



## **Quality Control of NGS data**

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```
@HWUSI-EAS100R:6:73:941:1973#0/1
GATTTGGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT
+
!''*(((((***+))%%%++)(%%%%).1***-+*''))**55CCF>>>>CCCCCCC65
```

1<sup>st</sup> row: sequence identifier (machine ID, x-y coordinates, additional info)

2<sup>nd</sup> row: The actual sequence

3<sup>rd</sup> row: starts with "+" and optionally the same identifier as in the 1<sup>st</sup> row

4<sup>th</sup> row: Quality score for each base in read



## **Phred Quality Scores**



## +SEQ\_ID !''\*((((\*\*\*+))%%%++)(%%%%).1\*\*

A quality value Q is an integer representation of the probability *p* that the corresponding base call is incorrect.

$$Q = -10 \log_{10} P$$
  $\longrightarrow$   $P = 10^{\frac{-Q}{10}}$ 

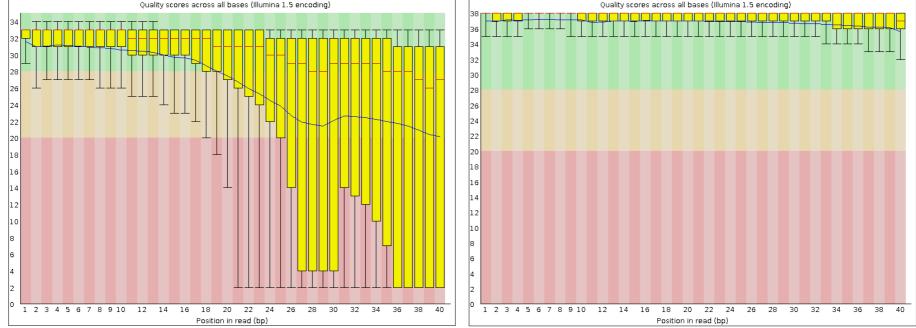
Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90%
20	1 in 100	99%
30	1 in 1000	99.9%
40	1 in 10000	99.99%
50	1 in 100000	99.999%













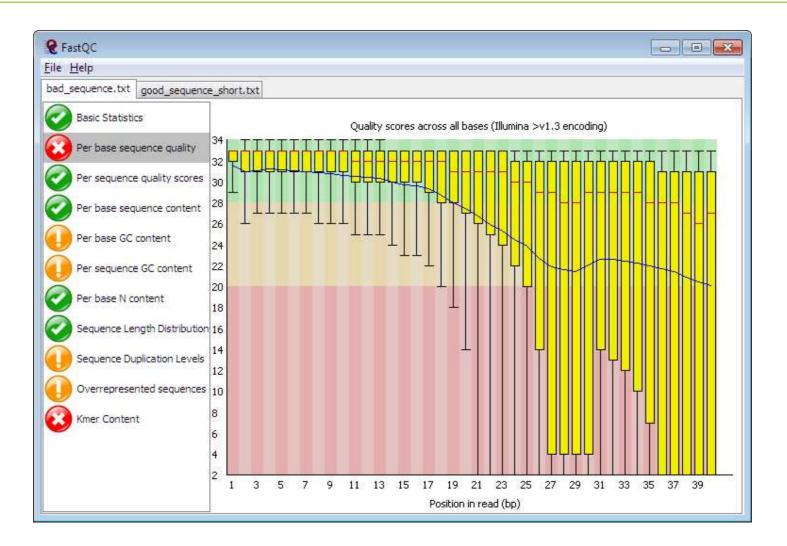


- Different NGS application have their own problem areas and requires their own QC strategy
- Today: Focus on QC for whole genome sequencing
- For variant calling it is important to look at quality score distribution, sequence length distribution and duplication levels.
- Thursday: More details on QC for RNA-seq















## https://www.bioinformatics.babraham.ac.uk/projects/ fastqc/