



# **Quality Control of NGS data**

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## FastQ files



```
@HWUSI-EAS100R:6:73:941:1973#0/1
GATTTGGGGTTCAAAGCAGTATCGATCAAATAGTAAATCCATTTGTTCAACTCACAGTTT
+
!''*((((***+))%%%++)(%%%%).1***-+*''))**55CCF>>>>>CCCCCCC65
```

1st row: sequence identifier (machine ID, x-y coordinates, additional info)

2<sup>nd</sup> row: The actual sequence

3rd row: starts with "+" and optionally the same identifier as in the 1st row

4th row: Quality score for each base in read



# **Phred Quality Scores**



```
+SEQ_ID
!''*((((***+))%%%++)(%%%%).1**
```

A quality value Q is an integer representation of the probability p that the corresponding base call is incorrect.

$$Q = -10 \log_{10} P$$
  $\longrightarrow$   $P = 10^{\frac{-Q}{10}}$ 

Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90%
20	1 in 100	99%
30	1 in 1000	99.9%
40	1 in 10000	99.99%
50	1 in 100000	99.999%



# **FastQC**



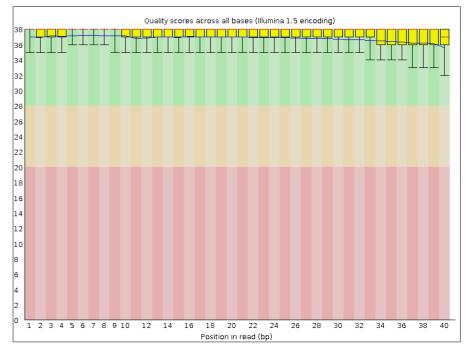


### **FastQC**

## Bad qualities:

# Quality scores across all bases (illumina 1.5 encoding) 34 32 30 28 26 24 22 20 18 16 14 12 10 8 6 4 2 0 1 2 3 4 5 6 7 8 9 10 12 14 16 18 20 22 24 26 28 30 32 34 36 38 40 Position in read (bp)

## Good qualities:





## What is QC?

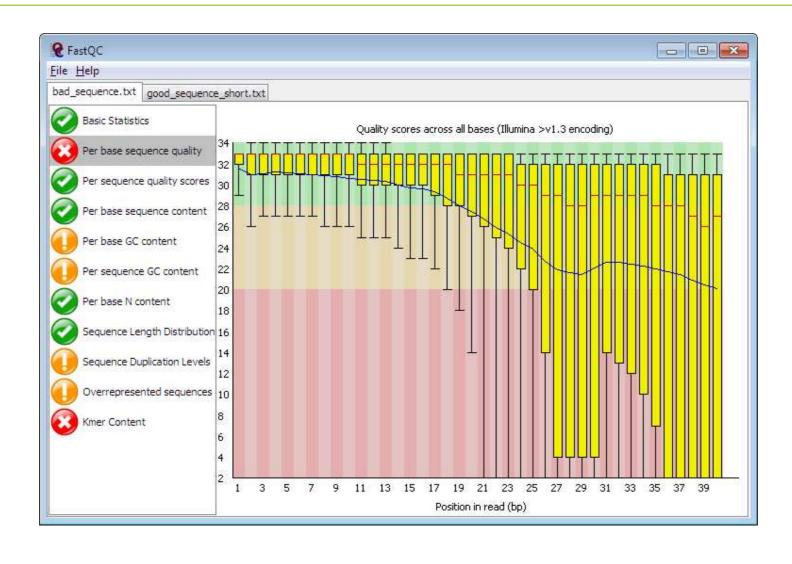


- Different NGS application have their own problem areas and requires their own QC strategy
- Today: Focus on QC for whole genome sequencing
- For variant calling it is important to look at quality score distribution, sequence length distribution and duplication levels.
- Thursday: More details on QC for RNA-seq



# **FastQC**







## FastQC link



https://www.bioinformatics.babraham.ac.uk/projects/fastqc/