

Variant-calling Workflow

Malin Larsson

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- Workflows
- Basic variant calling in one sample
- Basic variant calling in cohort
- Introduction to exercise

In separate talk Thursday at 9:

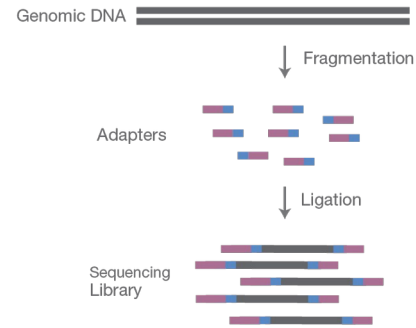
- GATK's Best practices



Illumina Sequencing

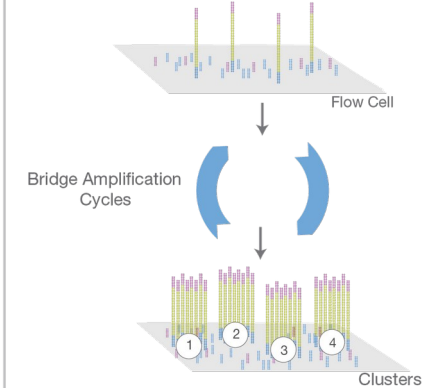
<https://www.youtube.com/watch?v=fCd6B5HRaZ8>

A. Library Preparation



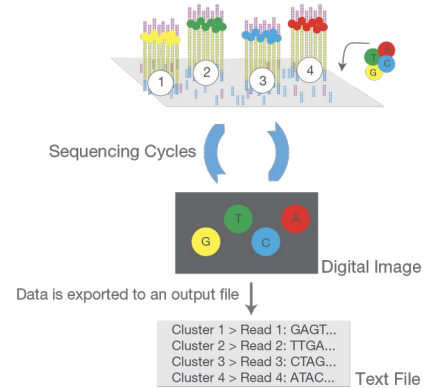
NGS library is prepared by fragmenting a gDNA sample and ligating specialized adapters to both fragment ends.

B. Cluster Amplification



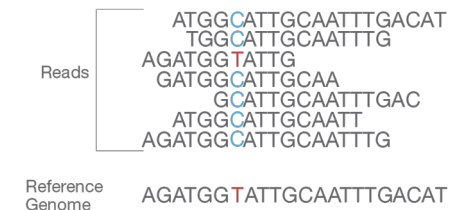
Library is loaded into a flow cell and the fragments are hybridized to the flow cell surface. Each bound fragment is amplified into a clonal cluster through bridge amplification.

C. Sequencing



Sequencing reagents, including fluorescently labeled nucleotides, are added and the first base is incorporated. The flow cell is imaged and the emission from each cluster is recorded. The emission wavelength and intensity are used to identify the base. This cycle is repeated "n" times to create a read length of "n" bases.

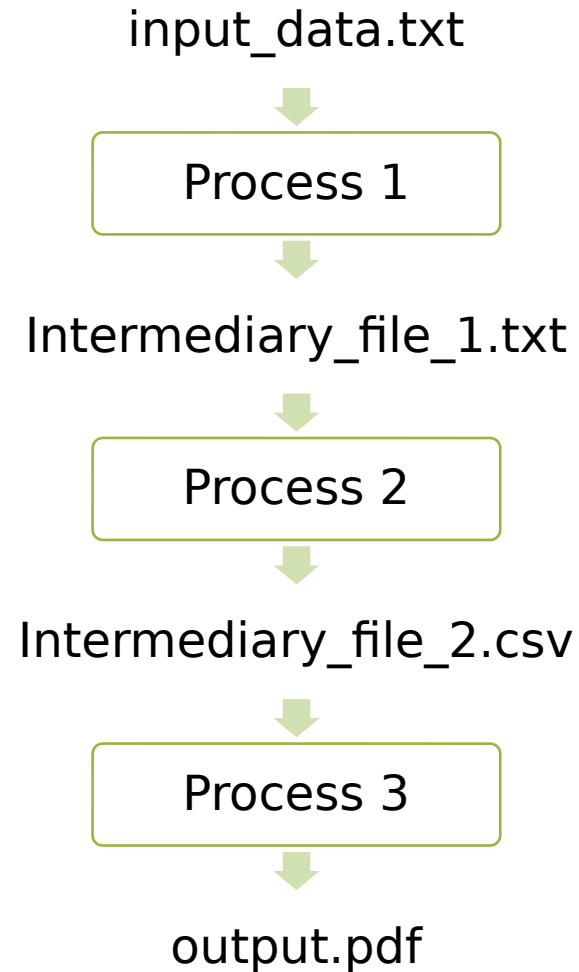
D. Alignment and Data Analysis



Reads are aligned to a reference sequence with bioinformatics software. After alignment, differences between the reference genome and the newly sequenced reads can be identified.

Workflows

What is a workflow



Workflow conventions



- Create a new output file in each process – don't over write the input file
- Use informative file names
- Include information of the process in output file name

Example: Basic variant calling in one sample



HG00097_1.fa
stq
HG00097_2.fa
stq

FASTQ files

Alignment

HG00097.ba
m

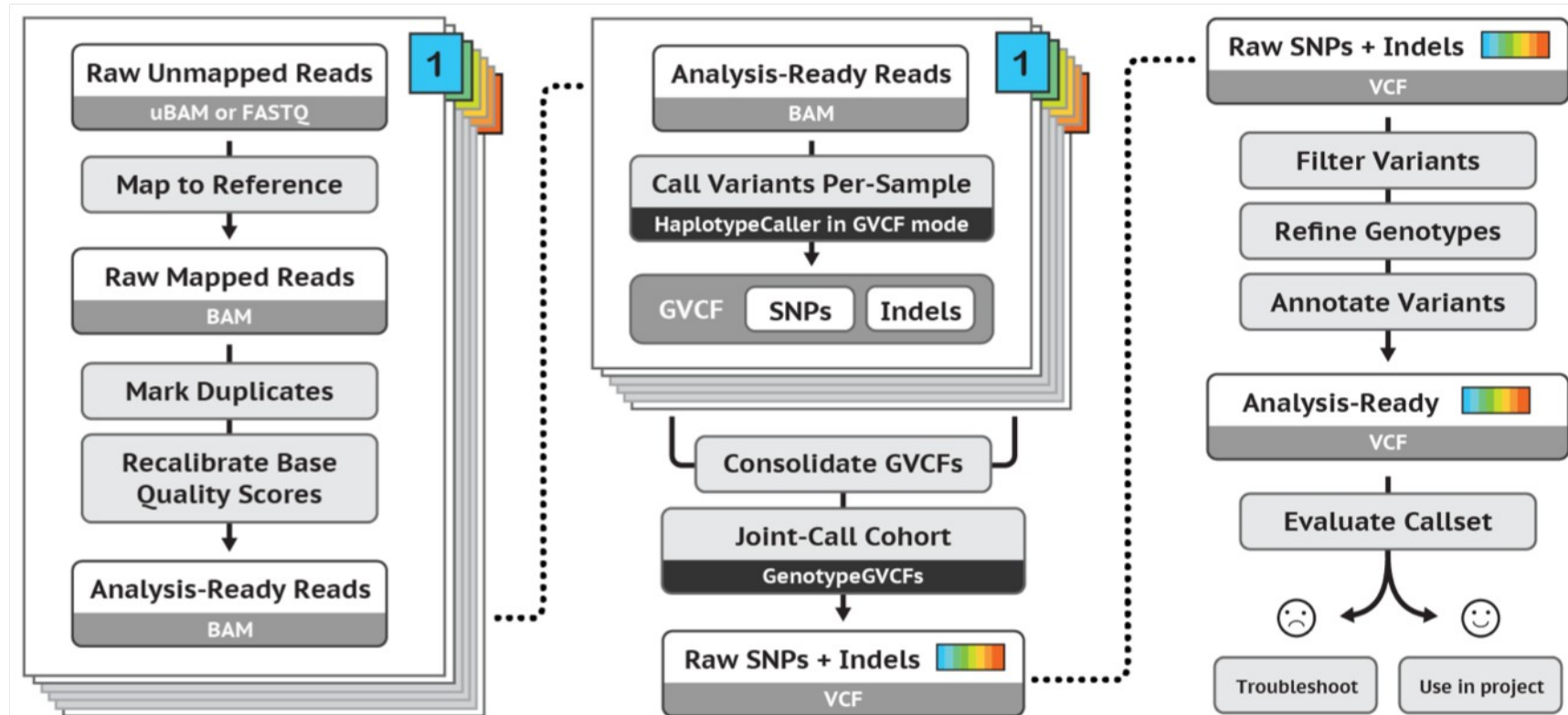
BAM files

VariantCalling

HG00097.vcf

VCF files





Basic Variant Calling in one sample

Alignment



HG00097_1.fa
stq
HG00097_2.fa
stq

FASTQ files

BWA mem

HG00097.ba
m

BAM files

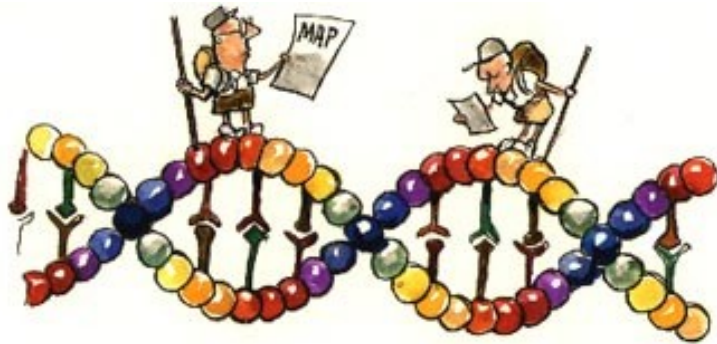
HaplotypeCall
er

HG00097.vcf

VCF files



The reference genome



A reference genome is a haploid nucleic acid sequence which represents a species genome.

