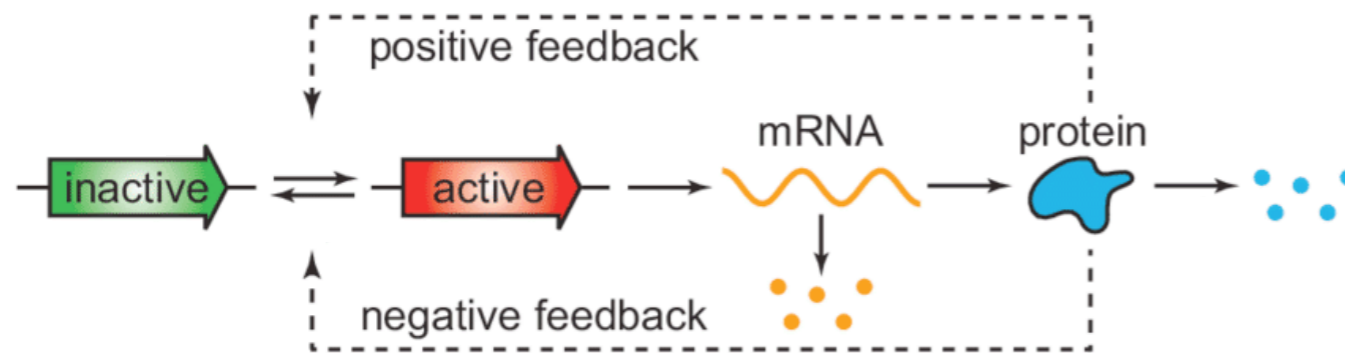
	v1	v2	v4	v3	v5	v7	v6
v1	0.0	1.0	1.0	2.0	2.0	2.0	3.0
v2	2.0	0.0	1.0	1.0	1.0	1.0	2.0
v4	inf	inf	0.0	inf	inf	inf	inf
v3	inf	inf	inf	0.0	1.0	inf	2.0
v5	inf	inf	inf	inf	0.0	inf	1.0
v7	1.0	2.0	2.0	3.0	3.0	0.0	4.0
v6	inf	inf	inf	inf	inf	inf	0.0



# 3. Paths

## Cycles and acyclic graphs

The **average path** gives a measure of network navigability (~feature relationships)



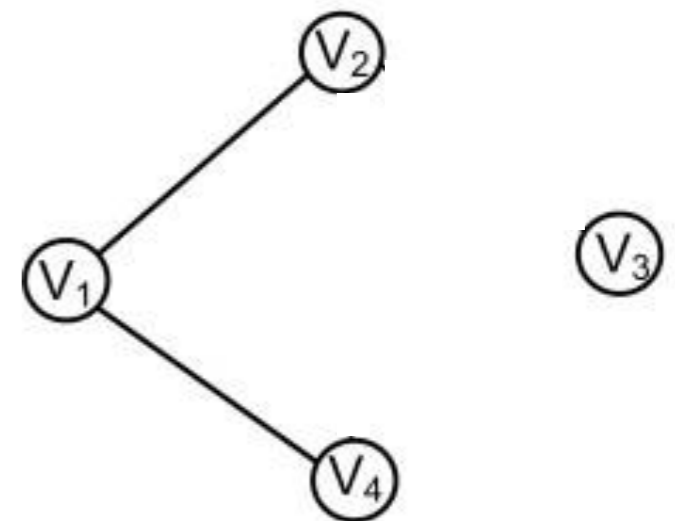
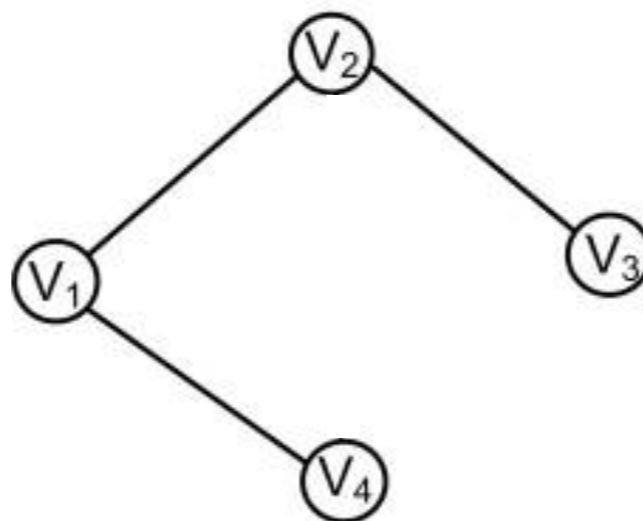
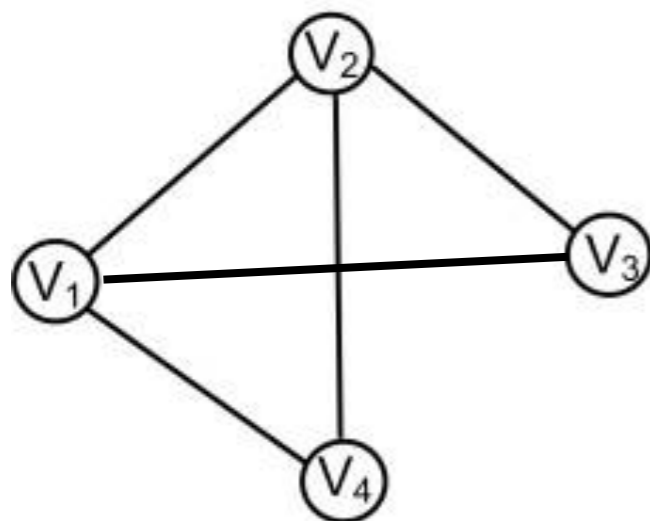
Average path length = 1.8

# 4. Connectivity

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**Node connectivity**  $\kappa(G)$ : minimum number of **nodes** whose removal renders the network disconnected

**Edge connectivity**  $\lambda(G)$ : minimum number of **edges** whose removal renders the network disconnected



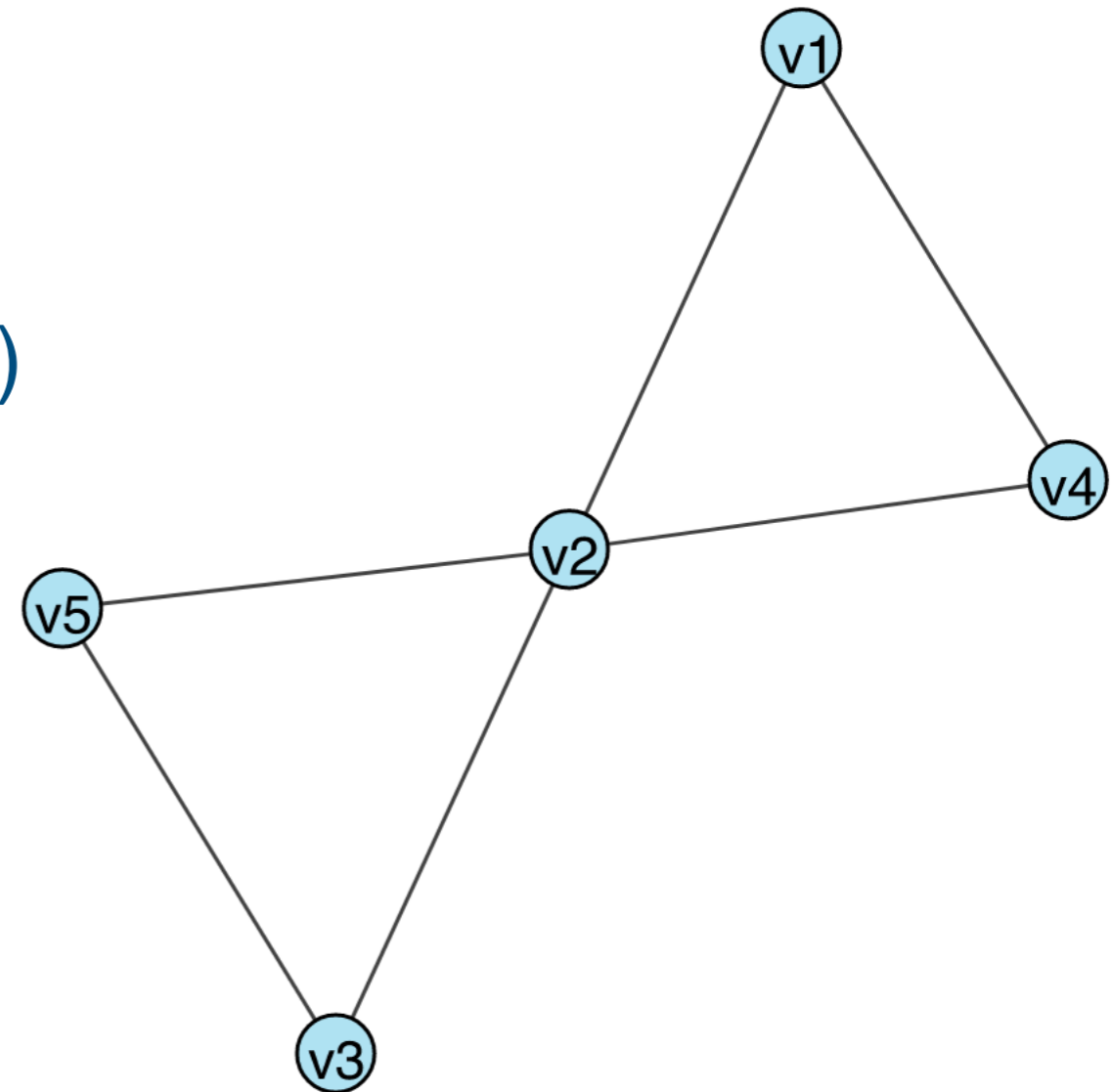
# 4. Connectivity

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$\kappa(G) = 1$ ; **cut:**  $v_2$

$\lambda(G) = 2$ ; **bridge:**  $((v_2, v_1) \ \& \ (v_2, v_4))$

**Local connectivity** may also be computed for any given pair of vertices  
(e.g.  $v_3, v_1$ :  $v_2$  and associated edges)



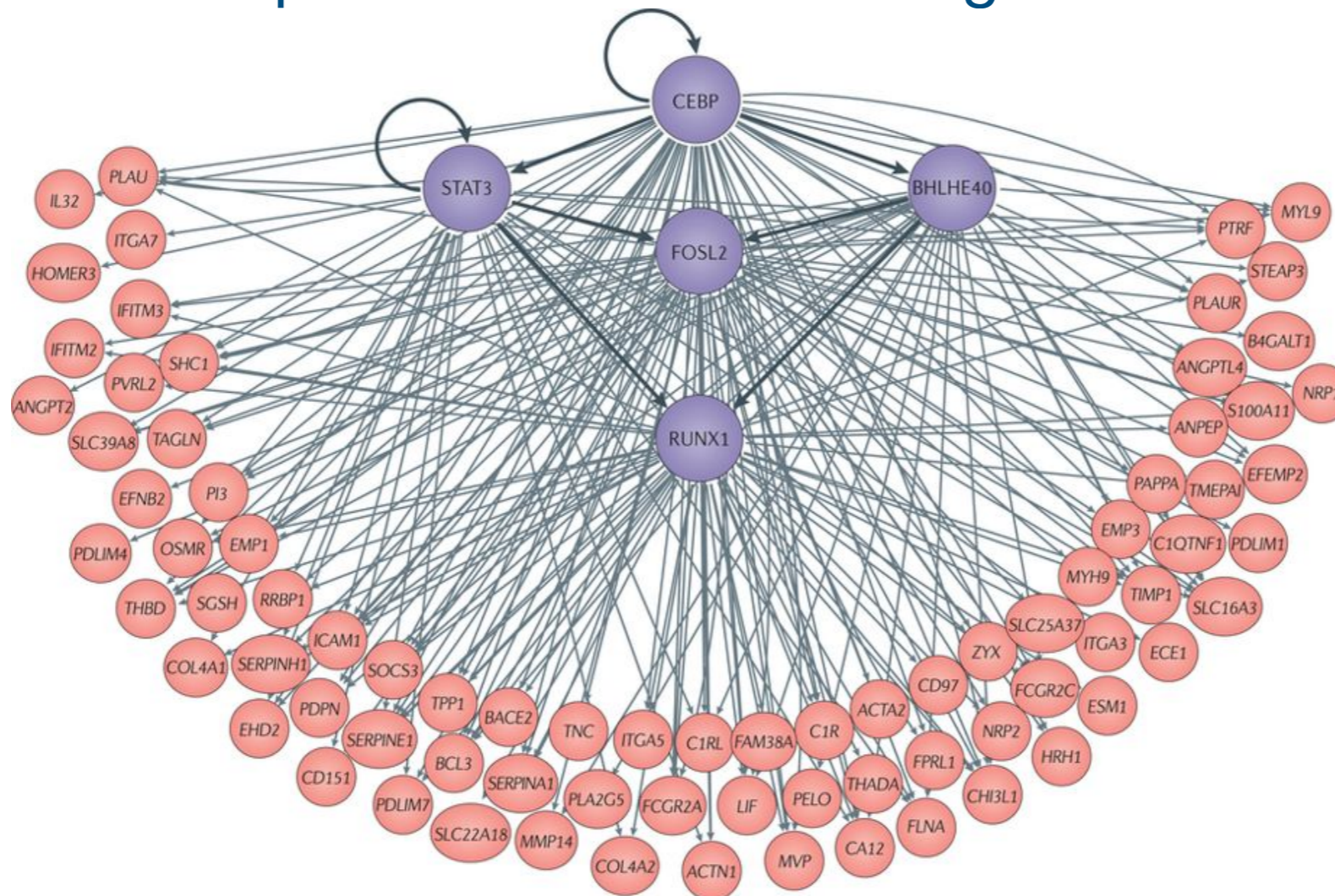
# 5. Centrality

Indicate the most central nodes in a network

Why look at the central nodes?

## Hubs

Example: Transcription Factor Master Regulators



# 5. Centrality

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Indicate the most central nodes in a network

Central nodes **possibly** most important in the network

There are many different measures of centrality:

- **Degree**
- **Eccentricity**
- **Betweenness**
- *Closeness*
- *Eigenvector*
- *PageRank*
- Katz
- Percolation
- Cross-clique
- ...