The first draft of the human genome contained 150,000 gaps.

GRCh37: 250 gaps

GRCh38 is the latest version of the human reference genome, but we will work with GRCh37 in the lab.

Keep track of the reference version!



The reference genome sequence is used as input in many bioinformatics applications for NGS data:

- mapping
- variant calling
- annotation

You must keep track of which version of the reference genome your data was mapped to.

The same version must be used in all downstream analyses.



- Most large files we work with, such as the reference genome, need an index
- Allows efficient access to the file
- Different indices for different file-types
- Bwa index = Burrows-Wheeler transform of reference genome (several files)
- Needs index: fasta, bam vcf files

Burrows-Wheeler Aligner



<http://bio-bwa.sourceforge.net>

Burrows-Wheeler Aligner

[Home](#)

Introduction

BWA is a software package for mapping low-divergent sequences against a large reference genome, such as the human genome. It consists of three algorithms: BWA-backtrack, BWA-SW and BWA-MEM. The first algorithm is designed for Illumina sequence reads up to 100bp, while the rest two for longer sequences ranging from 70bp to 1Mbp. BWA-MEM and BWA-SW share similar features for long-read support and split alignment, but BWA-MEM, which is the generally recommended for high-quality queries as it is faster and more accurate. BWA-MEM also has better performance than BWA-backtrack for 70bp reads.

FAQ

How can I cite BWA?

The short read alignment component (bwa-short) has been published by Li H. and Durbin R. (2009) Fast and accurate short read alignment with Burrows-Wheeler Transform. *Bioinformatics*, 25:1754-60. [PMID: 19451168]

If you use BWA-SW, please cite:
 Li H. and Durbin R. (2010) Fast and accurate long-read alignment with Burrows-Wheeler Transform. *Bioinformatics*, Epub. [PMID: 20111653]

(See also Errata below for a minor correction to the formulae in the above papers.)

There are three algorithms, which one should I choose?

For 70bp or longer Illumina, 454, Ion Torrent and Sanger reads, contigs and BAC sequences, BWA-MEM is usually the preferred algorithm. For short sequences, BWA-backtrack may be better. BWA-SW may be used for long reads.

BWA:

[SF project page](#)

Burrows-Wheeler transform of reference genome

0: googol\$	String Sorting →	0 6: \$googo l
1: oogol\$g		1 3: gol\$go o
2: ogol\$go		2 0: googol \$
3: gol\$goo		3 5: l\$goog o
4: ol\$goog		4 2: ogol\$g o
5: l\$googo		5 4: ol\$goo g
6: \$googol		6 1: oogol\$ g

Pos	i	S(i)		B[i]
	↓	↓		↓
X = googol\$		lo\$oogg		
		(6,3,0,5,2,4,1)		



module load bwa





HEADER SECTION

```
@HD      VN:1.6      SO:coordinate
@SQ      SN:2      LN:243199373
@PG      ID:bwa     PN:bwa     VN:0.7.17-r1188      CL:bwa mem -t 1 human_g1k_v37_chr2.fasta
HG00097_1.fq HG00097_2.fq
@PG      ID:samtools PN:samtools PP:bwa     VN:1.10      CL:samtools sort
@PG      ID:samtools.1PN:samtools PP:samtools VN:1.10      CL:samtools view -H HG00097.bam
```

ALIGNMENT SECTION

```
Read_001      99      2      3843448      0      101M      =      3843625      278
TTGGTTCCATATGAACTTT
Read_001      147      2      3843625      0      101M      =      3843448      -278
TTATTTTCATTGAGCAGTGGT
Read_002      163      2      4210055      0      101M      =      4210377      423
TGGTACCAAAACAGAGATAT
Read_003      99      2      4210066      0      101M      =      4210317      352
CAGAGATATAGATCAATGGGA
0IIFFFFIFFFIFIFIIIIIF
```

↑
 Read name
 (usually more complicated)

↑
 Reference sequence name

↑
 Start position

↑
 Sequence

↑
 Quality

Convert to Bam



Bam file is a binary representation of the Sam file



- Link *sample id, library prep, flowcell* and *sequencing run* to the reads.
- Good for error tracking!
- Often needed for variant calling
- Detailed description at <https://gatkforums.broadinstitute.org/gatk/discussion/6472/read-groups>

RGID = *combination of the sample id and run id*

RGLB = Library prep

RGPL = Platform (for us ILLUMINA)

RGPU = Run identifier *usually barcode of flowcell*

RGSM = Sample name

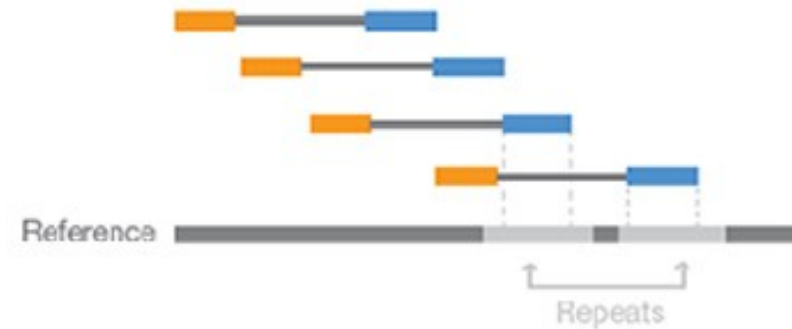
Paired-End data



Paired-End Reads



Alignment to the Reference Sequence



Paired-end sequencing enables both ends of the DNA fragment to be sequenced. Because the distance between each paired read is known, alignment algorithms can use this information to map the reads over repetitive regions more precisely. This results in much better alignment of the reads, especially across difficult-to-sequence, repetitive regions of the genome.

Paired-end data



ID_R1_001.fastq

```
@HISEQ:100:C3MG8ACXX:5:1101:1160:2  
197 1:N:0:ATCACG  
CAGTTGCGATGAGAGCGTTGAGAAGTATAATAGG  
AGTTAAACTGAGTAACAGGATAAGAAATAGTGAG  
ATATGGAAACGTTGTGGTCTGAAAGAAGATGT  
+  
B@CFFFFFFHHHHHGJJJJJJJJJJJFHHIIIIJJ  
JIHGIJJJJJIJJIJJJJIIJJJJJIIIEIHJIJ  
HGHHHHHDFEFEDDDDDCDDDCDDDDDDDCDC
```

ID_R2_001.fastq

```
@HISEQ:100:C3MG8ACXX:5:1101:1160:  
2197 2:N:0:ATCACG  
CTTCGTCCACTTTCATTATTCCTTTCATACATG  
CTCTCCGGTTTAGGGTACTCTTGACCTGGCCTT  
TTTTCAAGACGTCCCTGACTTGATCTTGAAACG  
+  
CCFFFFFFHHHHHJJJJIJJJJJJJJJJJJJJ  
JJJJJJJIJJIJGIJHBGHHIIIIJIIJJJJJJJI  
JJJHFFFFFFDDDDDDDDDDDDDDDEDCCDDDD
```

Variant calling



HG00097_1.fa
stq
HG00097_2.fa
stq

FASTQ files

BWA mem

HG00097.ba
m

BAM files

HaplotypeCall
er

HG00097.vcf

VCF files



Genetic variation



Genetic variation = differences in DNA among individuals of the same species

Detecting variants in reads



Reference:

...GTGCGTAGACTGCTAGATCGAAGA...

Sample:

...GTGCGTAGACTG**A**TAGATCGAAGA...

...GTGCGTAGACTG**A**TAGATCGAAGA...

...GTGCGTAGACTGCTAGATCGAAGA...

...GTGCGTAGACTGCTAGATCGAAGA...

...GTGCGTAGACTG**A**TAGATCGAAGA...