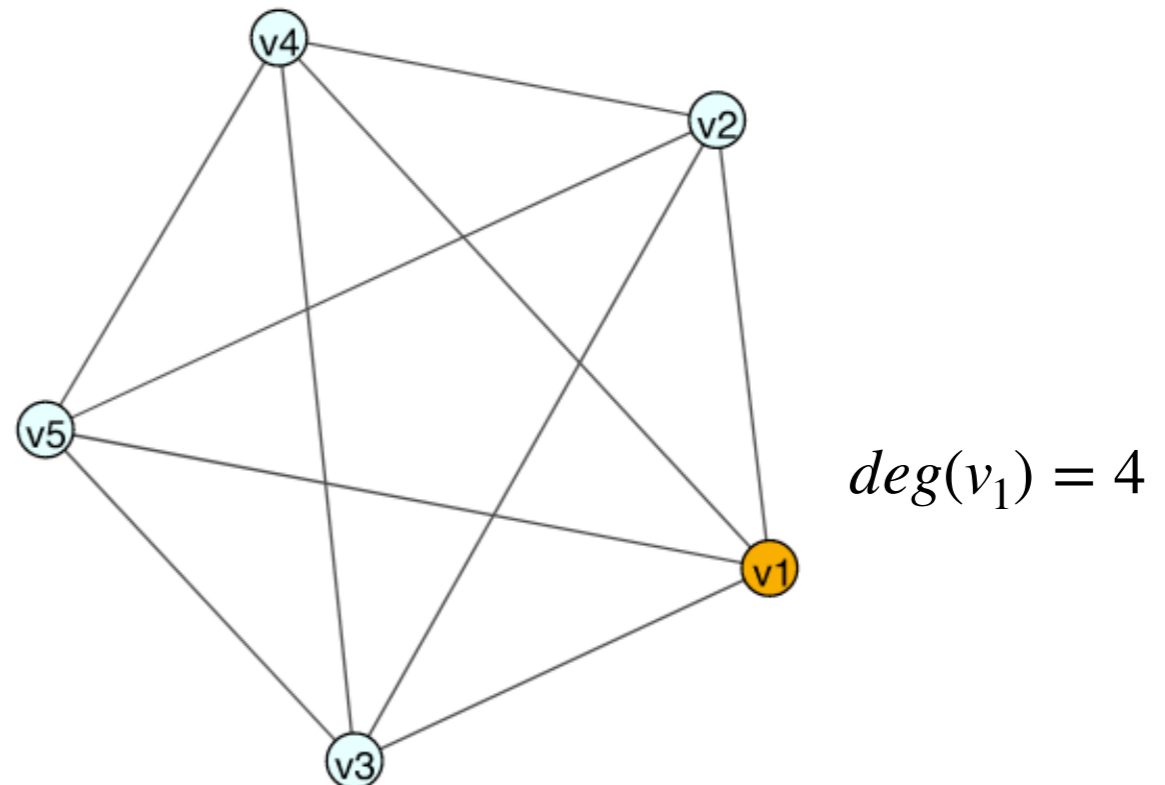
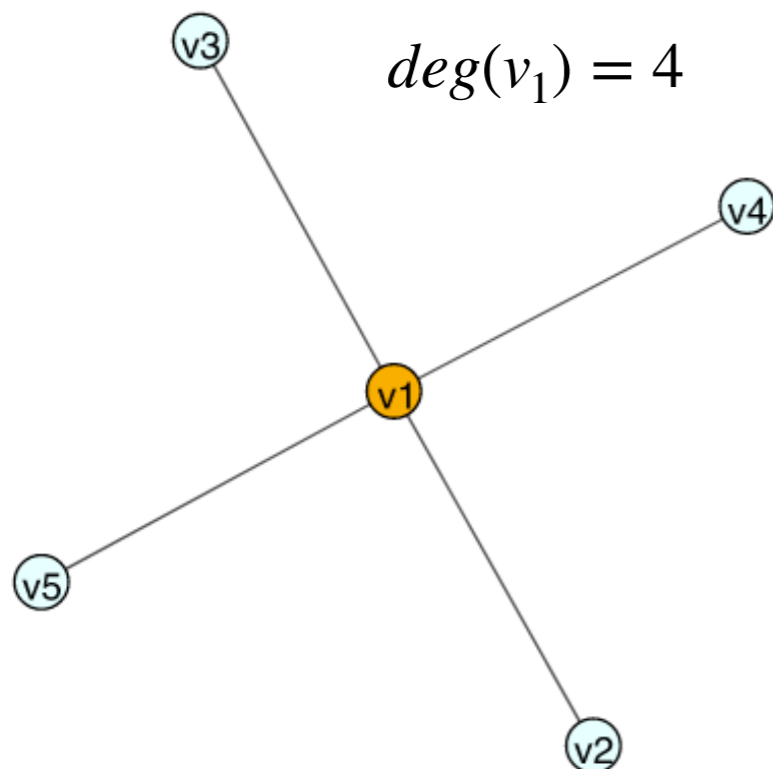
# 4. Centrality: degree centrality

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Degree indicates the number of connections with a node

$$d(v) = |N(i)|$$

where  $N(i)$  is the number of 1st neighbours of a node.



# 4. Centrality: degree centrality

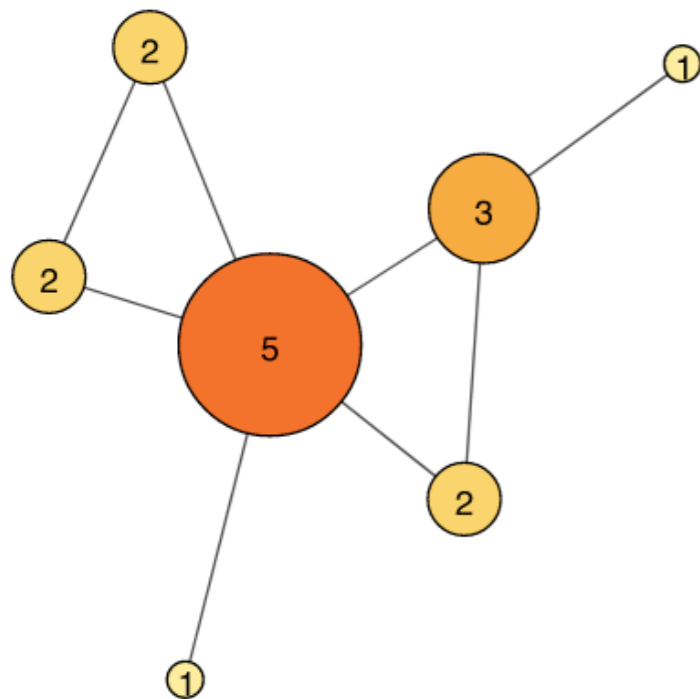
Undirected networks vs directed networks

**In-degree vs Out-degree**

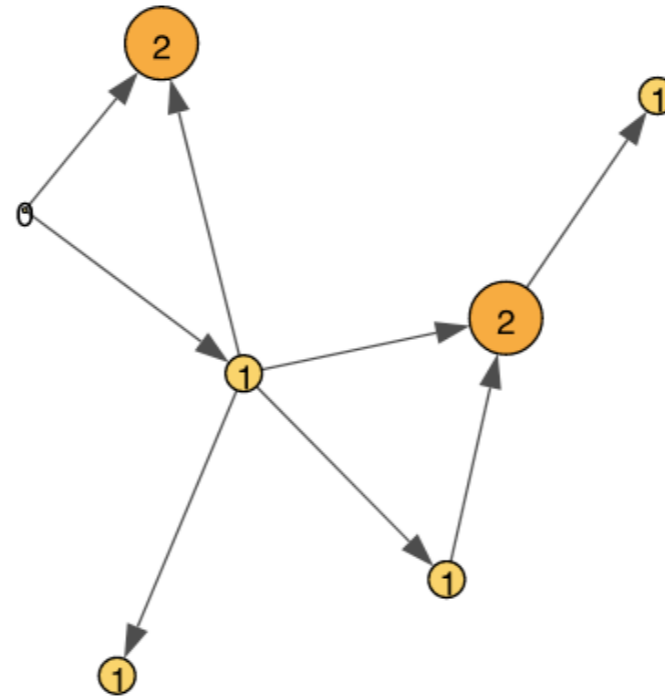
$$C_D(v_i) = \sum_{j=1}^N e_{ij}$$

Numbers indicate degree:

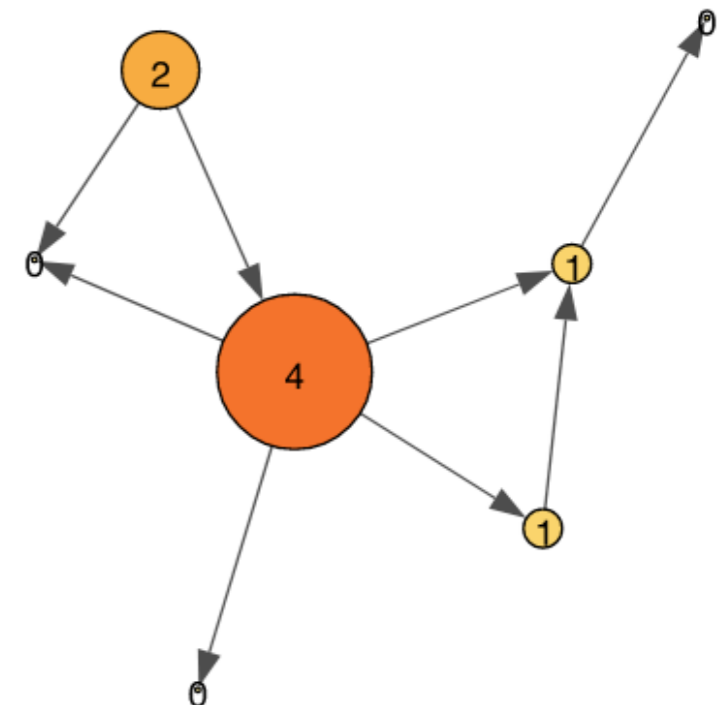
**Undirected**



**In-degree**



**Out-degree**



# 4. Centrality: degree centrality

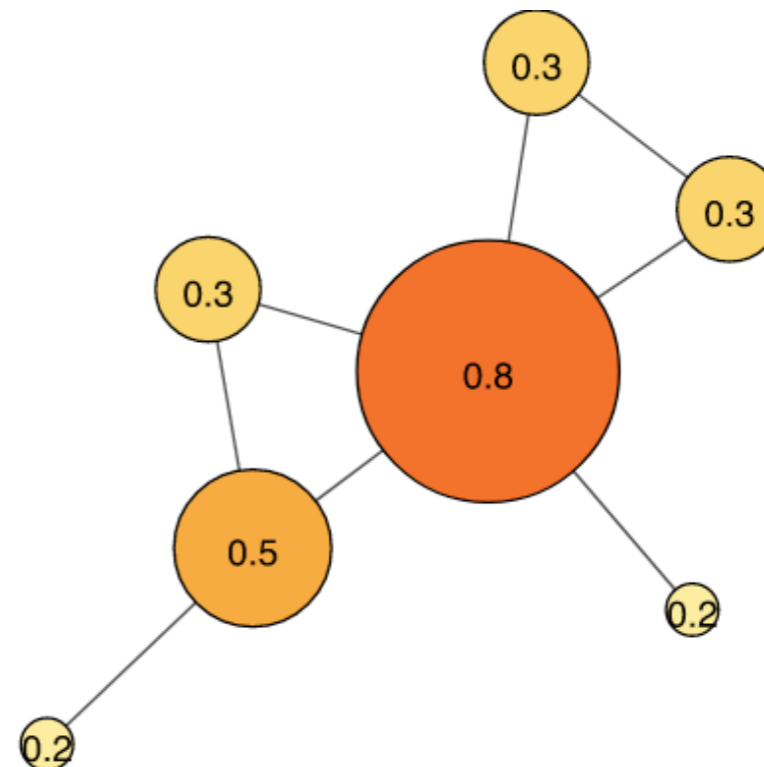
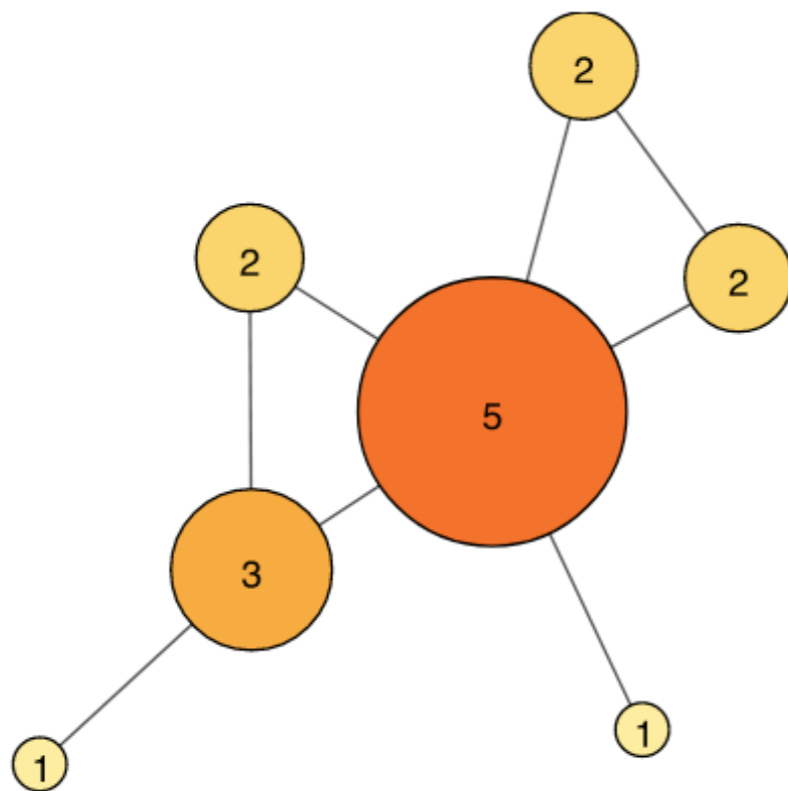
Degree centrality

$$C_D(v_i) = \sum_{j=1}^N e_{ij}$$

Normalized degree centrality

$$C_D(v_i) = \frac{\sum_{j=1}^N e_{ij}}{N-1}$$

Centrality normalization allows for comparison between networks of different sizes