...GTGCGTAGACTG**A**TAGATCGAAGA...

...GTGCGTAGACTGCTAGATCGAAGA...

...GTGCGTAGACTG**A**TAGATCGAAGA...

...GTGCGTAGACTGCTAGATCGAAGA...

...GTGCGTAGACTG**A**TAGATCGAAGA...



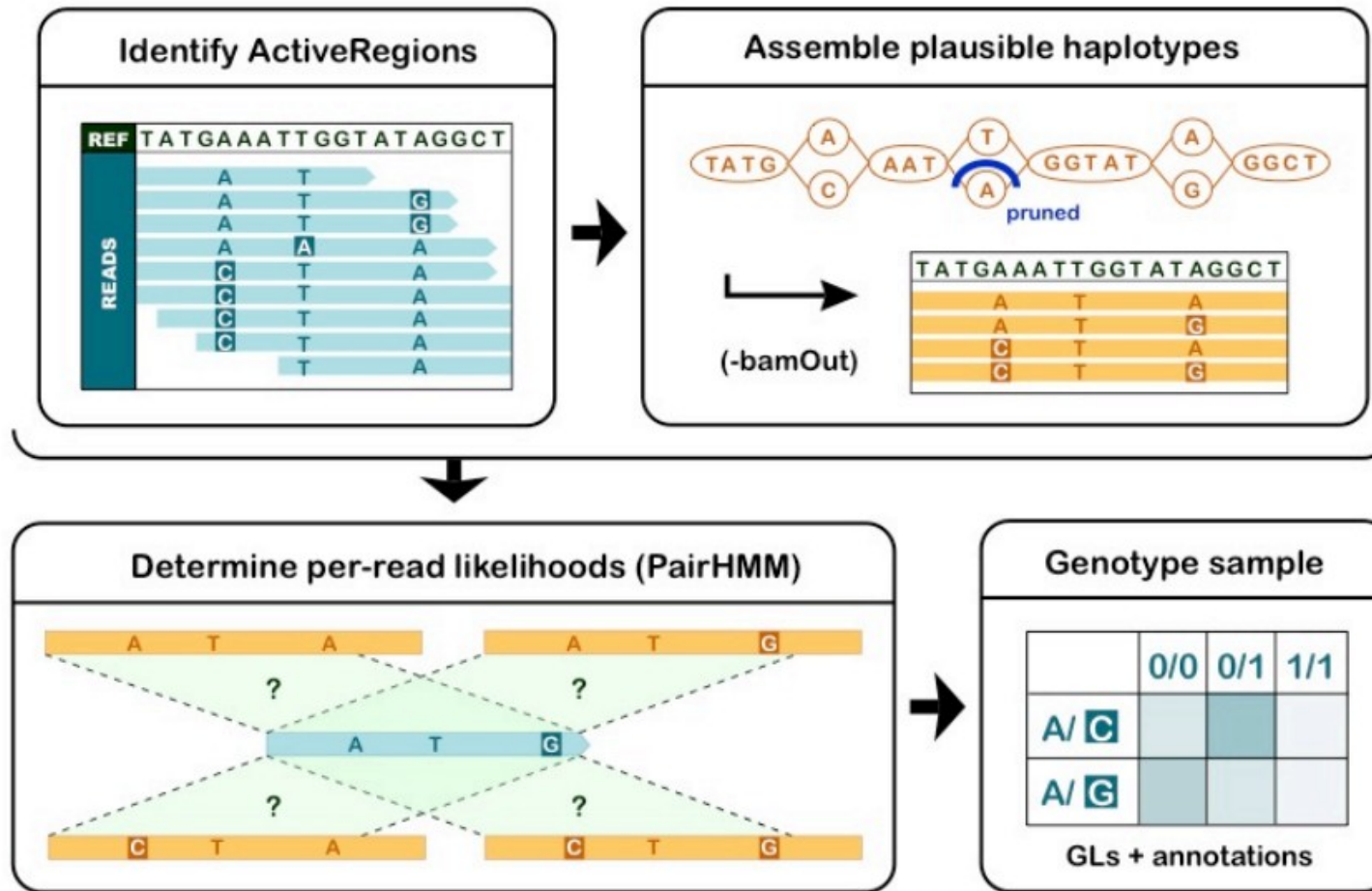
TGGGCTTTTCCAACAGGTATATCTTCCCCGCTAGCTAGCTAGCTACTTCAAATTCCT

Reference allele	AGCTAGCTA
Alternative allele	AGCTGGCTA

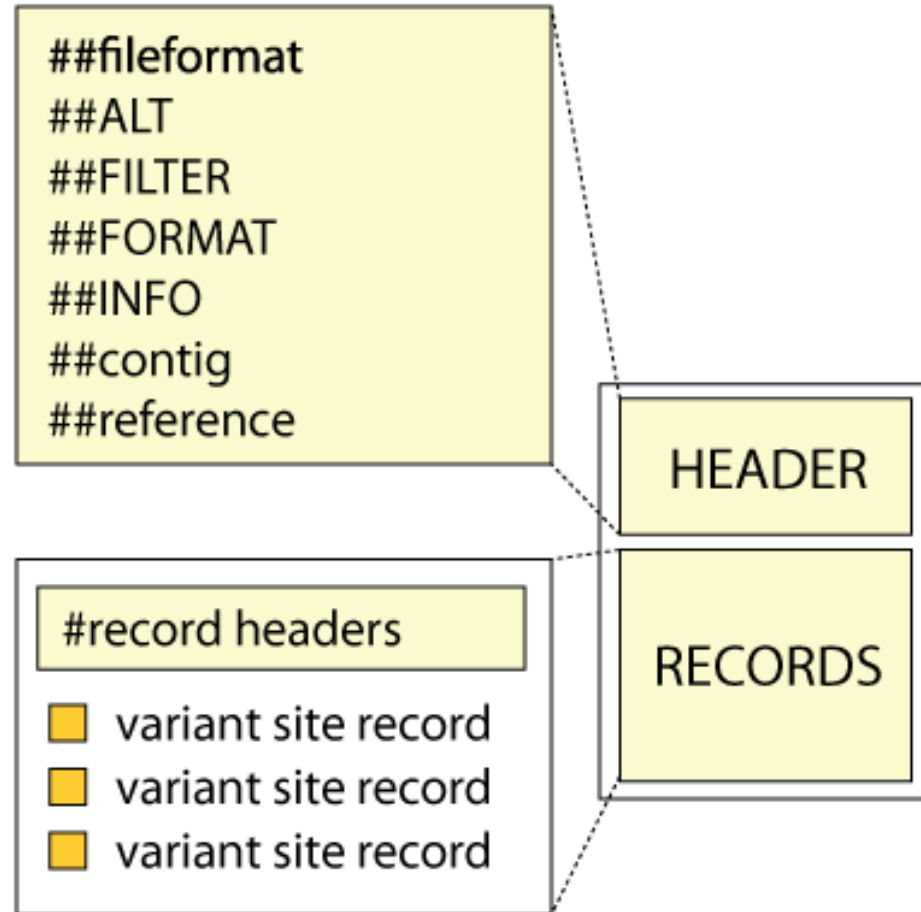
Reference allele = the allele in the reference genome

Alternative allele = the allele NOT in the reference genome

Variant Calling HaplotypeCaller



Variant Call Format (VCF)



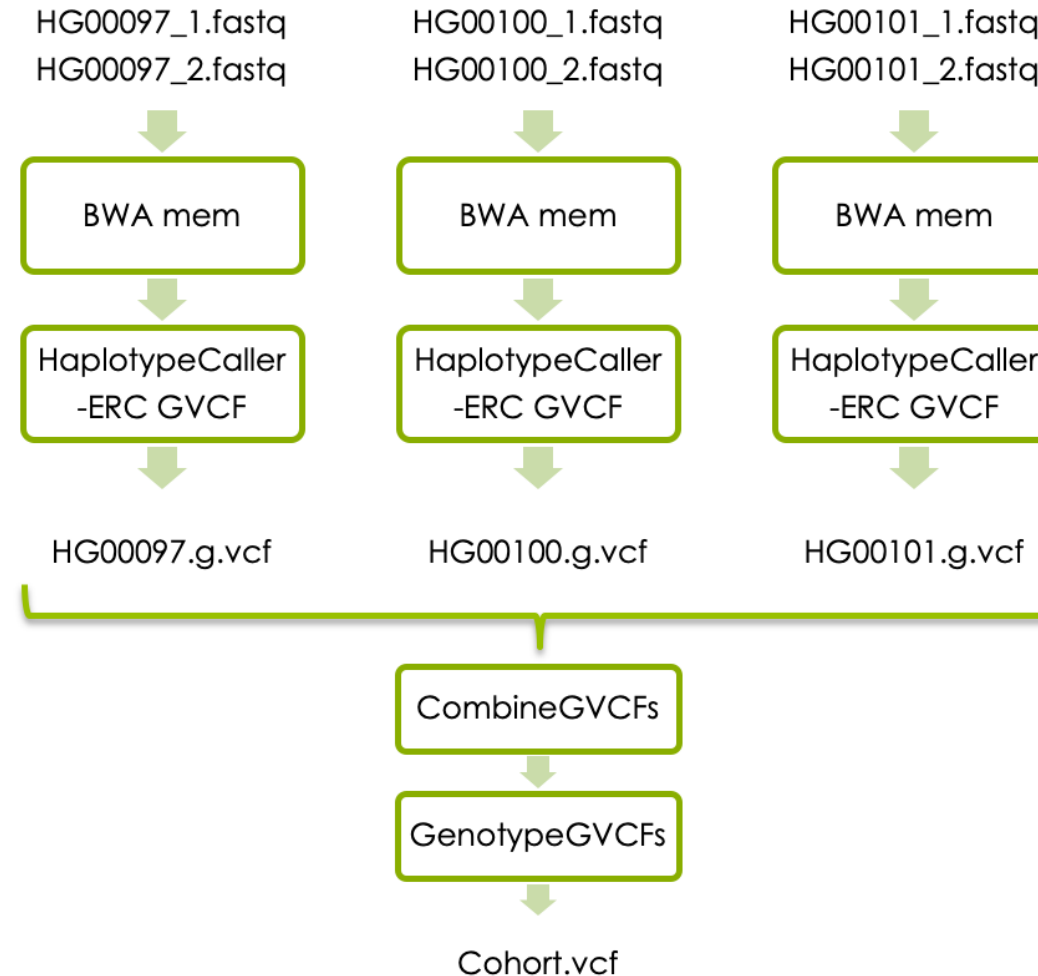
Variant Call Format (VCF)



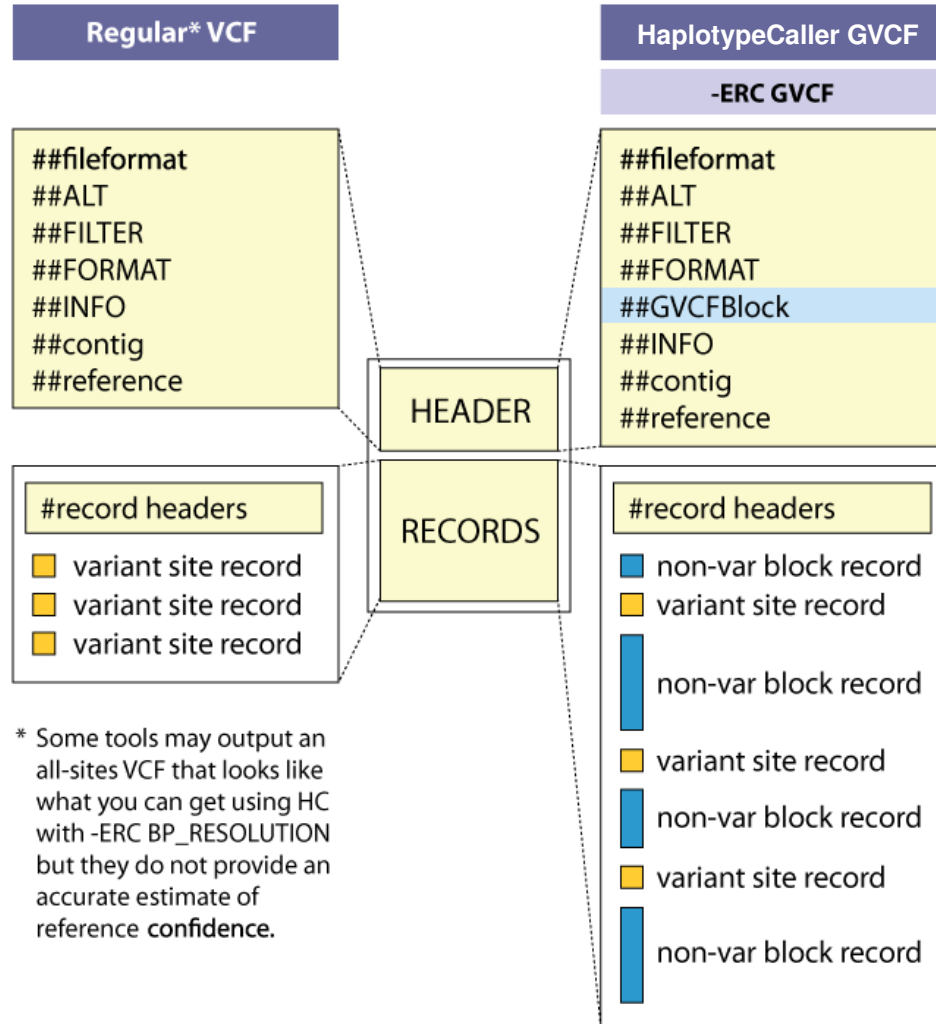
```
##fileformat=VCFv4.2
##FILTER=<ID=LowQual,Description="Low quality">
##FORMAT=<ID=AD,Number=R,Type=Integer,Description="Allelic depths for the ref and alt alleles in the order listed">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Approximate read depth (reads with MQ=255 or with bad mates are filtered)">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes, for each ALT allele, in the same order as listed">
##INFO=<ID=AF,Number=A,Type=Float,Description="Allele Frequency, for each ALT allele, in the same order as listed">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##contig=<ID=2,length=243199373>
##source=HaplotypeCaller
#CHROM  POS          ID          REF     ALT     QUAL     FILTER     INFO          FORMAT      HG00097
2       136220992    .           G       GT      30.64    .          AC=1;AF=0.500;AN=2  GT:AD:DP    0/1:3,2:5
2       136226814    .           GAC     G       44.60    .          AC=1;AF=0.500;AN=2  GT:AD:DP    0/1:4,2:6
2       136234279    .           C       T       102.60   .          AC=1;AF=0.500;AN=2  GT:AD:DP    0/1:3,4:7
2       136234284    .           C       T       102.60   .          AC=1;AF=0.500;AN=2  GT:AD:DP    0/1:3,4:7
2       136263277    .           T       A       148.60   .          AC=1;AF=0.500;AN=2  GT:AD:DP    0/1:8,5:13
...
...
```

Variant calling in cohort

Basic variant calling in cohort

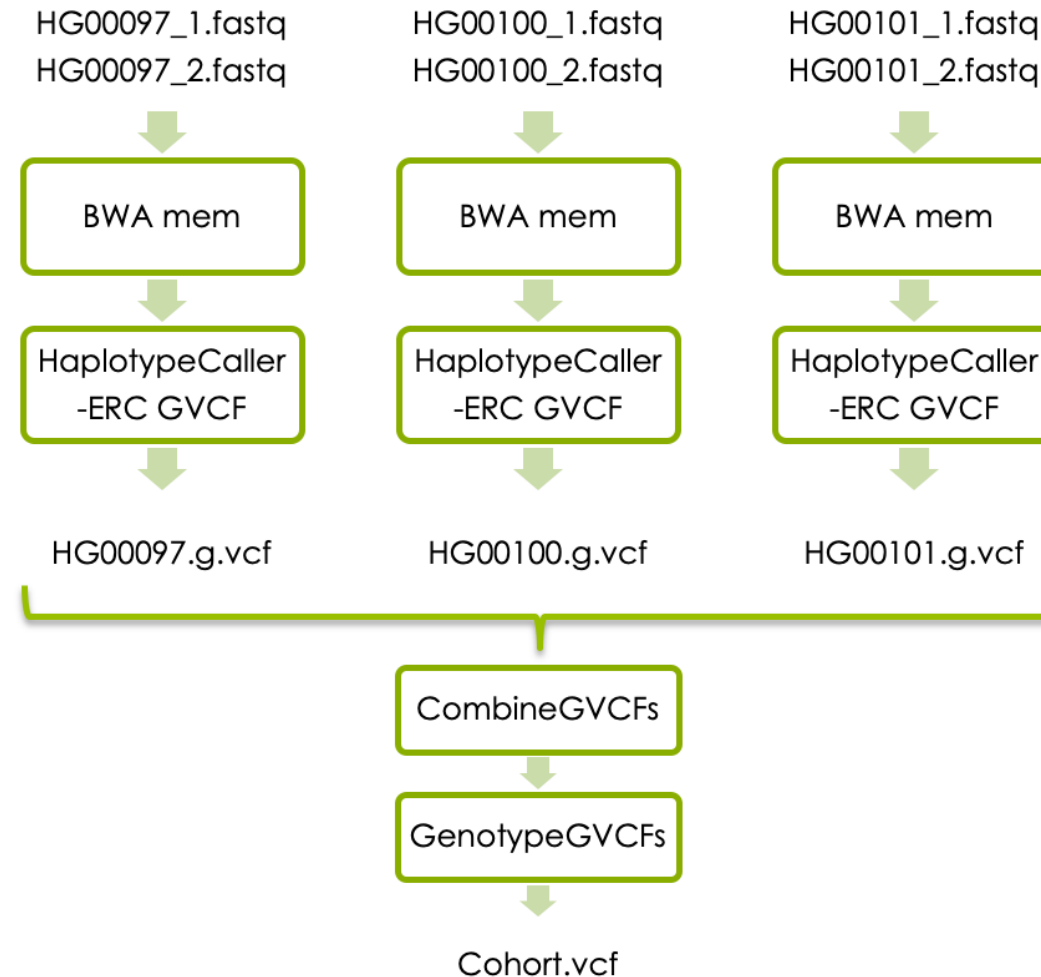


GVCF Files are valid VCFs with extra information



- GVCF has records for all sites, whether there is a variant call there or not.
- The records include an accurate estimation of how confident we are in the determination that the sites are homozygous-reference or not.
- Adjacent non-variant sites merged into blocks