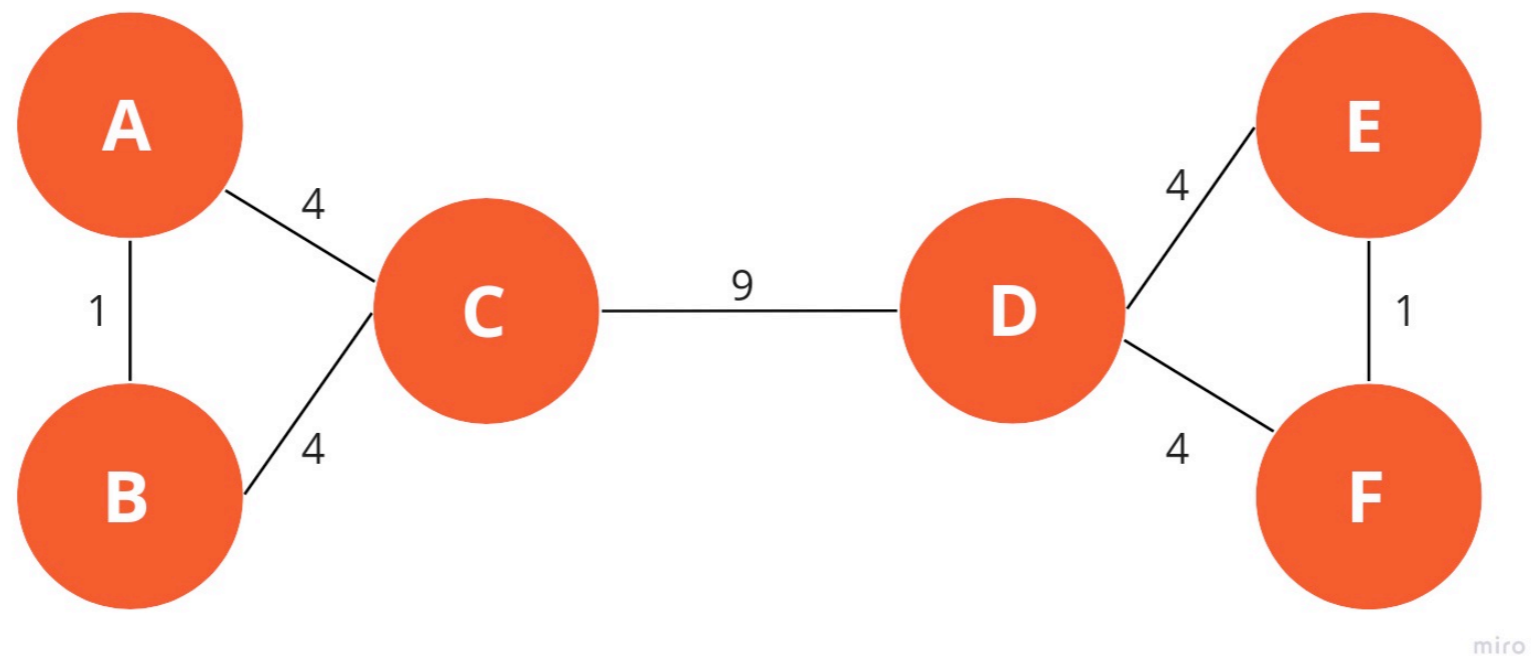




# 4. Centrality: betweenness centrality

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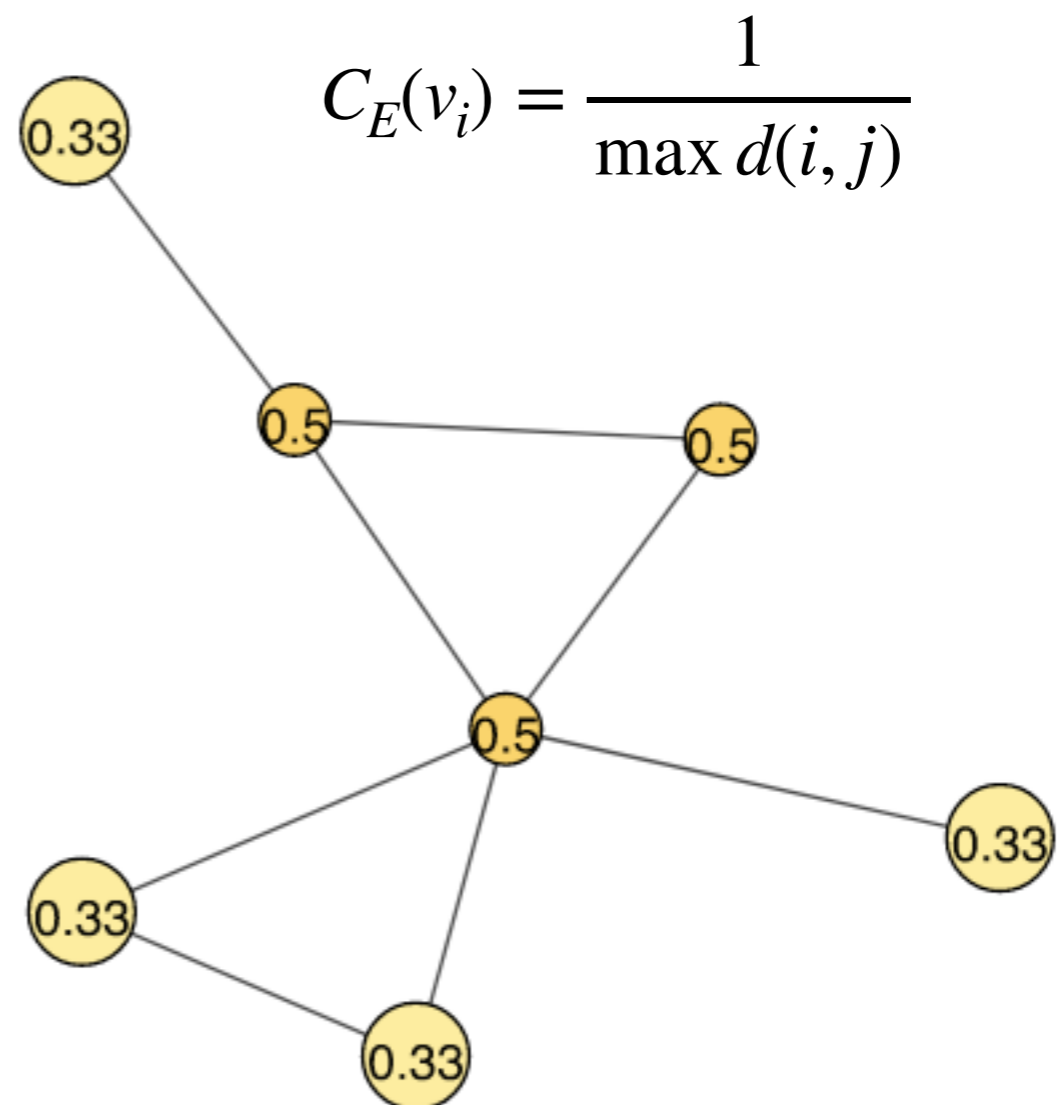
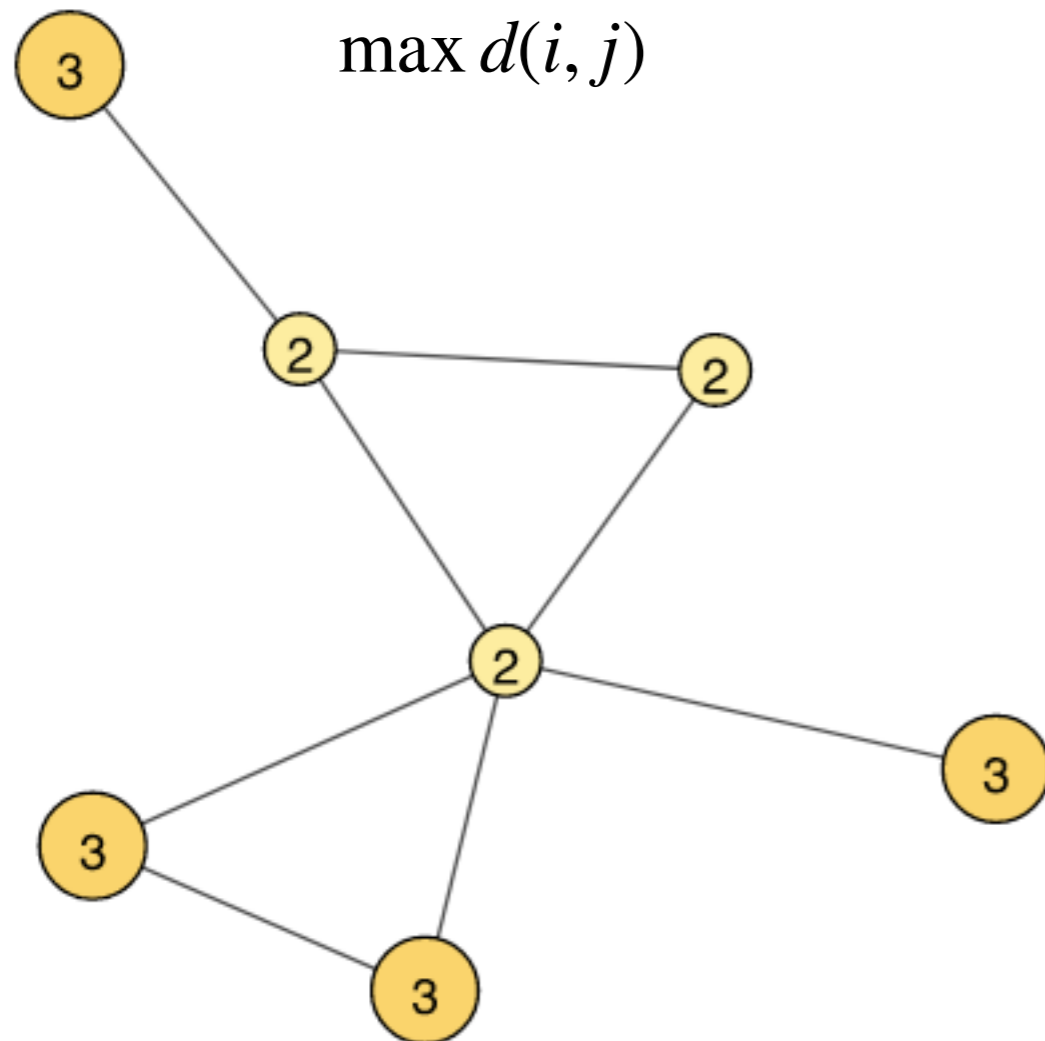
**Betweenness** considers the number of shortest paths passing through each edge



# 4. Centrality: eccentricity centrality

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**Eccentricity** considers a node's maximum shortest path to all other nodes



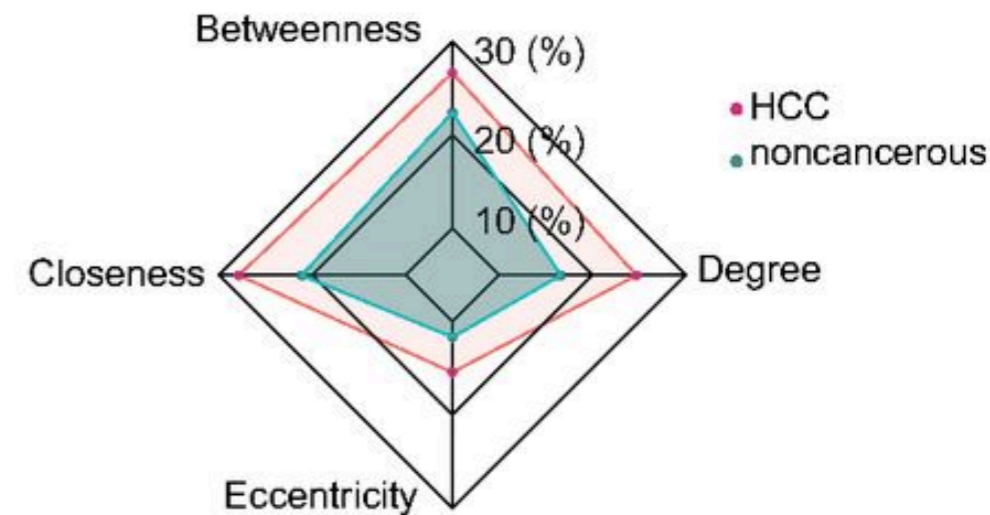
# 5. Centrality: limitations & influence

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Node centrality is used as proxy for **importance**

Should be:

1. Complement with experimental observations
2. Compute multiple metrics and summarise joint observations
3. Compute node **influence**, modifications of centrality
  - Accessibility
  - Dynamic influence
  - Impact
  - Expected force



Measure **information transmission** rather than *connectiveness*

# 6. Clustering coefficient

---

How likely is it that two connected nodes are part of a highly connected group of nodes?