Basic variant calling in cohort



Variant Call Format (VCF)



```
##fileformat=VCFv4.2
##ALT=<ID=NON_REF,Description="Represents any possible alternative allele at this location">
##FILTER=<ID=LowQual,Description="Low quality">
##FORMAT=<ID=AD,Number=R,Type=Integer,Description="Allelic depths for the ref and alt alleles in the order listed">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Approximate read depth (reads with MQ=255 or with bad mates are filtered)">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##INFO=<ID=AC,Number=A,Type=Integer,Description="Allele count in genotypes, for each ALT allele, in the same order as listed">
##INFO=<ID=AF,Number=A,Type=Float,Description="Allele Frequency, for each ALT allele, in the same order as listed">
##INFO=<ID=AN,Number=1,Type=Integer,Description="Total number of alleles in called genotypes">
##contig=<ID=2,length=243199373>
##source=CombineGVCFs
##source=GenotypeGVCFs
##source=HaplotypeCaller
```

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	HG00097	HG00100	HG00101
2	136045826	.	G	A	167.26	.	AC=1;AF=0.167;AN=6	GT:AD:DP	0/0:8,0:8	0/0:13,0:13	0/1:1,5:6
2	136046443	.	CGT	C	129.27	.	AC=3;AF=0.500;AN=6	GT:AD:DP	0/0:8,0:8	0/1:3,1:4	1/1:0,4:4
2	136047387	.	T	C	186.27	.	AC=1;AF=0.167;AN=6	GT:AD:DP	0/0:6,0:6	0/0:16,0:16	0/1:4,6:10
2	136048649	.	C	G	127.26	.	AC=1;AF=0.167;AN=6	GT:AD:DP	0/0:13,0:13	0/0:9,0:9	0/1:1,4:5
2	136052318	.	C	T	107.26	.	AC=1;AF=0.167;AN=6	GT:AD:DP	0/0:7,0:7	0/0:13,0:13	0/1:3,3:6

GATK's best practices for germline short variant discovery



Genome Analysis Toolkit

Variant Discovery in High-Throughput Sequencing Data



Developed in the Data Sciences Platform at the [Broad Institute](#), the toolkit offers a wide variety of tools with a primary focus on variant discovery and genotyping. Its powerful processing engine and high-performance computing features make it capable of taking on projects of any size. [Learn more](#)

Find answers to your questions. Stay up to date on the latest topics. Ask questions and help others.



Getting Started

Best practices, tutorials, and other info to get you started



Technical Documentation

Algorithms, glossary, and other detailed resources



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Purpose, usage and options for each tool



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GATK Showcase on Terra

Check out these fully configured workspaces



DRAGEN-GATK

Learn more about DRAGEN-GATK

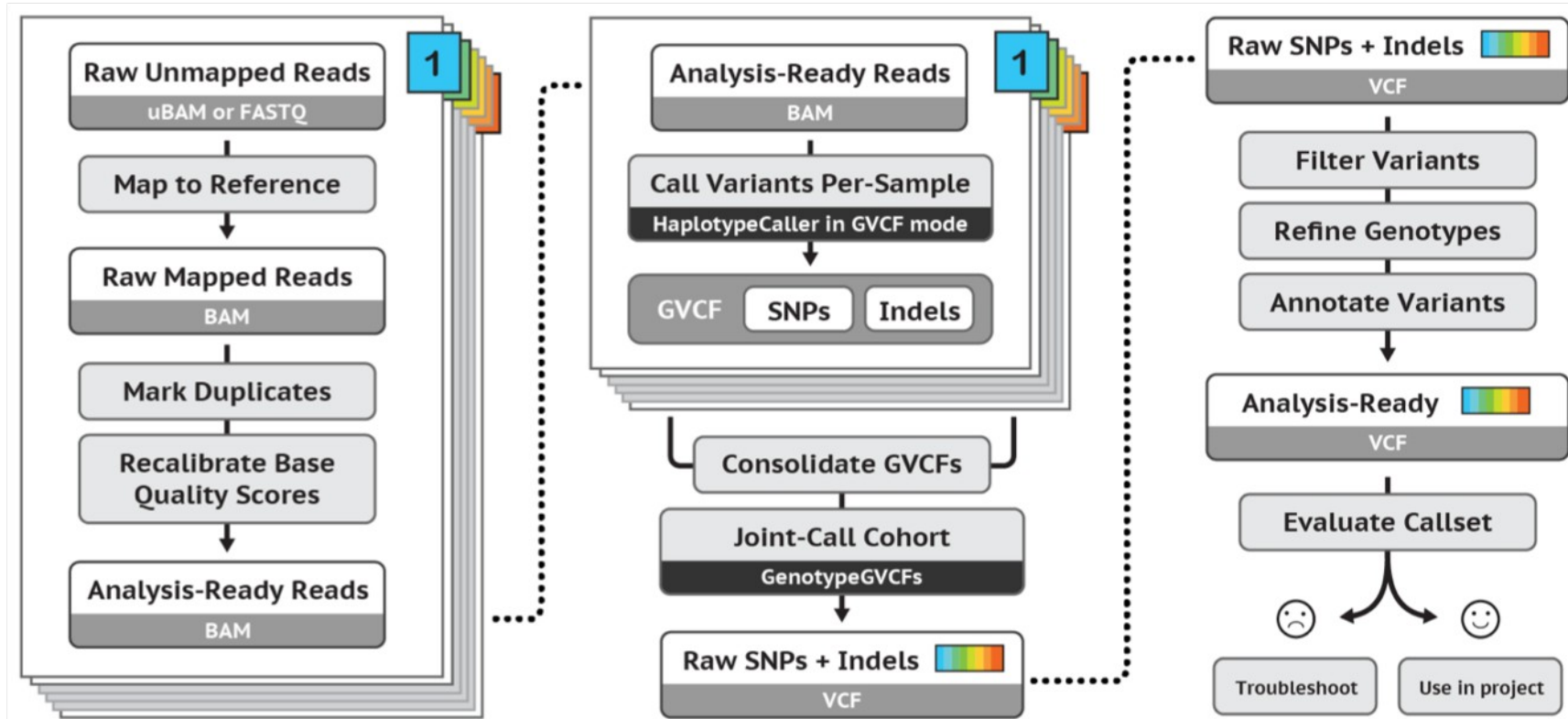


Download latest version of GATK

The GATK package download includes all released GATK tools

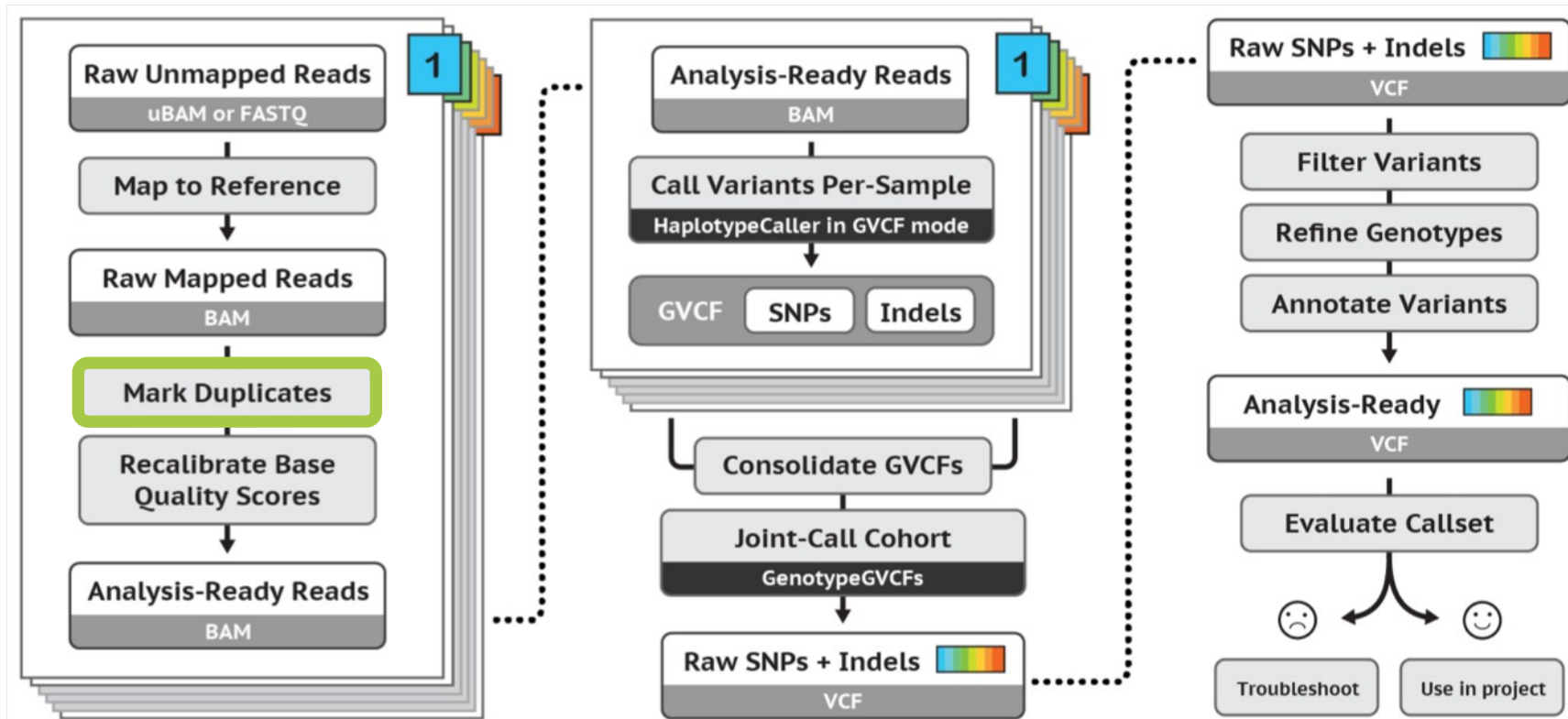
Run on Cloud

Run on HPC





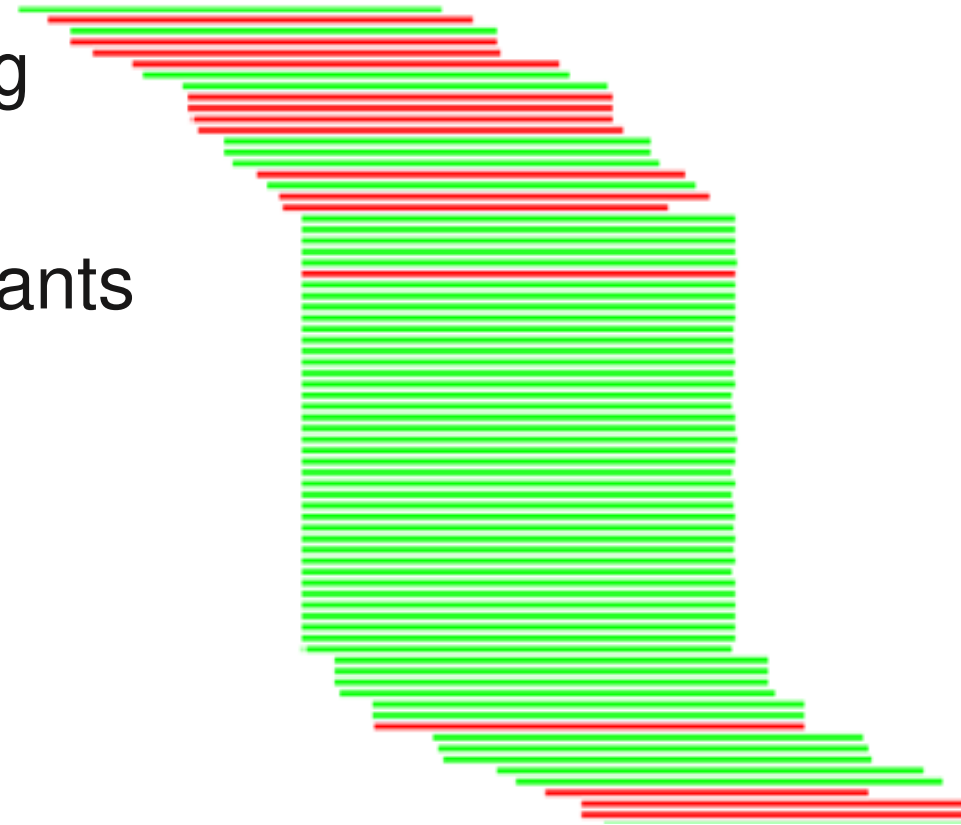
Mark Duplicates



Duplicate reads



- PCR duplicates - library preparation
- Optical duplicates - sequencing
- Don't add unique information
- Gives false allelic ratios of variants
- Should be removed/marked





Need Help?

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MarkDuplicates



[GATK](#) / [Tool Index](#) / 4.0.1.1

MarkDuplicates (Picard)

Follow



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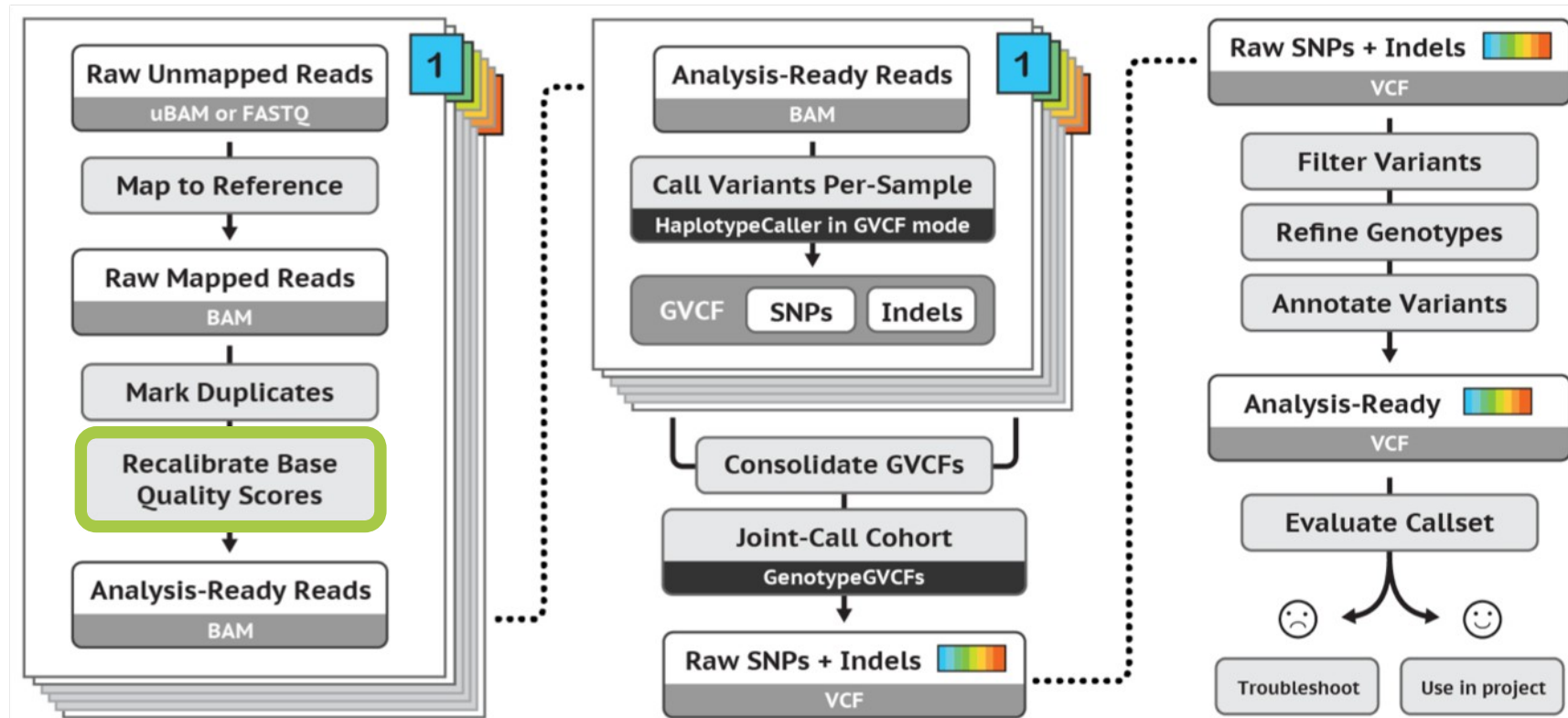
10 months ago · Updated

Identifies duplicate reads.