If node  $v_1$  is connected with  $v_2$  and  $v_3$ , it is very likely that  $v_2$  and  $v_3$  are also connected.

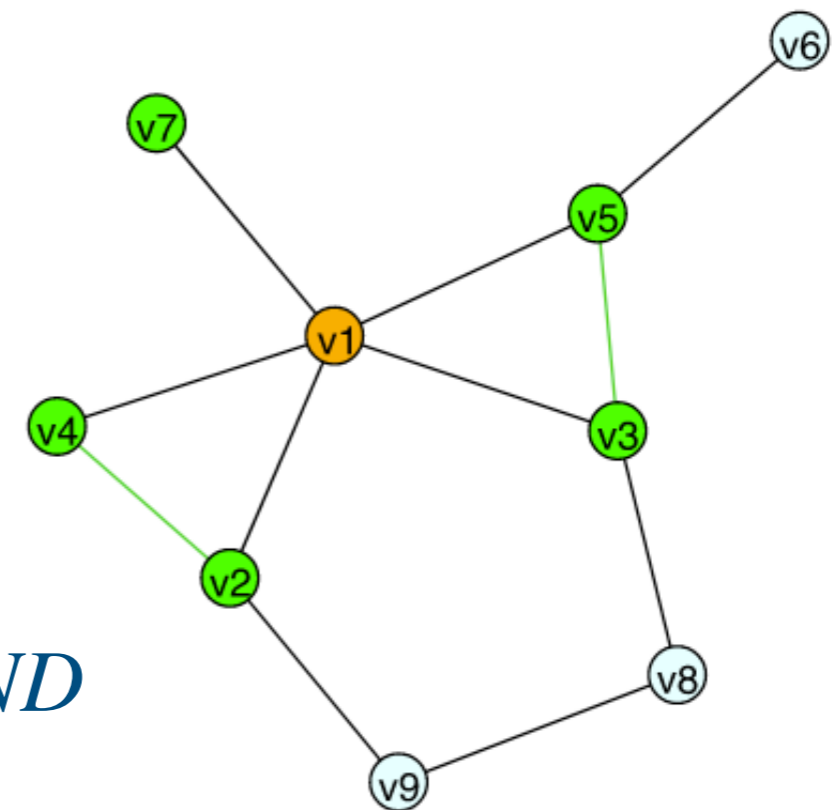
Takes into account degree of a node and the degree of its 1st neighbours

For node  $v_1$

- $deg(v_1) = k = 5$
- $n$  connections between 1st neighbours of  $v_1 = 2$

$$C_i = \frac{2 \cdot n}{k \cdot (k - 1)}$$

$$C(v_1) = \frac{2 \cdot 2}{5 \cdot 4} = 0.2 \quad C(v_7) = \frac{2 \cdot 0}{1 \cdot 0} = 0 \text{ or } ND$$

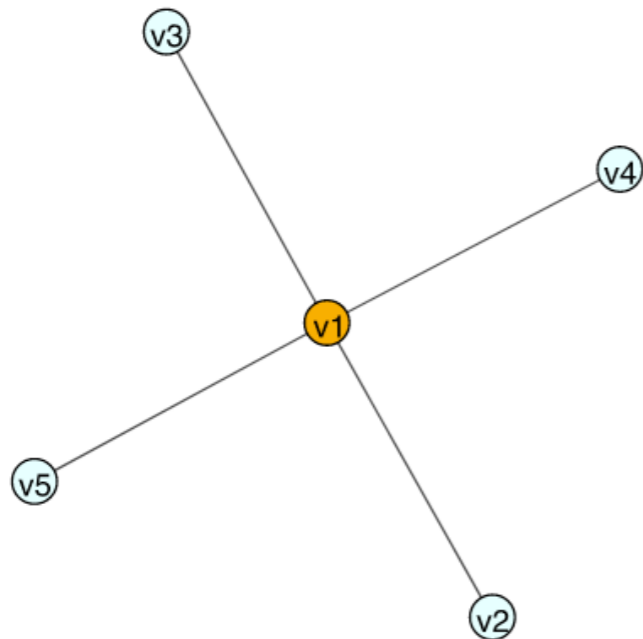


# 6. Clustering coefficient

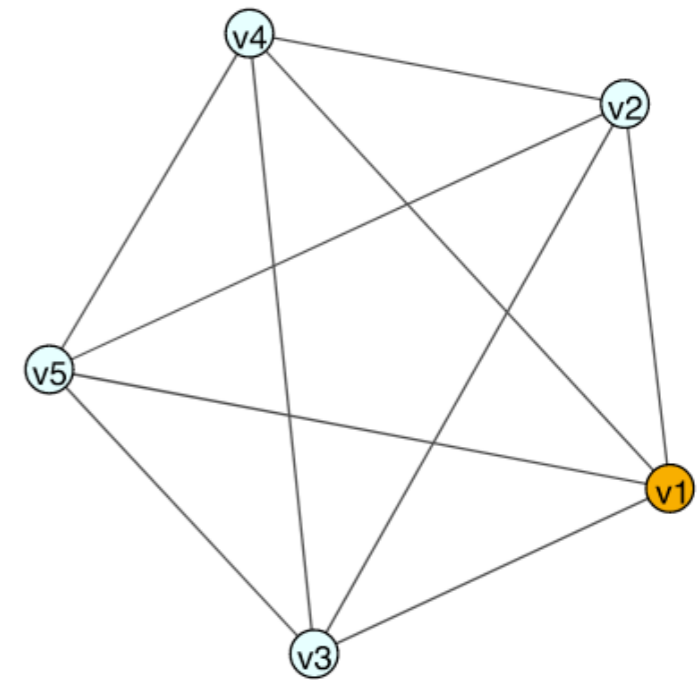
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$C_i = \frac{2 \cdot n}{k \cdot (k - 1)}$  gives the **fraction of possible interconnections** for neighbours of node  $i$

where  $\frac{k \cdot (k - 1)}{2}$  is the maximum number of triangles through a node



$$0 \leq C_i \leq 1$$



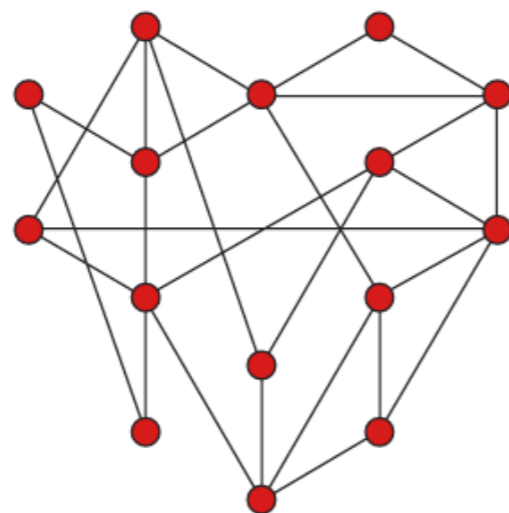
The global clustering coefficient  $C(G)$  is simply the average of its clustering coefficients

# What distinguishes biological networks from random?

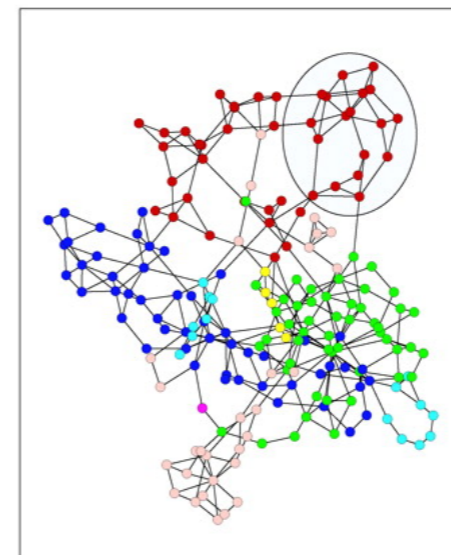
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Do metabolic networks display different network properties from random networks?