This tool locates and tags duplicate reads in a BAM or SAM file, where duplicate reads are defined as originating from a single fragment of DNA. Duplicates can arise during sample preparation e.g. library construction using PCR. See also [EstimateLibraryComplexity](#) for additional notes on PCR duplication artifacts. Duplicate reads can also result from a single amplification cluster, incorrectly detected as multiple clusters by the optical sensor of the sequencing instrument. These duplication artifacts are referred to as optical duplicates.

```
gatk --java-options -Xmx7g MarkDuplicates \  
  -I input.bam \  
  -O marked_duplicates.bam \  
  -M marked_dup_metrics.txt
```

Base Quality Score Recalibration (BQSR)





- During base calling, the sequencer estimates a quality score for each base. This is the quality scores present in the fastq files.
- Systematic (non-random) errors in the base quality score estimation can occur.
 - due to the physics or chemistry of the sequencing reaction
 - manufacturing flaws in the equipment
 - etc
- Can cause bias in variant calling
- **Base Quality Score Recalibration** helps to calibrate the scores so that they correspond to the real per-base sequencing error rate (phred scores)



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Base Quality Score Recalibration (BQSR) [Follow](#)



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5 days ago · Updated

BQSR stands for Base Quality Score Recalibration. In a nutshell, it is a data pre-processing step that detects systematic errors made by the sequencing machine when it estimates the accuracy of each base call.

*Note that this **base** recalibration process (BQSR) should not be confused with **variant** recalibration (VQSR), which is a sophisticated filtering technique applied on the variant callset produced in a later step. The developers who named these methods wish to apologize sincerely to anyone, especially Spanish-speaking users, who get tripped up by the similarity of these names.*

Contents

1. Overview
2. Base recalibration procedure details
3. Important factors for successful recalibration
4. Examples of pre- and post-recalibration metrics
5. Recalibration report

Articles in

[ActiveRe \(Haploty\)](#)

[Evaluatir and varia Mutect2\)](#)

[Local re-: determin Mutect2\)](#)

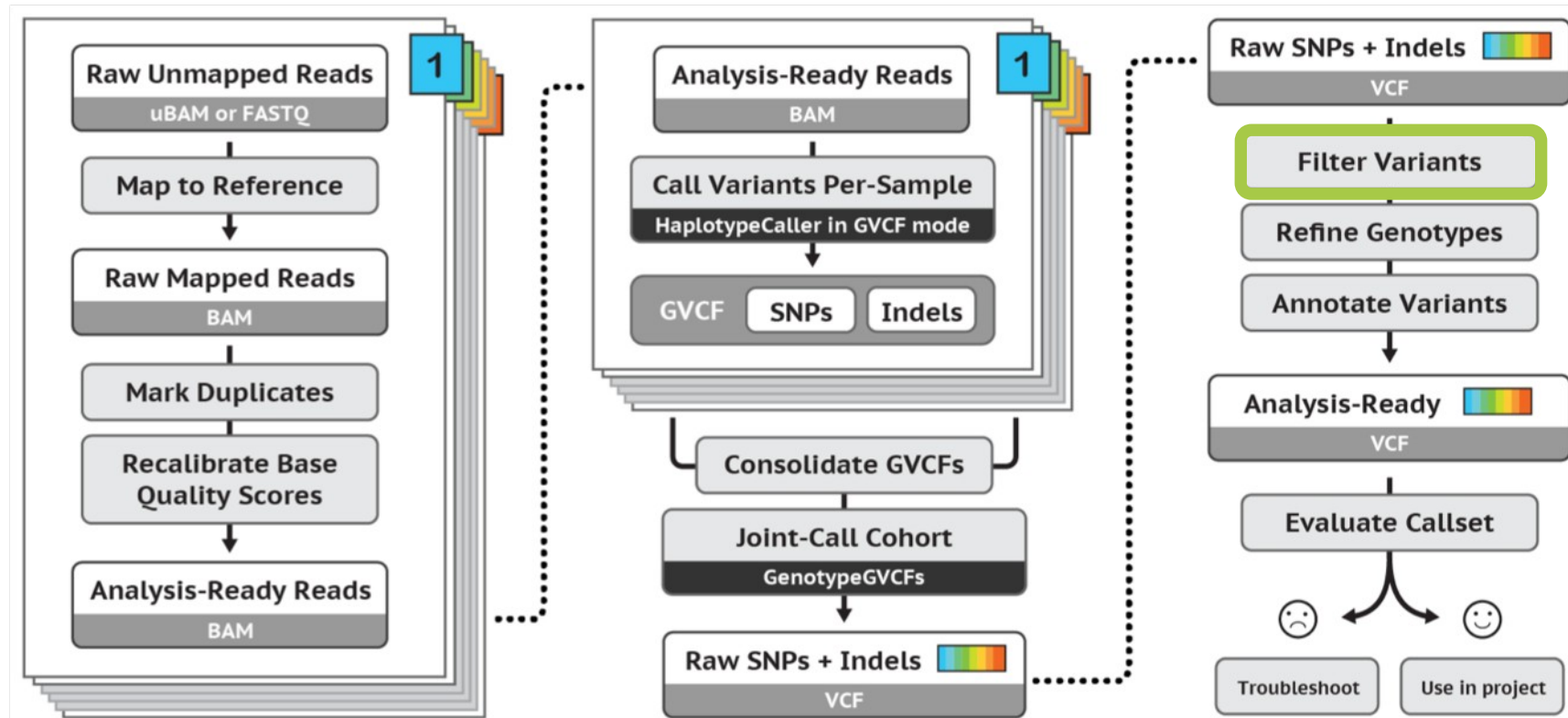
[Allele-sp: germline](#)

[Variant C](#)

[Evaluatir variant c:](#)



Filter variants

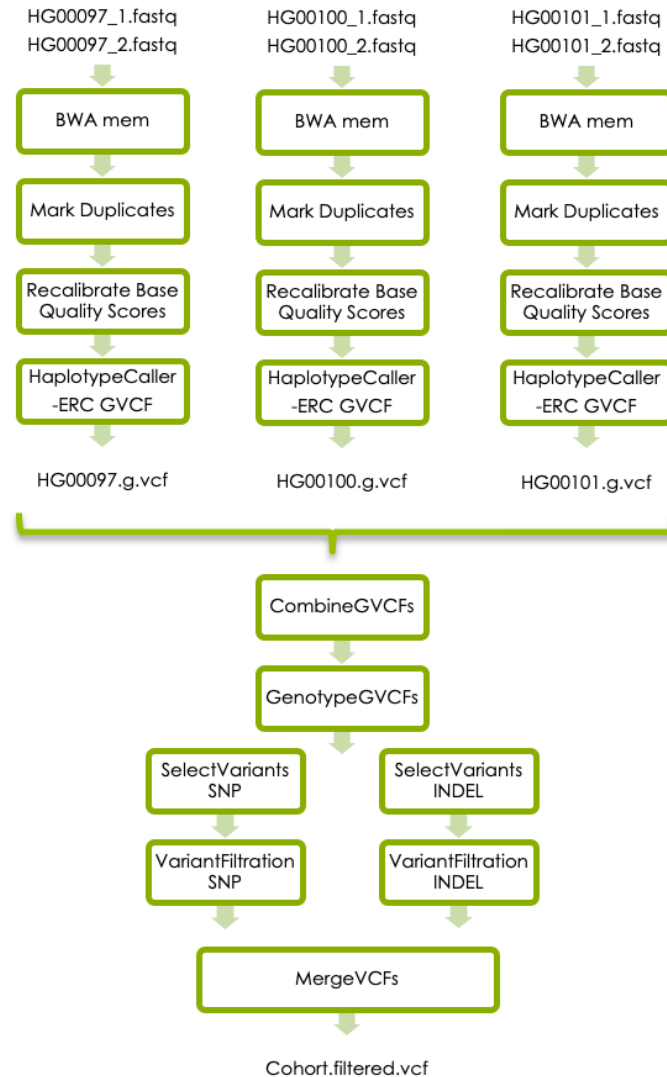


[https://software.broadinstitute.org/gatk/best-practices/
Germline short variant discovery \(SNPs + Indels\)](https://software.broadinstitute.org/gatk/best-practices/Germline-short-variant-discovery-(SNPs-+-Indels))



- Remove low quality variants
- Variant quality score recalibration (VQSR):
 - For large data sets (>1 WGS or >30WES samples)
 - GATK has a machine learning algorithm that can be trained to recognise "likely false" variants
 - **We do recommend to use VQSR when possible!**
- Hard filters:
 - For smaller data sets
 - Hard filters on information in the VCF file
 - For example: Flag variants with "QD < 2" and "MQ < 40.0"
 - GATK discussion on hard filters:
<https://gatkforums.broadinstitute.org/gatk/discussion/2806/howto-apply-hard-filters-to-a-call-set>