**Random network**



**Metabolic network**

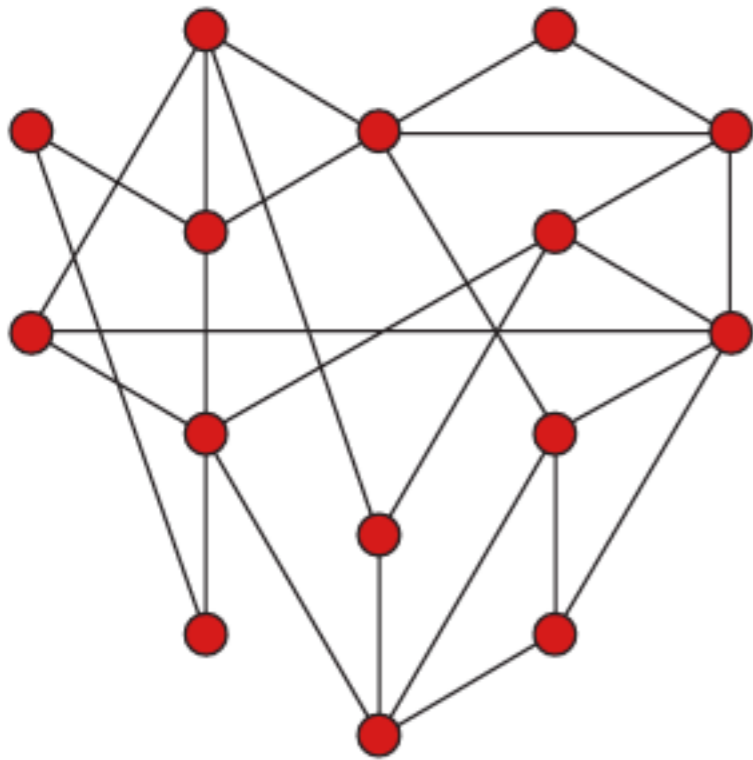


# 7. Degree and clustering coefficient distribution

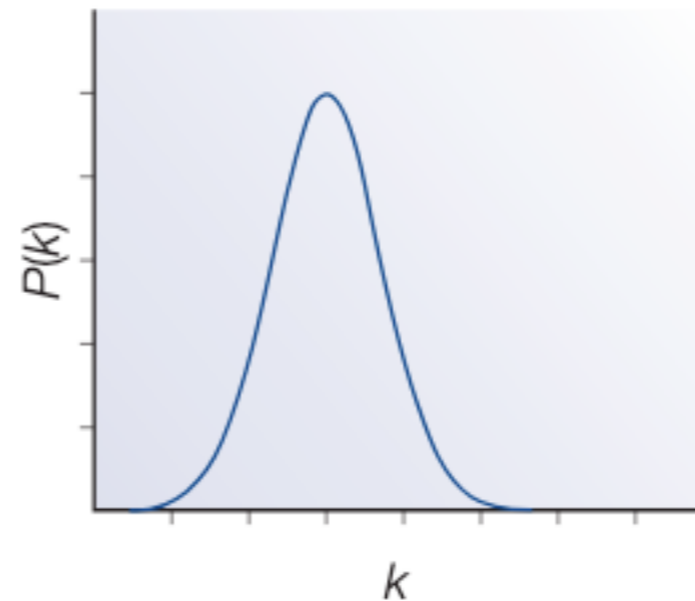
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Degree distributions allow us to compare network organization

**Random network**  
(e.g. Erdős-Rényi model)



**Poisson degree distribution**  
shows no highly connected nodes



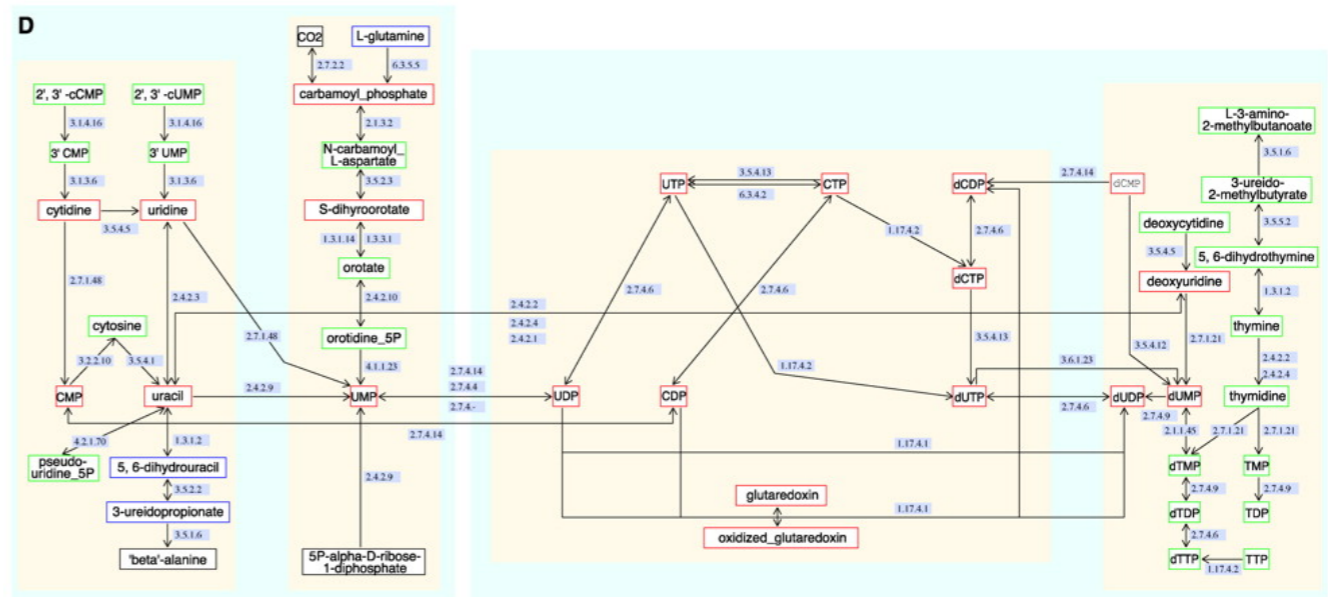
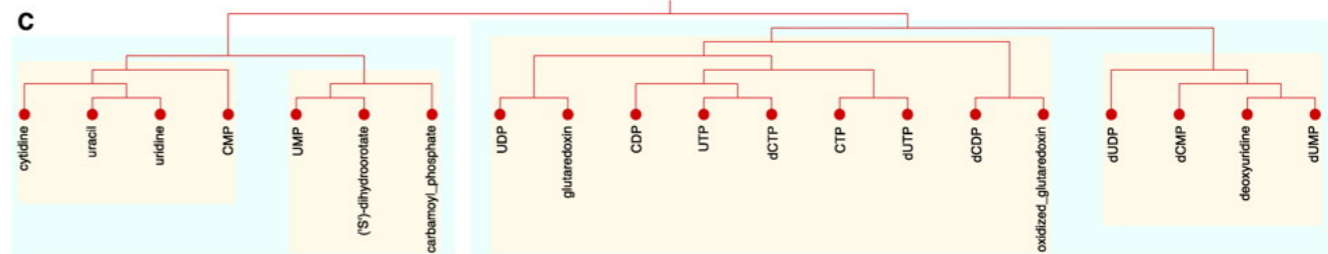
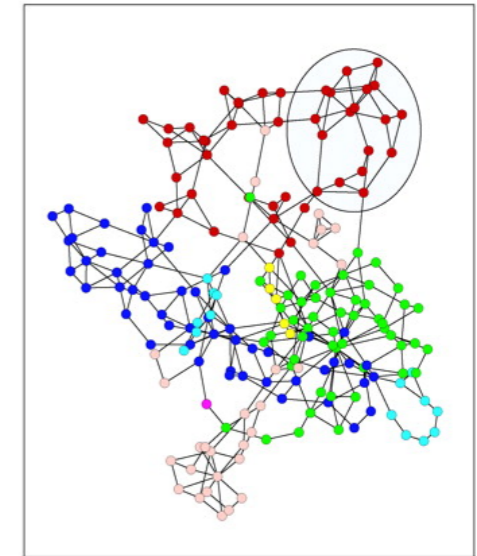
Most nodes have near  $\langle k \rangle$

# Metabolic networks show hierarchical topology

Metabolic networks of 43 organisms are organised into **small, tightly connected modules**

Their combination shows a hierarchical structure

B





# 7. Degree distribution

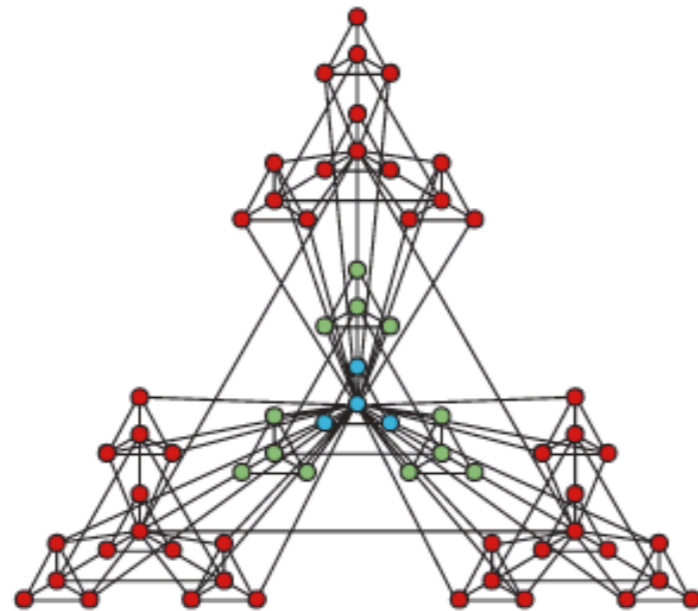
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Biological networks do not follow topology features of random networks.

Analysis of metabolic networks of 43 organisms shows common patterns

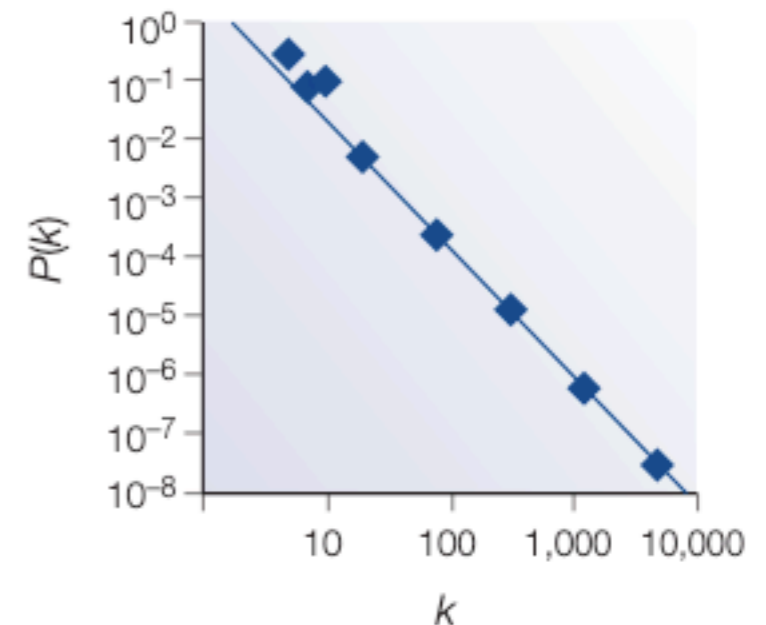
**Biological networks** tend to display high robustness to node failure: removal of <80% nodes still retains paths between any two nodes

**Hierarchical network**



**Degree distribution**

shows many with low degrees  
a few highly connected nodes



# 7. Degree and clustering coefficient distribution

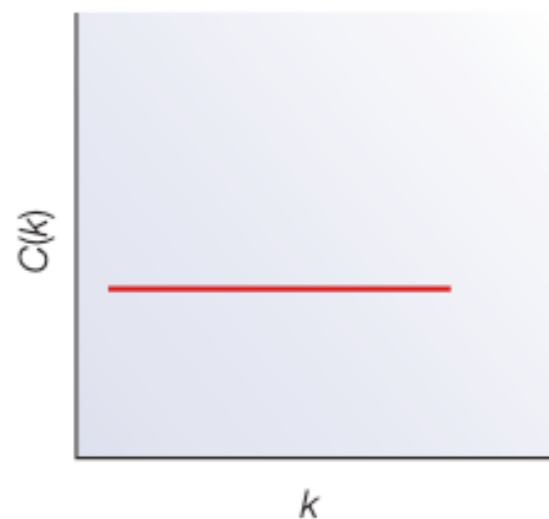
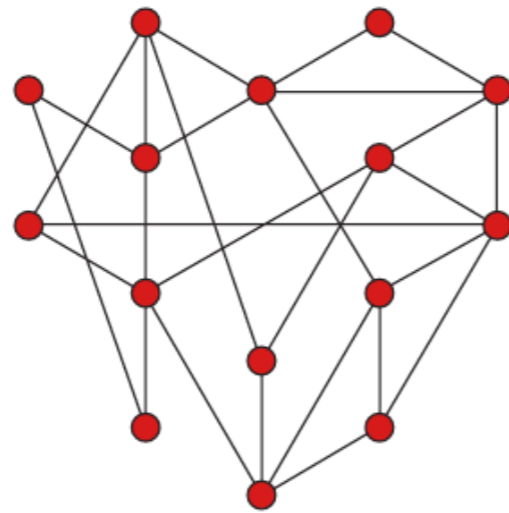
$C(k)$  shows no relationship with  $k$  in random networks: no modular organisation

$C(k) = k^{-1}$  in hierarchical networks

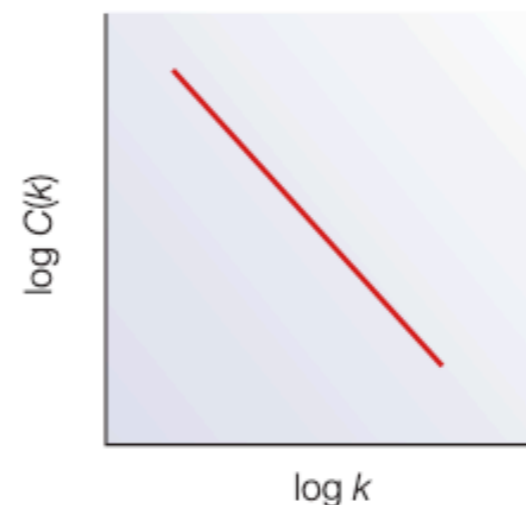
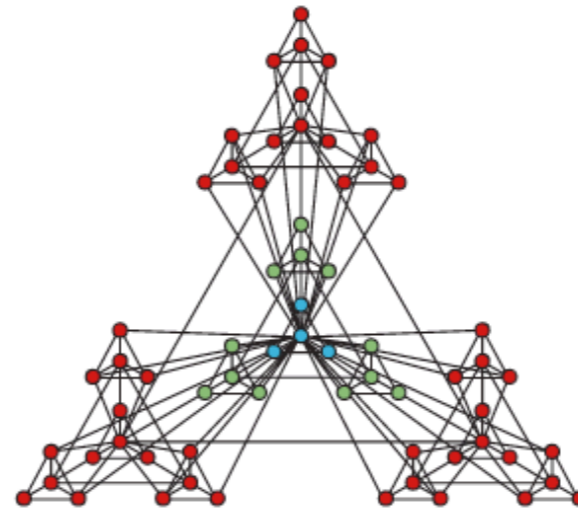
Sparsely connected nodes are part of highly modular areas

Communication between highly clustered neighbourhoods maintained by a few hubs

**Random network**



**Hierarchical network**



# 7. Small world

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Any two nodes can be connected in a small number of steps.

This is a property seen in **random networks** where the mean path length

$$l(G) \approx \log N \text{ for a network of size } N$$

Many biological networks show **ultra-small world** properties:

$$l(G) \approx \log(\log N)$$

Highly central hubs tend **not** to be connected in biological networks:  
they are **disassortative**

(social networks: **assortative**)



# Additional reading

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- [Analysis of Biological Networks](#) - General introduction into biological networks, network notation, and analysis, including graph theory.
- [Using graph theory to analyze biological networks](#) - overview of the usage of graph theory in biological network analysis
- [Survival of the sparsest: robust gene networks are parsimonious](#) - analysis of network complexity and robustness.
- [Network biology: understanding the cell's functional organization](#) - Overview of key concepts in biological network structure
- [Graph Theory and Networks in Biology](#) - extended perspective on how graph analysis is applied in biology
- [Modularity and community structure in networks](#)

Additional references displayed as hyperlinks in each figure.