GATK's best practices workflow



More details and links to GATK for each step is found in the lab instructions

Today's lab





- Low coverage WGS data
- 3 samples
- Small region on chromosome 2

About the samples:

<https://>

www.internationalgenome.org/data-portal/sample

The Lactase enzyme

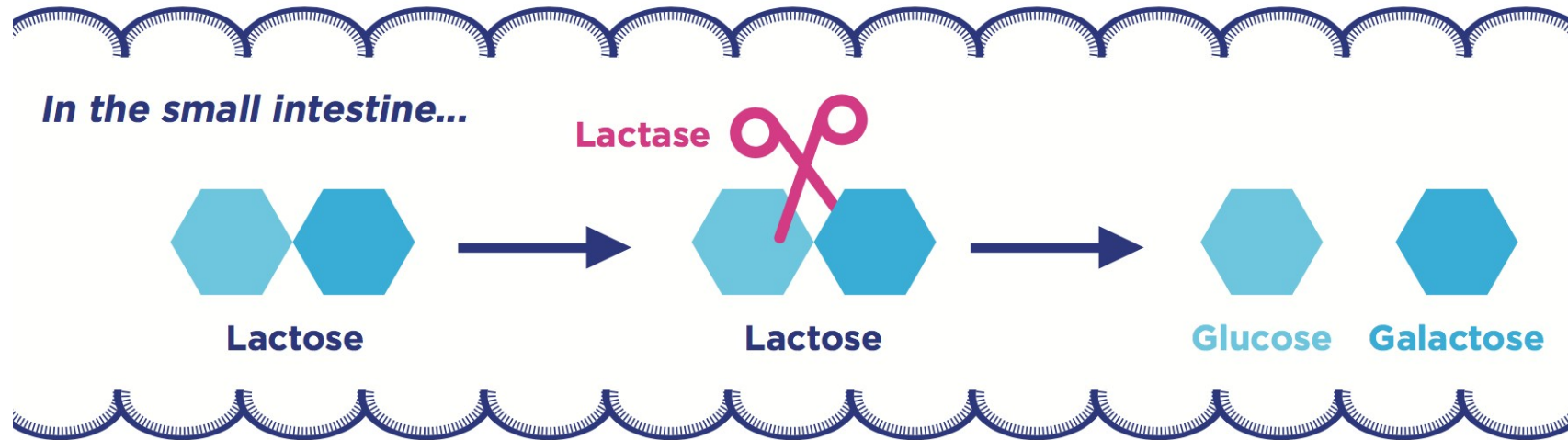


Figure 2. Lactose digestion in the intestine.

- All mammals produce lactase as infants
- Some human produce lactase in adulthood
- Genetic variation upstream of the *LCT* gene cause the lactase persistent phenotype (lactose tolerance)

part one:

variant calling in one sample



Basic variant calling in one sample

HG00097_1.fa
stq
HG00097_2.fa
stq

FASTQ files

BWA mem

HG00097.ba
m

BAM files

HaplotypeCall
er

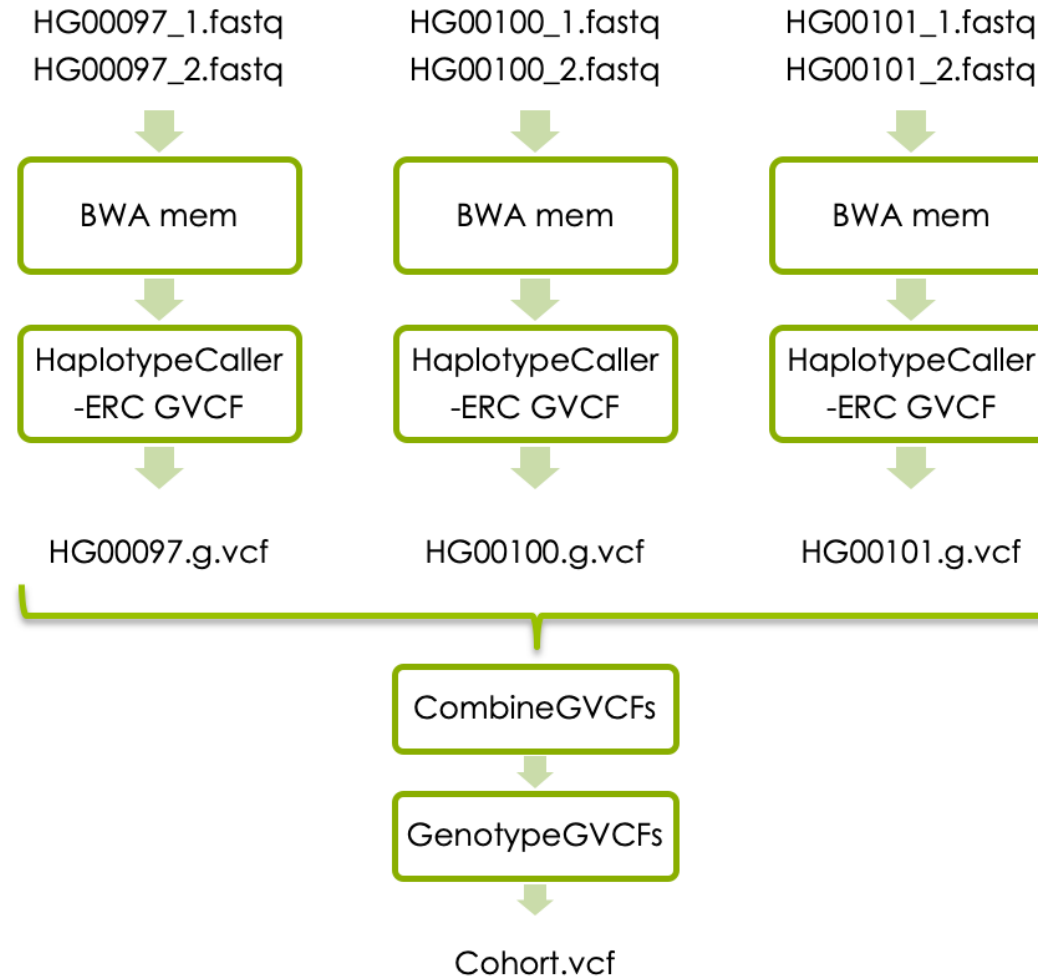
HG00097.vcf

VCF files



Part two (if you have time):
variant calling in cohort

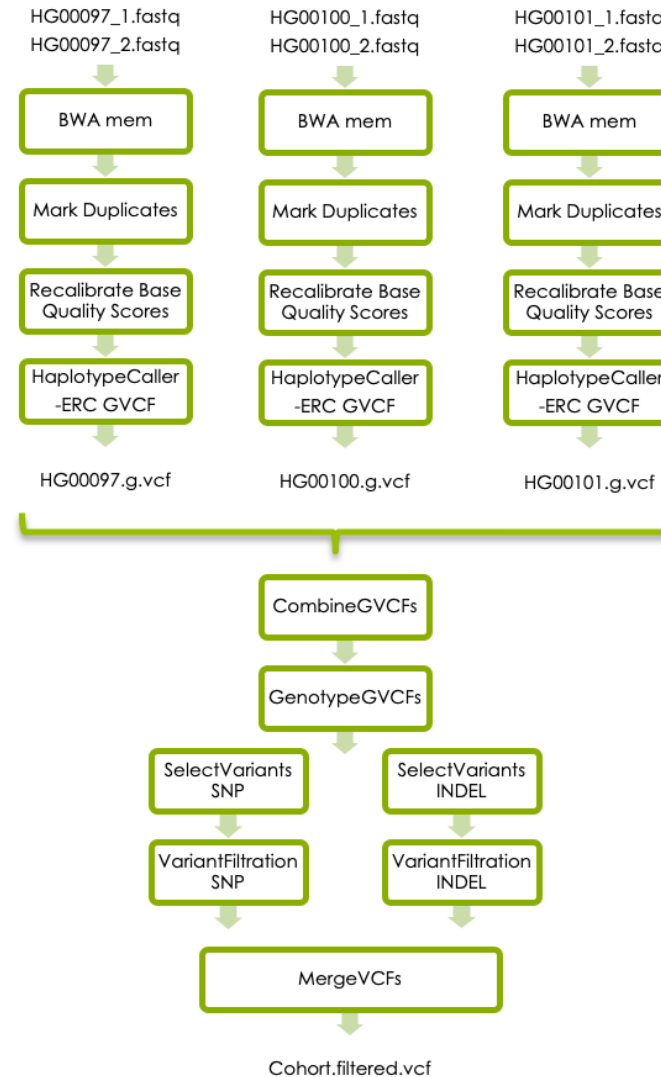
Joint variant calling workflow



Part three (if you have time):

**Follow GATK best practices for short
variant discovery**

GATK's best practises



First look at video about this linked from schedule!



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Questions?